RESEARCH LETTER

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Pneumocystis pneumonia risk among viral acute respiratory distress syndrome related or not to COVID 19



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Lymphopenia, corticosteroids and immunomodulatory therapeutics frequently used in COVID-19 patients with acute respiratory distress syndrome (C-ARDS) may be contributing factors to opportunistic infection such as *Pneumocystis jirovecii* pneumonia (PCP).

We conducted a retrospective study to compare the prevalence of PCP between patients with C-ARDS and those with non-SARS-CoV-2 viral ARDS (NC-ARDS).

Methods and some data from this cohort have been previously published [1]. There was no systematic protocol to search for PCP but in case of suspicion of PCP (respiratory symptoms with any consistent radiographic features), several analyses were performed on respiratory samples, such as broncho-alveolar lavage (BAL), blind protected sample, or sputum. It included direct examination (using May-Grünwald Giemsa (MGG), or immunofluorescence staining), detection of Pneumocystis jirovecii DNA by real-time polymerase chain reaction (qPCR) [2], and serum (1-3)- β -D-glucan. During the COVID-19 outbreak, immunofluorescence staining was not performed. PCP was defined as per the revised EORTC/MSGERC definition [3] as follows: proven in case of suspicion with positive direct examination; possible in case of suspicion with positive qPCR and positive BDG in ≥ 2 consecutive serum samples provided other etiologies have been excluded. SARS-CoV-2 and other viruses were not considered a priori as host factors. Patients with positive qPCR but lacking the other criteria for possible PCP were classified as colonized.

The primary endpoint was the difference in prevalence of PCP between C-ARDS and NC-ARDS patients.

No statistical sample size calculation was performed a priori, and sample size was equal to the number of patients treated during the study period. All patients were included only once.

Between October 1, 2009, and April 29, 2020, ninety patients had C-ARDS (positive RT PCR test for SARS-CoV-2), while 82 patients had viral NC-ARDS. Our study comprises 120 patients with fungal analyses on respiratory samples obtained from 81 C-ARDS and 39 NC-ARDS patients. NC-ARDS patients had more comorbidities were more often immunocompromised, and had lower lymphocyte counts than C-ARDS patients (Table 1). C-ARDS patient received less steroid than NC-ARDS patients because they were included before randomized trials demonstrating decreased mortality with dexamethasone.

Pneumocystis analyses were performed on a mean of 3.1 respiratory sample per patient (range 1–15). Direct examination was performed in a total of 72 samples, with two positive cases. qPCR was performed in a total of 368 samples (294 blind protected samples, 72 BAL, and three sputum). All qPCR were negative in C-ARDS patients, while five (13%) NC-ARDS patients had at least

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 Table 1
 Characteristics of patients with Pneumocystis jirovecii research according to C-ARDS and NC-ARDS patients

	NC-ARDS (n = 39)	C-ARDS (n = 81)	p-value 0.32	
Age, median [IQR]	61.8 [56.1–69.3]	58 [52–69.5]		
Male gender	28 (72%)	65 (80%)	0.30	
Medical history				
Mc Cabe			< 0.0001	
No underlying condition	13 (33%)	70 (86%)		
Ultimately fatal	16 (41%)	10 (12%)		
Rapidly fatal disease	10 (26%)	1 (1%)		
Charlson comorbidity index	3 [2–4]	1 [0-2]	< 0.0001	
Diabetes mellitus	11 (28%)	38 (47%)	0.051	
Congestive heart failure (NYHA 3-4)	3 (8%)	6 (7%)	> 0.99	
Supraventricular arrhythmia	5 (13%)	8 (10%)	0.76	
Hypertension	16 (41%)	52 (64%)	0.016	
COPD	2 (5%)	8 (10%)	0.50	
Chronic renal failure	8 (21%)	13 (16%)	0.55	
Dialysis	3 (8%)	2 (3%)	0.33	
Stroke	1 (3%)	3 (4%)	> 0.99	
Liver cirrhosis (Child C)	1 (3%)	0 (0%)	0.33	
Current smoking	8 (21%)	21 (26%)	0.52	
Immunosuppression conditions				
Overall	32 (82%)	13 (16%)	< 0.0001	
Solid cancer	2 (5%)	4 (5%)	> 0.99	
Blood cancer	15 (38%)	0 (0%)	< 0.0001	
Organ transplant	9 (23%)	5 (6%)	0.013	
HIV infection	3 (8%)	3 (4%)	0.39	
Sickle cell disease	1 (3%)	3 (4%)	> 0.99	
Others	4 (10%)	1 (1%)	0.038	
Clinical characteristics upon ICU admission				
IGS2	51 [37–68]	35 [27–43]	< 0.0001	
Baseline SOFA, median [IQR]	9 [6–12]	7 [4–8]	< 0.0001	
ARDS classification (Berlin definition)			0.046	
Mild	12 (31%)	10 (12%)		
Moderate	18 (46%)	44 (54%)		
Severe	9 (23%)	27 (33%)		
Norepinephrine, n (%)	20 (51.3%)	35 (43.2%)	0.41	
Serum creatinine (µmol/L)	147 [83–226]	82 [66–124]	0.001	
White blood cell count (\times 10 ⁹ /L)	5.4 [3–14.8]	8.1 [5.5–11.9]	0.44	
Lymphocyte count ($\times 10^9/L$)	0.4 [0.2–0.9]	0.8 [0.5–1.2]	0.01	
Documented bacterial co-infections	18 (46%)	13 (16%)	< 0.0001	
Treatments during the first 24 h	, ,	, ,		
Antibiotics	39 (100%)	81 (100%)	> 0.99	
Antiviral treatment	26 (67%)	65 (80%)	0.10	
Corticosteroids (any dose)	21/38 (55%)	10/79 (13%)*	< 0.0001	
Corticosteroids (low dose)	20/38 (53%)	8/79 (10%)*	< 0.0001	
Corticosteroids (high dose) #	1/38 (3%)	2/79 (3%)*	> 0.99	
ARDS treatment during ICU stay	(,	\/	. 0.22	
Corticosteroids (any dose)	24 (63%)	32 (41%)*	0.02	
Corticosteroids (low dose)	22 (58%)	22 (28%)*	0.002	
Corticosteroids (low dose) #	2 (5%)	10 (13%)*	0.22	
Prone position	20 (51%)	71 (88%)	< 0.0001	

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Table 1 (continued)

	NC-ARDS (n=39)	C-ARDS (n = 81)	p-value	
Neuromuscular blockade	25 (64%)	74 (91%)	< 0.0001	
Inhaled nitric oxide	6 (15%)	28 (35%)	0.03	
Extra-corporeal membrane oxygenation	5 (13%)	20 (25%)	0.13	
Organ support and outcome during ICU stay				
Renal replacement therapy during ICU stay	19 (49%)	29 (36%)	0.18	
Norepinephrine, n (%)	32 (82%)	61 (75%)	0.41	
ICU length of stay among survivors, days	17 [10–28]	30 [22–46]	0.09	
Death at day 28	15 (39%)	30 (37%)	0.88	
Death in the ICU	17 (44%)	32 (40%)	0.67	
Pneumocystis jirovecii samples and analysis				
Total samples, mean (range)	1.5 (1–4)	3.8 (1–15)	< 0.001	
Sputum examination, mean (range)	0.08 (0-1)	0	0.01	
Broncho-alveolar lavage, mean (range)	1.5 (0–4)	0.19 (0-2)	< 0.001	
Blind protected sample, mean (range)	0	3.6 (1–15)	< 0.001	
Direct examination (IF or MGG)	1.5 (0–4)	0.19 (0-2)	< 0.001	
qPCR	1.5 (1–4)	3.8 (1–15)	< 0.001	
Serum (1–3)-BDG	0.5 (0-4)	(0-4) 4 (1-10)		

COPD = chronic obstructive pulmonary disease, HIV = human immunodeficiency virus, SAPS II = Simplified Acute Physiology Score II, SOFA = sequential organ failure assessment, ICU = intensive care unit; *two missing values because two patients received dexamethasone or placebo in a randomized controlled trial; #denotes more than 1 mg/kg of prednisone or equivalent

Table 2 Characteristics of patients with a positive *Pneumocystis jirovecii* qPCR

Patient, age, sex	Underlying disease	Date of PCP diagnosis	Viral association	Respiratory sample	Direct examination (IFI or MGG)	Pneumocystis qPCR	BDG (pg/ ml)*	Time between ICU admission and positive sample (day)	Treatment
P1, 58y, M	Diabetus mellitus Congestive heart failure	14/01/2014	Coronavirus, Rhinovirus	BAL	Negative	36.7	NA	1	No
P2, 73y, M	Renal trans- plantation Diabetus mellitus Congestive heart failure	29/08/2015	Coronavirus	BAL	Positive	32	106	0	Yes (sulfameth- oxazole)
P3, 52y, M	Myasthenia (steroid, aza- thioprine)	04/07/2012	Respiratory syncytial virus	BAL	Negative	39.8	NA	1	No
P4, 32y, F	Acute lymphoblas- tic leukemia (methotrex- ate and aracytabine)	08/01/2019	Metapneu- movirus	BAL	Positive	27.9	188	0	Yes (sulfameth- oxazole)
P5, 67y, M	Cirrhosis, rheumatoid polyarthritis (steroid)	22/04/2019	Coronavirus NL63	BAL	Negative	36.6	NA	1	No

M = male, F = female, P = patient; $BDG = (1-3)-\beta-D-glucan,*BDglucan not performed in the lab before 2013. <math>BDglucan$ was performed using the Fungitell kitTM (Cape Cod Inc, USA) with a positivity threshold of 80 pg/mL; qPCR of $P_ijirovecii$ was performed using a region of the mitochondrial large subunit rRNA gene (LSU) after DNA extraction with a Qiasymphony kit (Qiagen, Courtaboeuf, France)

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one positive PCR, with a median cycle threshold of 36.6 [30–38.3].

Two NC-ARDS patients fulfilled proven PCP diagnostic criteria, with a positive direct examination, a single β-D-glucan > 80 pg/mL (Table 2), and received treatment for PCP.

Three other NC-ARDS patients were classified as colonized, while no patient fulfilled possible PCP diagnostic criteria. Time between ICU admission and positive sample for PCP (Table 2) was short (< 2 days) like in other invasive fungal infections (i.e. invasive pulmonary aspergillosis) in severe influenza infection or ARDS.

In this study of patients with viral ARDS, we found a low risk for possible or proven PCP. Our findings are in accordance with two smaller studies in France [4, 5] retrieving a low risk of *Pneumocystis* colonisation in COVID-19 patients. In our cohort, qPCR was positive in 13% of NC-ARDS. This result is in accordance with a previous study showing 7% of positive qPCR in ICU-admitted influenza patients [6].

The strengths of our study are the analysis of a large ARDS cohort with fungal analyses. Our study also has limitations: monocentric design, NC-ARDS patients more frequently immunocompromised, and a long cohort period.

Abbreviations

C-ARDS: Coronavirus disease 19 related acute respiratory distress syndrome; NC-ARDS: Non-coronavirus disease 19 viral ARDS; PCP: *Pneumocystis jirovecii* Pneumonia.

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Authors' contributions

KR, RA and FB contributed to the study design, analysis and interpretation of data. KR, RA and AMD drafted the initial manuscript and approved the article final version. AFH, SF, AMD and the COVID-PCP group contributed to the interpretation of data, critical revision of intellectual content and approval of the submitted version of the article. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets supporting the conclusions are included within the article.

Declarations

Ethics approval and consent to participate

This observational study was approved by the Ethical Review Board of the French Society for Intensive Care Medicine (*Société de Réanimation de Langue Française*). As per the French law, no informed consent was required for this type of studies.

Consent for publication

Not applicable.

Competing interests

All authors report no conflict of interest relevant to this study.

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