

REVIEW

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Pharmacokinetics–pharmacodynamics issues relevant for the clinical use of beta-lactam antibiotics in critically ill patients

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Abstract

Antimicrobials are among the most important and commonly prescribed drugs in the management of critically ill patients and beta-lactams are the most common antibiotic class used. Critically ill patient's pathophysiological factors lead to altered pharmacokinetics and pharmacodynamics of beta-lactams.

A comprehensive bibliographic search in PubMed database of all English language articles published from January 2000 to December 2017 was performed, allowing the selection of articles addressing the pharmacokinetics or pharmacodynamics of beta-lactam antibiotics in critically ill patients.

In critically ill patients, several factors may increase volume of distribution and enhance renal clearance, inducing high intra- and inter-patient variability in beta-lactam concentration and promoting the risk of antibiotic underdosing. The duration of infusion of beta-lactams has been shown to influence the $fT > \text{minimal inhibitory concentration}$ and an improved beta-lactam pharmacodynamics profile may be obtained by longer exposure with more frequent dosing, extended infusions, or continuous infusions.

The use of extracorporeal support techniques in the critically ill may further contribute to this problem and we recommend not reducing standard antibiotic dosage since no drug accumulation was found in the available literature and to maintain continuous or prolonged infusion, especially for the treatment of infections caused by multidrug-resistant bacteria.

Prediction of outcome based on concentrations in plasma results in overestimation of antimicrobial activity at the site of infection, namely in cerebrospinal fluid and the lung. Therefore, although no studies have assessed clinical outcome, we recommend using higher than standard dosing, preferably with continuous or prolonged infusions, especially when treating less susceptible bacterial strains at these sites, as the pharmacodynamics profile may improve with no apparent increase in toxicity.

A therapeutic drug monitoring-guided approach could be particularly useful in critically ill patients in whom achieving target concentrations is more difficult, such as obese patients, immunocompromised patients, those infected by highly resistant bacterial strains, patients with augmented renal clearance, and those undergoing extracorporeal support techniques.

Keywords: Critical care or intensive care or critically ill, Sepsis or septic shock, Antibiotics, Pharmacokinetics, Pharmacodynamics

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Background

Antimicrobials are among the most important and commonly prescribed drugs in the management of critically ill patients and beta-lactams are the most common antibiotic class used because of their broad spectrum of activity and high tolerability [1, 2].

Early and appropriate antibiotic administration improves clinical outcome of septic patients [3–7]. In the presence of septic shock, besides conflicting results [8, 9], each hour delay is associated with a measurable increase in mortality and other negative endpoints (e.g., length of stay in ICU, acute kidney injury, acute lung injury, and global organ injury assessed by the Sepsis-Related Organ Assessment score) [10, 11].

Choosing the appropriate antimicrobial for the bacterial activity spectrum is crucial but the correct dosage regimen (both dose and frequency) is, at least, of the same importance for successful clinical cure and microbiological eradication [11].

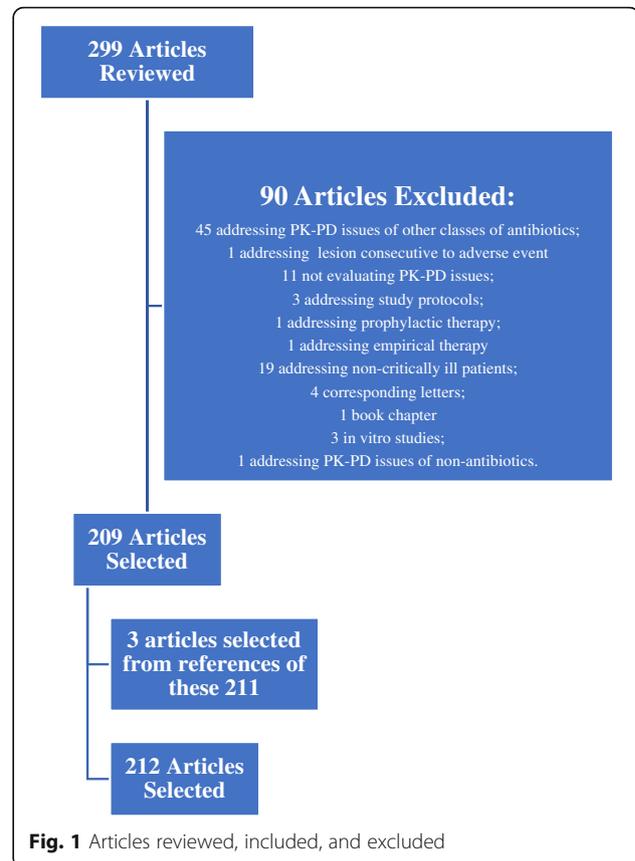
Unlike organotropic drugs, where it is easy to titrate dose to achieve a clinical response, antibiotics may take 24–72 h to present signs of resolution of infection, making it difficult to determine the most appropriate dosage [1, 2].

We conducted a comprehensive bibliographic search in the PubMed database of all English language articles published from January 2000 to December 2017, using the following keywords: critical care or intensive care or critically ill and sepsis or septic shock and antibiotics and pharmacokinetics or pharmacodynamics. Articles not addressing beta-lactam pharmacokinetics (PK) or pharmacodynamics (PD) in critically ill patients were excluded. A small number of articles derived from references in the articles selected were also reviewed. In the end, 214 studies were included in our review (Fig. 1).

Beta-lactam PD characteristics

Knowledge of the antimicrobial PD characteristics (inhibition of growth, rate and extent of bactericidal action, and post-antibiotic effect (PAE)) provides a more rational basis for determination of optimal dosing regimens in terms of the dose and the dosing interval.

The antimicrobial activity of drugs is usually assessed by determination of the minimum inhibitory concentration (MIC) and the minimum bactericidal concentration (MBC) of the drug under specific conditions in vitro. These in vitro conditions are very different from those expected at the site of infection, where the milieu is frequently acidic and anaerobic, and tissue protein may bind a variable amount of the drug. Additionally, these parameters do not provide information on the time course of the antimicrobial effect—the fluctuating levels that are present in a patient treated with the drug—and are measured against a standard bacterial inoculum (about 10 colony-forming units (CFU) per millilitre [5])



that does not necessarily correspond to bacterial densities at site of infection (10 CFU per gram of tissue or pus [8–10]). For bactericidal drugs, the MBC is generally not more than fourfold higher than the MIC. The size of the residual bacterial population at the end of each dosing interval, and ultimately the efficacy of the antimicrobial regimen, depends on the interplay of a variety of bacterial, drug, and host factors that include the size of the initial bacterial population, the potency (MIC and MBC) and PK characteristics of the antimicrobial agent, the rate and extent of any bactericidal effect, the presence of a post-antibiotic effect (PAE), the rate of re-growth of persistent organisms, and the state of host defences [13].

Three PD indices describe optimal killing associated with antibiotics: $fT > MIC$, which is the amount of time that the unbound drug concentration remains above the MIC of the infecting organism; C_{max}/MIC , which is the ratio between the maximum concentration of the drug and the MIC of the infecting organism; and AUC_{0-24}/MIC , which is the ratio between total area under the concentration–time curve (AUC) over 24 h and the MIC of the infecting organism.

Beta-lactams are time-dependent antimicrobials whose efficacy is mainly related to $fT > MIC$ [1, 2, 12–14].

Increasing drug concentrations much above the MBC does not enhance bacterial killing and the bactericidal action of these drugs is relatively slow. When drug levels at the site of infection fall below the MIC, the relatively large residual population can resume growth quickly because most beta-lactams either have no or only a short PAE [12]. McKinnon et al. compared the PD of cefepime and ceftazidime and observed that patients with $fT > MIC$ of 100% had significantly greater rates of clinical cure and bacteriological eradication than patients with $fT > MIC$ of $< 100\%$ [15].

It is suggested that 50% $fT > MIC$ of the dosage interval is needed to ensure standard efficacy with these antimicrobials, whereas 100% $fT > MIC$ of the dosage interval should be ensured for optimal exposure in immunocompromised patients. A further improvement in efficacy is observed when antibiotic concentrations are four to five times greater than MIC [2, 12, 13]. The percentage of time above MIC that correlates with efficacy varies among different beta-lactam groups, being greater for cephalosporins and aztreonam than for penicillins, and greater for penicillins than for carbapenems. Also, variations occur among different bacterial species, being less for staphylococci, for which beta-lactams have a PAE, than for streptococci and Gram-negative bacilli, for which beta-lactams do not have a PAE [2, 13].

Beta-lactam PK issues in the critically ill

Discovered antibiotics are evaluated in vitro and tested in animals, initially for toxicity and subsequently for efficacy. The antibiotic dose and frequency are based on these in vitro or animal in vivo PK/PD studies. These dosing regimens are then tested on healthy human volunteers for tolerability, with clinical efficacy studies undertaken in non-critically ill patients. After the launch of the drug onto the general market, the same dosing regimen is used in critically ill patients; however, this is likely to lead to suboptimal outcomes in the ICU [5], especially with more resistant bacterial strains [16] and in the immunocompromised population [17].

Beta-lactams are hydrophilic drugs and so their volume of distribution (Vd) is low and similar to that of extracellular water. Variations in the extracellular fluid content and/or in renal or liver function may be considered the most relevant and frequent pathophysiological mechanisms possibly affecting drug disposition in critically ill patients. Other factors may contribute to altered antibiotic concentrations: an interesting case-report by Taccone et al. [18] related the case of an obese septic patient with *Pseudomonas aeruginosa* pneumonia treated with meropenem. The PD target ($t > 4 \times MIC > 40\%$ of the dosing interval) was only achievable by dosing 3 g q6h at 3 h extended infusion and was associated with clinical improvement.

Compared with healthy volunteers and non-critically ill patients, in critically ill patients capillary leakage and edema, fluid therapy, pleural effusion, ascites, indwelling post-surgical drainage, and hypoalbuminemia may increase Vd and cause antibiotic dilution in plasma and extracellular fluids. Some pathophysiological factors may also enhance (trauma, burns, the hyperdynamic condition of the early phase of sepsis, the use of hemodynamically active drugs) or reduce (renal failure, muscular wastage, bedridden patients) renal clearance and consequently may alter plasma and extracellular antibiotic concentrations (with implications on time over MIC), induce high intra- and inter-patient variability, and promote the risk of antibiotic underdosing [1, 2, 12, 14, 19–35]. Extracorporeal support techniques also contribute to antibiotic concentration variability [36].

PK studies on ICU septic patients reported, overall, increased Vd with significant daily concentration fluctuations between and within patients [5, 36–41]. Clearance of drugs is also affected and usually related to creatinine clearance [1, 42–44]. A single-center study of 17 ICU patients with ventilator-associated pneumonia (VAP) described the PK profile of ertapenem and concluded that, because of its highly protein-bound profile, hypoalbuminemia resulted in a higher protein-unbound fraction with consequences for drug distribution and elimination [38]. Ulldemolins et al. [39] found the same while studying the PK profile of flucloxacillin. Ramon-Lopez et al. [45] described high PK variations (between and within patients) for meropenem in 12 burn ICU patients that were mostly related to age, body weight, and serum albumin. Carlier et al. [37] investigated the adequacy of piperacilin/tazobactam dosing and its trough variability during an entire 7-day antibiotic course in 11 ICU patients with pneumonia and normal renal function. Six of them failed to achieve the PK/PD target of 100% $fT > MIC$ at least once during the treatment course and considerable antibiotic concentration variability was found within and between patients. The *DALI study*, a large multicenter prospective study evaluated 248 ICU patients treated for infection with beta-lactams and found large variations on beta-lactam blood concentrations. The achievement of the PK/PD targets was highly inconsistent, with one fifth of the patients not achieving their most conservative PK/PD target of 50% $fT > MIC$ and better outcomes were described with higher drug exposure, at least for less severely ill patients [5].

Septic patients with acute renal failure may have suboptimal antibiotic concentrations in the first days of therapy when the recommended dosing adjustment for renal failure is used [46]. Taccone et al. [40] studied the PK profiles of four beta-lactams (ceftazidime, cefepime, piperacilin/tazobactam, and meropenem) over the first 24 h of treatment in 80 septic ICU patients. They

concluded that, besides high intra- and inter-patient PK variability, standard first doses of broad-spectrum β -lactams provided inadequate levels to achieve target serum concentrations for extended periods of time.

Augmented renal clearance has probably more impact than altered Vd on the PK of beta-lactams [25, 27, 47–54]. Roberts et al. [23] described the PK of cefazolin in plasma and interstitial fluid of subcutaneous tissue in post-trauma critically ill patients and demonstrated that increasing creatinine clearance (ClCr) or decreasing serum albumin concentrations will reduce the likelihood of achieving optimal cefazolin exposure in subcutaneous interstitial tissue. In the presence of augmented renal clearance (ClCr > 130 mL/min), a much higher dose of cefazolin is required to obtain similar relative drug exposures [23]. Conil et al. [43] found that higher ClCr values (> 50 mL/min) did not provide trough concentrations of piperacilin (4 g three times a day) sufficient enough to attain the MIC for many pathogens in many of the patients studied.

Hypoalbuminemia has also been associated with altered PK. Wong et al. [55] described a linear correlation between the percentage protein binding of flucloxacillin and the plasma albumin concentration, though this was not true for ceftriaxone. Also, plasma albumin concentrations and in vitro binding data from healthy volunteers should not be used to predict unbound concentrations of ceftriaxone in ICU patients [56].

Use of extracorporeal support techniques in critical care

Acute kidney injury (AKI) occurs in 50 to 65% of critically ill patients and in approximately two-thirds of patients within the first 24 h after admission to the intensive care unit (ICU) [57]. Critically ill patients are usually supported with one of the forms of continuous renal replacement therapy (CRRT)—continuous venous-venous hemofiltration, hemodiafiltration, hemodialysis (CVVHF, CVVHDF, CVVHD, respectively)—or with sustained low-efficiency dialysis (SLED). Molecules are transported across the filter membrane by the mechanism of convection (driven by the pressure gradient—CVVHF), diffusion (driven by the concentration gradient—CVVHD, SLED), or both (CVVHDF).

Employing CRRT complicates antibiotic dosing to a significantly higher extent than standard hemodialysis due to the high number of variables, including Vd, flow of the dialysis fluid, replacement fluid infusion site (pre- or post-dilution mode), type and surface of the used membrane, and the difference between delivered and prescribed RRT dose.

Vd in AKI may be significantly different from published population estimates derived from healthy subjects. Besides the decreased plasma protein concentrations in

acutely ill patients, uremic solutes, such as hippurate and indoxyl sulfate, alter drug binding to albumin in chronic renal failure and might do so in acute renal failure, although this has not been tested. The free fraction of many drugs is increased in renal failure, even though the Vd for total drug may increase due to movement of unbound drug into interstitial or total body water [57–59].

Overall, a tendency for antibiotic underdosing in critically ill patients on CRRT or SLED likely exists. The mode and dose of CRRT vary quite widely from center to center and from report to report, making it very difficult to create generally applicable beta-lactam dosing guidelines for critically ill patients under CRRT. Additionally, antibiotic concentrations may vary depending on the degree of extraction and residual renal function, which is variable, difficult to assess, and rarely considered despite its relevant contribution to antibiotic clearance in patients undergoing CRRT (Tables 1 and 2) [60–96].

Globally, we recommend not reducing standard antibiotic dosage since no drug accumulation was found in the available literature and to maintain continuous or prolonged infusion in critically ill patients on CRRT, SLED, or EDD, especially for the treatment of multidrug-resistant bacteria. Although usually not available in clinical routines, a therapeutic drug monitoring (TDM)-guided strategy has potential benefit to ensure appropriate antibiotic therapeutic targets.

Extracorporeal membrane oxygenation (ECMO) has become an essential tool for severe cardiorespiratory failure in critically ill patients. It is thought to introduce additional confounding factors to the already altered PK properties of beta-lactams in this subset of patients. Sequestration of antibiotics in the ECMO circuit and the associated systemic inflammation can further increase the antibiotic Vd and reduce clearance [74, 97–99]. However, very few in vivo studies have been performed in this subset of patients (Table 3). Globally, they show no significant statistical variation in Vd and clearance, but while probability of target attainment (PTA) with standard ICU dosage regimens was achieved when treating for highly susceptible Gram-negative bacteria, antibiotic concentrations were below those desired to treat more resistant strains.

Longer exposure regimens: continuous infusion, extended infusion, or reduced-interval dosing

The duration of infusion of beta-lactams has been shown to influence their $fT > MIC$. Improved PD profiles of beta-lactams may be obtained by longer exposure with more frequent dosing, extended infusions, or continuous infusions. Several studies reported PD benefits for target attainment of extended and continuous infusions, especially considering highly resistant bacterial strains, even using smaller daily doses [1, 2, 36, 41, 103–143].

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Fish et al. [67]	To more fully characterize PK disposition of imipenem in critically ill adult patients during currently used CWH or CWHDF regimens	Imipenem-cilastatin	Prospective open-label study of imipenem-cilastatin administered as the combination product in a fixed 1:1 ratio Adult ICU patients with CWH (n = 6 patients) or CWHDF (n = 6 patients) Imipenem administered at 0.5 g every 8 to 12 h (total daily doses of 1 to 1.5 g/day) by intravenous infusion over 30 min Pre- and post-membrane plasma and corresponding ultrafiltrate or dialysate samples were collected at 1, 2, 4, and 8 or 12 h (depending on dosing interval) after completion of the drug infusion	Patients on CWHDF had significantly higher CLs compared to patients on CWH (P = 0.01), but this difference was not significant when normalized for total body weight (P = 0.477) The observed t1/2 was overall similar between both patient groups (P = 0.860) No significant differences were found in Vd, S, and Sa, or ultrafiltration rates Mean Cmax, Cmin, and AUC0–24 values in patients receiving 0.5 g bid during CWH versus CWHDF were 17.5 g/mL vs 14.1 g/mL, 1.1 g/mL vs 1.0 g/mL, and 129.5 gh/mL vs 110.3 gh/mL, respectively, and in patients receiving 0.5 g tid they were 18.5 g/mL vs 17.1 g/mL, 1.9 g/mL vs 1.1 g/mL, and 183.3 gh/mL vs 140.6 gh/mL, respectively Doses of 0.5 g bid and 0.5 g tid generally provided T > MIC of at least 40 to 50% and 50 to 60%, respectively (MICs of < 2 g/mL) Doses of 0.5 g every 6 h (2.0 g/day) were modeled based on the PK parameters observed in this study and the T > MIC calculated: at 2.0 g/day, organisms with a MIC of < 4 g/mL had a T > MIC of at least 50 to 80%. And for those with MIC < 8 g/mL, T > MIC ranged from 34 to 62%	CWH and CWHDF contribute to imipenem clearance to a greater degree than previously reported. Imipenem doses of 1.0 g/day appear to achieve concentrations adequate to treat most common Gram-negative pathogens (MIC up to 2 g/mL) but doses of 2.0 g/day or more may be required to adequately treat and prevent resistance in pathogens with higher MICs (MIC > 4 to 8 g/mL)
Malone et al. [68]	To more fully characterize the PK disposition of cefepime in critically ill adult ICU patients during CWH or CWHDF	Cefepime	Prospective, observational study 12 adult ICU patients receiving CRRT for severe renal failure (5 patients on CWH, 7 patients on CWHDF) Cefepime regimens included either 1 g or 2 g doses administered intravenously every 12 or 24 h (total daily doses of 1 to 4 g/day) Sampling was performed as soon as possible after initiation of the CRRT and drug therapy. Pre- and post-membrane venous blood samples were obtained 1, 2, 4, and 8 h after the completion of the drug infusion and just before administration of the next dose	Drug clearance during CRRT (CL CRRT) and %CLS were significantly higher (P = 0.002 and 0.018, respectively), and t1/2 was significantly lower (P = 0.005) among patients receiving CWHDF than in patients receiving CWH. The mean cefepime S during CWH and Sa during CWHDF were estimated at 0.86 ± 0.04 and 0.78 ± 0.10, respectively, indicating that cefepime is extensively cleared across the CRRT membrane Approximately 40 and 59% of cefepime CLs was attributed to membrane clearance during CWH and CWHDF, respectively, indicating that the clearance of cefepime was	It appears that CWHDF is more efficient than CWH in eliminating cefepime. However, the present study included too few subjects and too much variability was observed within the data to demonstrate this conclusively. Cefepime regimens of 0.25 to 1.0 g/day as recommended by the manufacturer for anuric patients or those receiving conventional hemodialysis would likely be subtherapeutic against all but the most highly susceptible pathogens when administered to patients receiving CRRT The authors recommend cefepime doses of 2 g daily under most

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (Continued)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Mueller et al. [69]	To determine the PK of piperacillin-tazobactam in critically ill patients with acute anuric renal failure treated by CWH	Piperacillin-tazobactam	Prospective, observational study 8 adult ICU patients on CWH Doses and dosing schedules were chosen empirically by the attending physicians and were administered intravenously over 15 min Pre-dialyzer blood samples and dialyzer-outlet dialysate samples were taken before drug administration, at 10 and 30 min after infusion, and at 1, 2, 4, 6, 8, 12, 20, 22, and 24 h after infusion	substantially enhanced during both CRRT techniques Values for cefepime Vd were also statistically different ($P = 0.03$) between CWH and CWHDF groups, but changes in Vd were only weakly correlated with changes in t1/2 (Spearman's rank correlation coefficient of 0.641; $P = 0.133$) All pathogens isolated from study patients had cefepime MICs of < 4 g/mL, and doses as low as 1 g/day would predictably provide adequate treatment during either CWH or CWHDF Cefepime doses of 2 g/day would be expected to achieve favorable concentrations in serum against susceptible pathogens (MIC < 8 g/mL) with T > MIC greater than 50%. Cefepime (2 g/day) during either CWH or CWHDF would also be predicted to achieve favorable T > MIC > 80% against pathogens with intermediate susceptibility (MIC < 16 g/mL) CWHDF clearance of piperacillin was 37% (median, with a range of 13 to 100%) and the CWHDF clearance of tazobactam was 38% (median, with a range of 32 to 92%) of CLtotal Vd was 0.31 ± 0.07 and 0.24 ± 0.09 for piperacillin and tazobactam, respectively t1/2 was 4.3 ± 1.2 and 5.6 ± 1.3 for piperacillin and tazobactam, respectively Simulations of 4 g of piperacillin and 0.5 g of tazobactam administered every 12 h and 2 g of piperacillin and 0.25 g of tazobactam administered every 8 h resulted in times above MIC of > 50% for piperacillin with susceptible (MIC of piperacillin 16 mg/liter; time above MIC, 48 to 100%) and intermediate susceptible (MIC 32 mg/liter; time above MIC, 17 to 100%) pathogens in seven of eight patients	circumstances in critically ill patients receiving CRRT. However, considering more resistant strains, doses of 4 g/day should be considered for empirical therapy in life-threatening nosocomial infections, while awaiting results of culture and susceptibility testing Patients with residual renal function and patients that receive CRRT with higher dialysate flow rates or higher additional hemofiltrate flow rates might have higher CL of piperacillin-tazobactam, resulting in higher dosage needs

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (*Continued*)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Roberts et al. [70]	To assess the variability of antibiotic trough concentrations, the influence of effluent flow rates on such concentrations, and the incidence of suboptimal antibiotic dosage	Meropenem and piperacillin-tazobactam	Prospective, observational, multicenter study, conducted within the multicenter RENAL study. It assessed the effect of post-dilutional higher intensity (40 mL/kg/h effluent rate) or lower intensity (25 mL/kg/h effluent rate) CRRT. Patients were randomized to receive either higher or lower intensity effluent flow rate 24 adult ICU patients with AKI on CWH. 17 patients were on meropenem and 7 patients on piperacillin-tazobactam	There was marked variability in trough concentrations for all antibiotics: 6.7-fold for meropenem; 3.8-fold for piperacillin; 10.5-fold for tazobactam When investigating trough concentration variability according to effluent flow rate, no statistically significant differences were found using univariate analysis Meropenem trough concentrations were 10.1 ± 8.7 and 15.0 ± 6.4 for low and high effluent flow rates, respectively Piperacillin trough concentrations were 83.6 ± 63.8 and 204.0 ± 105.0 for low and high effluent flow rates, respectively 100% T > MIC was achieved with meropenem (for MIC 2 mg/L) and piperacillin-tazobactam (for MIC 16 mg/L) 76% T > MIC was achieved for meropenem (for MIC 8 mg/L) 86% T > MIC was achieved for piperacillin-tazobactam (for MIC 64 mg/L)	It appears that CRRT effluent flow rates cannot be used independently to guide dose adjustment Trough concentrations failed to achieve the higher therapeutic target in 24% of patients receiving meropenem and 14% of patients receiving piperacillin, which is of concern Dose-adjusting to achieve a target concentration that exceeds the appropriate MIC but is less than potentially toxic concentrations seems desirable but cannot be reliably achieved with empirical dosing
Banyai et al. [71]	To study the PK of ceftiprome in critically ill patients with acute kidney failure treated by CVWH and to develop an optimal dosing regimen in patients with CWH	Ceftiprome	Prospective, observational study 8 adult ICU anuric patients with acute kidney failure on CVWH All patients received a dosage of 2 g ceftiprome over a period of 30 min, every 8 h after starting the hemofiltration Blood samples were collected from the arterial and venous line of the extracorporeal circuit immediately at baseline and at 60, 120, 180, 240, 300, 360, and 420 min after the start of the first infusion. Additional blood samples were collected immediately before the end and 30 min after the end of each infusion, up to a total study period of 48 h Ultrafiltration samples were collected from the outlet of the ultrafiltrate compartment of the hemofilter at corresponding times	C_{max} 14.8 ± 3.2 µg/mL (10.8 to 19.7) C_{min} 3.1 ± 0.8 µg/mL Post hemofiltration to pre-hemofiltration ratio of 0.23 ± 0.10 Total drug removal of 78.0% ± 8.8% Elimination t _{1/2} was 2.36 ± 0.59 h (1.6 to 3.2 h). The calculated Vd was 118 ± 36 L Total body clearance and hemofiltration clearance were 589.1 ± 164.5 mL/min and 43.3 ± 7.8 mL/min, respectively The calculated AUC was 60.4 ± 16.0 mg/L·h	Highest levels of ceftiprome were significantly lower compared with values observed in healthy volunteers and in patients on hemodialysis Plasma ceftiprome concentrations remained above 4 µg/mL for 62% and above 8 µg/mL for 25% of the dosing interval, respectively In patients infected with an intermediate susceptible <i>Pseudomonas aeruginosa</i> (MIC90 ~ 8 mg/L), no sufficient antimicrobial efficacy can be expected Comparable low trough levels of 3.1 ± 0.8 µg/mL suggest a dosage recommendation of 2 g ceftiprome every 8 h

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (Continued)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Eyler et al. [72]	To determine the PK of ertapenem in critically ill adults receiving CWHD or CWHDF	Ertapenem	Prospective, open-label, first-dose PK study 8 adult ICU patients with suspected or confirmed Gram-negative infections receiving CWHD (4 patients) or CWHDF (4 patients) 1 g ertapenem was administered as a half-hour intravenous infusion Blood samples were collected from the CWHD circuit at the sampling port, just before the hemodialysis filter at time zero (baseline), 30 min (end of infusion), and 1, 2, 4, 8, 12, 18, and 24 h after the start of the ertapenem infusion. At the same time points, effluent was also collected from the effluent port of the CWHD/F circuit	CLS, unbound 48 mL/min VC, unbound 32 l VP, unbound 21 l Cl _{dial} , unbound 36 mL/min PTA (40%T > MIC) and fraction of the dosage interval spent above the MIC for different regimens were For MIC of 0.5 mg/L 500 mg q12h, 1.0 and 0.999 500 mg q24h, 1.0 and 0.999 750 mg q24h, 1.0 and 0.999 1000 mg q24h, 1.0 and 0.999 For MIC of 1 mg/L 500 mg q12h, 1.0 and 0.999 500 mg q24h, 0.99 and 0.916 750 mg q24h, 1.0 and 0.999 1000 mg q24h, 1.0 and 0.999 For MIC of 2 mg/L 500 mg q12h, 1.0 and 0.992 500 mg q24h, 0.962 and 0.563 750 mg q24h, 0.999 and 0.750 1000 mg q24h, 0.999 and 0.917	The unbound fraction (20 to 40%) was markedly increased compared to those reported for healthy volunteers (5 to 15%) At the effluent rates, ertapenem was cleared to a substantial degree During the study sampling period, the dose of 1 g every 24 h produced unbound ertapenem concentrations that remained above 2µg/mL for an average of 90% of the dosing interval, achieving the PD targets in all eight patients Monte Carlo simulations revealed that 99.9% of simulated subjects would achieve unbound ertapenem concentrations above 2µg/mL for at least 40% of the interval, with concentrations remaining above 2µg/mL for a median of 92% (range, 33 to 100%) of the dosing interval Particularly in patients where organisms with high MICs are suspected, it may be necessary to use doses > 500 mg q24h. Concentrations remained above 2µg/mL for an interquartile range of only 50 to 67% of the dosing interval
Vossen et al. [73]	To characterize the PK profile of 1000 mg doripenem q8h for critically ill patients receiving CRRT	Doripenem	Prospective, open-label, observational study 13 adult ICU patients under CRRT (5 on CWH, 5 on CWHD, 3 on CWHDF) All patients received 1000 mg every 8 h at a 30-min infusion Blood and dialysate samples were drawn from the arterial (input), venous (output), and effluent dialysate ports of the dialysis machine before the first administration of doripenem and at 0.5, 1, 2, 3.5, 7, 8, 9, 16, 17, 24, 24.5, 25, 26, 27.5, 31, 32, 33, 40, 41, and 48 h following the start of the first infusion	All patients: AUC ₀₋₈ (mg·h/L) 78.58 ± 10.32; Cl _{tot} (L/h) 8.07 ± 1.77; C _{ipre} -post filter (mL/min) 36.06 ± 14.27; Sc 0.150 ± 0.053; Cl _{Sc} (mL/min) 5.20 ± 1.95; Vd total 59.26 ± 26.47; t _{1/2} (h) 5.39 ± 2.84 Patients on CWH (n = 3): AUC ₀₋₈ (mg·h/L) 87.15 ± 8.13; Cl _{tot} (L/h) 6.53 ± 1.00; C _{ipre} -post filter (mL/min) 63.82 ± 5.83; Sc 0.129 ± 0.033; Cl _{Sc} (mL/min) 6.45 ± 1.81; Vd total 51.05 ± 10.18; t _{1/2} (h) 5.72 ± 1.96 Patients on CWHD (n = 5): AUC ₀₋₈ (mg·h/L) 77.59 ± 12.31; Cl _{tot} (L/h) 7.48 ± 1.43; C _{ipre} -post filter (mL/min) 34.03 ± 7.22; Sc 0.164 ± 0.067; Cl _{Sc} (mL/min) 5.63 ± 2.30; Vd total 71.01 ± 34.98; t _{1/2} (h) 6.80 ± 3.37 Patients on CWHDF (n = 5): AUC ₀₋₈ (mg·h/L) 76.15 ± 4.50; Cl _{tot} (L/h) 9.29 ± 1.28; C _{ipre} -post filter (mL/min) 26.99 ± 5.34; Sc 0.145 ± 0.035; Cl _{Sc} (mL/min) 4.68 ± 1.43; Vd total 50.80	The mean hemofilter clearance rates observed slightly exceeded those reported in the literature The sieving coefficients observed differed dramatically from those reported previously The prefilter/postfilter clearance values found in our trial are within the range of values reported previously for imipenem and meropenem There was an uncharacteristically low clearance for CWHDF patients, which may be attributed to the larger membrane size and higher membrane K _{UF} employed for the CWHF and CWHD groups. The usual expectation for beta-lactam antimicrobials would be quite the opposite: CWHDF clearance values should be higher than CWHD or CWH clearance values. However, this is true only if the same flow rates

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (Continued)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Carlier et al. [74]	To describe the PK of cefepime in septic shock patients requiring CRRT and to investigate whether PK/PD targets are achieved with current dosing strategies as well as to investigate the potential advantages of alternative dosing regimens	Cefepime	Prospective, observational study 13 adult ICU patients with septic shock and on CRRT (CWHF or CWHDF) Patients received 2 g q8h or q12h. The dose was administered as a 30-min intravenous infusion Blood samples were drawn on the day of inclusion and then every second day during CRRT treatment whenever possible. On each sampling day, blood samples were drawn immediately before antibiotic administration (0 h) and then at 1, 2, and 5 h and at 6 or 12 h (depending on the antibiotic regimen) after the start of the infusion	<p>\pm 6.34; t1/2 (h) 3.87 \pm 0.80</p> <p>The simulations conducted show that the proposed dose level of 1000 mg every 8 h is superior to lower doses for reaching the desired plasma doripenem concentration of 8 mg/L. Although mean trough concentrations in all dosing regimens exceeded 4 mg/L during steady state, the individual outcomes were highly variable</p> <p>At a dose of 500 mg every 8 h, only 39.5% of the simulated patients showed trough concentrations that were constantly above the lower threshold (4 mg/L) during steady state. At a dose of 1000 mg every 8 h, plasma concentrations still fell below 8 mg/L, but not lower than 4 mg/L, for 60.5% of the simulated patients</p> <p>CL (L/h) 4.5 Vd (L) 40.6 PTA at different UFR considering a MIC of 16 mg/L UFR 1000 mL/h 1 g q12h 100%T > MIC 64% and 60%T > MIC 95% 2 g q12h 100%T > MIC 89% and 60%T > MIC 99% 1 g q8h 100%T > MIC 95% and 60%T > MIC 100% 2 g q8h 100%T > MIC 99% and 60%T > MIC 100% 1 g q6h 100%T > MIC 100% and 60%T > MIC 100% UFR 1500 mL/h 1 g q12h 100%T > MIC 31% and 60%T > MIC 80% 2 g q12h 100%T > MIC 82% and 60%T > MIC 98% 1 g q8h 100%T > MIC 87% and 60%T > MIC 99% 2 g q8h 100%T > MIC 96% and 60%T > MIC 100% 1 g q6h 100%T > MIC 97% and 60%T > MIC 100% UFR 2000 mL/h 1 g q12h 100%T > MIC 9% and</p>	<p>and membrane materials are chosen. If the 1-g q8h regimen is used, 39.5% of the patients will reach a trough level of 8 mg/L at the end of each dosing interval. To attain sufficient drug exposure during the first dosing interval the administration of an initial "loading dose" of 20.4 mg/kg of body weight is suggested</p> <p>The broad therapeutic index of beta-lactams favors higher dosing, providing safety margin for more-effective RRT modalities</p> <p>Antibiotic clearance was proportional to UFR, with important variability between patients both for clearance and Vd</p> <p>A dose of 2 g q8h or 1 g q6h leads to optimal target attainment (100% T > MIC) whilst minimizing the probability of reaching toxic trough concentrations for patients treated with a high UFR (1500–2000 mL/min). However, the optimal dose for patients treated with lower UFRs (\leq 1000 mL/h) when aiming for the high target was 1 g q8h</p>

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (Continued)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Seyler et al. [75]	To evaluate whether the recommended doses of broad-spectrum beta-lactams result in appropriate serum concentrations in ICU patients with severe sepsis and septic shock receiving CRRT	Meropenem Piperacilin-tazobactam Cefepime Ceftazidime	Prospective, open-label study 53 adult ICU patients with severe sepsis or septic shock on CRRT (CVH, n = 19 or CWHDF, n = 34) Meropenem 1 g q12h (n = 17) Piperacilin-tazobactam 4.5 g q6h (n = 16) Cefepime 2 g q12h (n = 8) Ceftazidime 2 g q12h (n = 12) Each antibiotic dose was administered as a 30-min infusion Serum concentrations of these antibiotics were determined from samples taken before (t = 0) and 1, 2, 5, and 6 or 12 h (depending on the beta-lactam regimen) after the administration of each antibiotic Series of measurements were separated into those taken during the early phase (< 48 h from the first dose) of therapy and those taken later (> 48 h)	60%T > MIC 51% 2 g q12h 100%T > MIC 73% and 60%T > MIC 95% 1 g q8h 100%T > MIC 79% and 60%T > MIC 95% 2 g q8h 100%T > MIC 92% and 60%T > MIC 100% 1 g q6h 100%T > MIC 93% and 60%T > MIC 100%	The recommended doses for broad-spectrum beta-lactams are generally insufficient to maintain therapeutic serum concentrations greater than four times the MIC of <i>P. aeruginosa</i> Applying results to other MICs, the observed concentrations for all antibiotics were adequate in 90% of patients only for MICs lower than the clinical breakpoint of <i>Pseudomonas</i> spp., which correspond to MICs of sensitive <i>Enterobacteriaceae</i> In the first 48 h of treatment, 29%, 34%, 100%, and 62% of our patients treated with meropenem, piperacilin-tazobactam, cefepime, and ceftazidime, respectively, never reached the PK target. After 48 h of treatment, the drug concentrations obtained were higher (significantly different only for meropenem), but they remained insufficient in many patients At the onset of sepsis in patients receiving CRRT, similar beta-lactam doses to those used in the absence of renal failure should be given during the first 48 h of therapy Dose reduction should be considered thereafter to avoid drug accumulation. Considering the large PK variability, therapeutic drug monitoring of beta-lactams should be performed to optimize antibiotic efficacy

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (*Continued*)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Roberts et al. [76]	To evaluate variability in CL and Vd and to assess the effect of CRRT prescription on extracorporeal and systemic antibiotic CL and Vd in patients treated with CRRT of different intensities	Meropenem Piperacillin-tazobactam	Nested cohort prospective multicenter observational PK study within a randomized controlled trial of CRRT intensity Patients were randomly assigned to receive post-dilutional hemodiafiltration as either a higher (40 mL/kg body weight/h effluent flow rate) or lower (25 mL/kg body weight/h effluent flow rate) intensity rate Sampling occurred each day (1) immediately before antibiotic dosing, (2) after completion of their intravenous infusion, and (3) at 4 h after completion of infusion It occurred at 65 time points for meropenem and 29 time points for piperacillin-tazobactam in 24 patients	Ceftazidime Day < 48 h (n = 8) 38% Day > 48 h (n = 7) 71% Mean hemodiafiltration clearance of meropenem, piperacillin, and tazobactam did not differ significantly between higher vs lower CRRT intensity: 23 (16–29) vs 21 (15–28), P = 0.4802; 22 (21–31) vs 24 (17–31), P = 0.9091; 37 (34–49) vs 56 (41–66), P = 0.0642, respectively Systemic clearance and Vd were Meropenem 38 mL/min (23–95) and 17.5 L Piperacillin 59 mL/min (37–115) and 18.7 L Tazobactam 113 mL/min (45–248) and 49.3 L	The prescribed intensity of CRRT did not adequately predict extracorporeal clearance or Sd, CLs, Vd, or half-life Systemic CL and elimination half-life did not differ according to CRRT dose, and so the CRRT prescription may not be useful for guiding antibiotic prescribing. In many cases, extracorporeal CL accounted for more than 30% of the observed systemic CL for that antibiotic, which is a suggested threshold for adjustment of the dosing regimen Drug monitoring may be the most practical method for ensuring that antibiotic therapeutic targets are achieved in critically ill patients receiving CRRT
Ohchi et al. [77]	To investigate PK characteristics of doripenem in patients receiving high-flow vs conventional flow intensity CWHDF	Doripenem	Prospective, observational study Adult ICU Two patients with AKI on high-flow CWHDF. Patients on conventional CWHDF were described in a previous study Doripenem administered as 250 mg single dose infusion over 1 h Blood samples were collected at 1 (just prior to the end of antibiotic infusion), 2, 3, 4, 7, and 12 h after initiating the infusion High flow CWHDF parameters: blood flow 100 mL/min; dialysate flow rate 1500 mL/h; filtration flow rate 900 mL/h	Conventional CWHDF AUC 74.15 ± 15.5 mg·h/L Cis 58 ± 12.7 mL/min CI dialysis 13.5 ± 1.6 mL/min t1/2 7.9 ± 3.7 h High-flow CWHDF AUC 35.2 mg·h/L Cis 118 mL/min CI dialysis 41.9 mL/min t1/2 2.9 h	Extracorporeal clearance increases in proportion with the intensity dialysis rate The daily dose thus must be increased to 1.0–1.5 g, the same dosage used when creatinine clearance is > 50 mL/min
Arzuaga et al. [78]	To study the PK of piperacillin and tazobactam during CRRT in ICU patients with various degrees of renal impairment.	Piperacillin-tazobactam	Prospective, observational study 14 adult ICU patients on CWHDF, grouped according to severity (CLCR 10 mL/min, 10 < CLCR < 50 mL/min, and CLCR > 50 mL/min) Piperacillin 4 g and tazobactam 0.5 g were administered every 6 or 8 h, by 20-min intravenous infusion Prefilter blood and ultrafiltrate samples were collected at 0, 0.3, 0.5, 0.75, 1, 3, 6, and 8 h (in case of administration every 8 h) after the	CLCR < 10 mL/min (n = 4) CLCR: 8.67 ± 2.31 mL/min UF flow: 27.1 ± 7.8 mL/min Sc: PIP 0.42 ± 0.25; TZ 0.76 ± 0.26 Cmax: PIP 365.6 ± 232.3; TZ 384 ± 13.4 mg/L t1/2: PIP 7.8 ± 4.2; TZ 7.9 ± 3.0 h Cis: PIP 50.0 ± 53.0; TZ 50.4 ± 38.3 mL/min Hemofiltration CI: PIP 11.45 ± 6.5 l; TZ 20.9 ± 12.6 mL/min AUC: PIP 761.43 ± 49,748; TZ 23218 ±	The contribution of the hemofiltration clearance to the total clearance increased with the degree of renal insufficiency. Correct doses of these drugs should take into account this observation to avoid clinical failures due to underdosing For both drugs, significant differences were documented in the majority of the PK parameters when patients with CLCR > 50 mL/min were compared to patients with CLCR ≤ 10 mL/min

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (Continued)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
			administration of the antibiotic. Time 0 was considered just before the beginning of the 20-min infusion	<p>27,943 mg/h/L</p> <p>Vd: PIP 21.0 ± 11.7; TZ 18.9 ± 7.1 l</p> <p>10 < CLCR > 50 mL/min (n = 5)</p> <p>CLCR 25.20 ± 7.73 mL/min</p> <p>UF flow: 30.3 ± 4.3 mL/min</p> <p>Sc: PIP 0.38 ± 0.37; TZ 0.73 ± 0.32</p> <p>Cmax: PIP 244.5 ± 122.1; TZ 31.5 ± 5.1 mg/L</p> <p>t1/2: PIP 4.2 ± 2.3; TZ 4.1 ± 0.9 h</p> <p>Cls: PIP 90.6 ± 29.9; TZ 68.2 ± 26.2 mL/min</p> <p>Hemofiltration Ci: PIP 12.2 ± 13.2; TZ 21.9 ± 9.6 mL/min</p> <p>AUC: PIP 4544.5 ± 25,525; TZ 23218 ± 27,943 mg.h/L</p> <p>Vd: PIP 26.8 ± 19.8; TZ 21.6 ± 3.0 l.</p> <p>CLCR > 50 mL/min (n = 5)</p> <p>CLCR: 82.40 ± 20.03 mL/min</p> <p>UF flow: 20.0 ± 7.5 mL/min</p> <p>Sc: PIP 0.23 ± 0.07; TZ 0.86 ± 0.30</p> <p>Cmax: PIP 160.6 ± 93.2; TZ 15.7 ± 6.6 mg/L</p> <p>t1/2: PIP 2.6 ± 0.8</p> <p>TZ 5.0 ± 3.9 h</p> <p>Cls: PIP 265.2 ± 152.2; TZ 180.1 ± 73.9 mL/min</p> <p>Hemofiltration Ci: PIP 4.8 ± 3.3; TZ 19.6 ± 15.3 mL/min</p> <p>AUC: PIP 17,328 ± 11,134; TZ 2098 ± 1030 mg.h/L</p> <p>Vd: PIP 44.9 ± 20.4; TZ 60.3 ± 34.6 l</p> <p>t > MIC90 obtained were 100% for all the pathogens in patients with creatinine clearance < 10 mL/min. In patients with a creatinine clearance between 10 and 50 mL/min, t > MIC90 was 100% for pathogens with MIC90 ≤ 32, but only 50% for microorganisms with an MIC90 of 64. However, in patients with creatinine clearance > 50 mL/min, as piperacillin elimination was faster, t > MIC90 was only 55.5% and 16.6% for pathogens with MIC90 values of 32 and 64, respectively</p>	<p>The observed sieving coefficient of piperacillin and tazobactam plus the effluent amount gave a relevant extracorporeal clearance only in the severe renal impairment group, with more than 25% of total clearance for both drugs</p> <p>To increase the t > MIC90 index, piperacillin-tazobactam combination every 4 h could be a better dosage regimen in patients presenting CLCR > 50 mL/min</p>
Isla et al. [79]	To describe the PK of meropenem in critically ill patients with different degrees of renal impairment undergoing CWHF or CWHD	Meropenem	Prospective, observational study 20 adult ICU patients on CWHF or CWHD Grouped into 3 categories according to the renal function: 7 with severe failure, ClCr less than 10 mL/min	<p>No significant differences depending on renal impairment were found in the Sc. No differences were found in the Sc obtained by CWHF and the Sc obtained by CWHD; both membranes showed a similar</p>	<p>Differences in meropenem PK in critically ill patients undergoing CRRT with different degrees of renal impairment have been observed, and they should be taken into account when dosing critically ill</p>

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (*Continued*)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Uldemolins et al. [80]	To describe the PKs of meropenem in critically ill patients with septic shock and CRRT, to identify the sources of PK variability in these patients, and to perform different dosing simulations to assess their probability of target attainment by MIC, in order to provide empirical dosing recommendations based on clinical characteristics	Meropenem	Prospective, observational, multicentre study 30 adult ICU patients with septic shock and CRRT, either CVWHF (n = 4) or CVHDF (n = 26) Patients were prescribed meropenem at 500 mg q12h over 30 min (n = 1); 500 mg q8h over 30 min (n = 2) or as a 3-h infusion (n = 3); 500 mg q6h as a 3-h infusion (n = 1); 1000 mg q12h over 30 min (n = 6) as a 3-h infusion (n = 1) or as a 4-h infusion (n = 8); 1000 mg q8h over 30 min (n = 8) as a 3-h infusion (n = 5) or as a 4-h-infusion (n = 1); or 2000 mg q8h over 30 min (n = 1) Blood samples were collected at 24 h of CRRT and meropenem	permeability to meropenem Total clearance was significantly higher in group III than in the other two groups. This finding could be attributable to the lower t1/2 (1.51 ± 0.52 h in group III versus 2.73 ± 0.68 h and 3.72 ± 0.82 h in groups II and I, respectively) and to the high Vd observed in those patients (1.31 ± 0.90 L/kg in group III, 0.37 ± 0.10 L/kg in group II, and 0.57 ± 0.29 L/kg in group I) The contribution of CRRT to total clearance diminished in the extent that CLCR increased. Although there were no statistically significant differences between groups I and II, CL _{CRRT} was significantly lower in group III In group I patients, trough plasma concentrations were above 4 µg/mL, with the exception of the only patient who received 500 mg/8 h. In group II, plasma concentrations were above 2 µg/mL during the entire dose interval, except in the patient to whom 1000 mg/8 h was administered In spite of the higher doses the patients of group III received, 4 of 6 patients showed concentrations below 0.5 µg/mL The study model failed to identify CRRT intensity to be a significant modifier of meropenem CL, which may lead to the hypothesis that even the lowest CRRT intensities studied may be enough to maximize meropenem clearance and that higher intensities may add little to total meropenem CL There were no differences between CRRT techniques, likely because of the underrepresentation of CVWHF (4 out of 30 patients) in the study population For the attainment of a PD target of 100% of the T > MIC, fixed doses would be required, depending on the MIC of the bacteria, but the infusion time would depend on	patients In those patients with no renal impairment the risk of underdosing and clinical failure is important, and the administration of meropenem 2000 mg every 8 h did not reach plasma levels to ensure adequate T > MIC values against many bacteria

Table 1 PK/PD studies of beta-lactams in patients undergoing CRRT (Continued)

Study	Endpoints	Antibiotic	Design and type of CRRT	Results	Conclusions
Bouman et al. [81]	To compare the observed $Cl_{C_{WHF}}$ (calculated from measured data) and the predicted $Cl_{C_{WHF}}$ (calculated from the F_{up}) To determine whether dose adjustment according to the predicted CWH removal provides an estimate as reliable as that according to the observed CWH removal	Amoxicillin Ceftazidime Flucloxacillin	therapy. For bolus sampling, 6 samples were collected at 10 min pre-dose; at 0, 15, and 60 min and between 3 and 6 h after the end of the infusion; and just before the next dose. For extended infusion sampling, samples were collected at 10 min pre-dose; 0, 60, and 120 min after the end of the infusion; and just before the next dose CRRT settings: the median intensity on the day of the study was 34.7 mL/kg/h (range, 18.7 to 60.1 mL/kg/h), and the median blood flow was 200 mL/min (range, 130 to 250 mL/min) Prospective, observational study 45 adult ICU oligoanuric patients on CWHF During the sampling period a single antimicrobial drug was administered to 31 patients, two drugs to 9 patients, and a combination of 3 to 5 Dosages were: amoxicillin 1000 mg q6h; ceftazidime 1000 mg q6-12 h; flucloxacillin 2000 mg q4-6 h Blood flow rate was 150 mL/min and warmed substitution fluids were administered in predilution at a flow rate of 2000 mL/h. If a negative fluid balance was required, the ultrafiltration flow was increased and the substitution flow was constant Blood samples were collected from the afferent (pre-hemofilter) and efferent (post-hemofilter) line of the extracorporeal circuit and from the ultrafiltrate line. Samples were collected at 2, 4, and 6 h for agents given every 6 h; at 2, 4, and 8 h for agents given every 8 h; and at 2, 6, and 10 h for agents given every 12 or 24 h	residual diuresis: oligo-anuric patients would benefit from a 30-min bolus, while a 3-h extended infusion would be more appropriate for those patients with preserved diuresis For the attainment of the classic PD target for carbapenems (40% of the $T > MIC$), a standard dose of 500 mg q8h as a bolus over 30 min would be sufficient for all cases For the attainment of a more aggressive target, such as a C_{min}/MIC ratio of 5, doses of 1000 mg q8h as a 3-h infusion or higher would be required All the studied agents were easily filtered ($SC > 0.7$) with the exception of flucloxacillin There was a high interindividual variability in the SC values of the studied drugs, in particular for ceftazidime and to a lesser degree for amoxicillin and flucloxacillin The correlation between observed and predicted clearance was significant ($P = 0.003$) only when all drugs were combined, not for the individual antimicrobial drugs. Despite the nonsignificant correlation, the difference between predicted and observed clearance for all drugs was small, with the exception of ceftazidime	There was no significant correlation between predicted and observed CWH drug removal. However, for clinical practice, dose adjustment according to the predicted CWH removal provides a more reliable estimate than that according to the observed CWH removal Although there is interpatient variability between the observed and predicted $Cl_{C_{WHF}}$ values for some antibiotics, its effect on dosing strategies is not necessarily clinically relevant: flucloxacillin has an important non-renal elimination route, and therefore the $F_{C_{WHF}}$ value was extremely low and not affected by the wide interindividual variation in observed $Cl_{C_{WHF}}$. Also, wide therapeutic range, such as with ceftazidime, makes it safe to use the predicted $Cl_{C_{WHF}}$ for dose adjustment

AKI acute kidney injury, *AUC* area under the curve, *Cl_C* creatinine clearance, *Cl_{dialysis}* extracorporeal clearance, *C_{max}* maximal concentration, *C_{min}* minimal concentration, CRRT continuous renal replacement therapy, *C_{WHF}* continuous venous-venous hemofiltration, *C_{WHDF}* continuous venous-venous hemodiafiltration, *C_{WHDF}* continuous veno-venous hemodiafiltration, *F_{up}* unbound fraction of a drug, *MIC* minimal inhibitory concentration, *PTA* probability of target attainment, *S* saturation coefficient, *t_{1/2}* half-life, *T > MIC* time above MIC, *UFR* ultrafiltration rate, *V_d* volume of distribution

Table 2 PK/PD studies of beta-lactams in patients with sustained low-efficiency dialysis or extended daily dialysis

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Kielstein et al. [82]	To evaluate PK of meropenem in critically ill patients with renal failure undergoing EDD	Meropenem	Prospective clinical study Adult ICU patients with anuric acute renal failure being treated with EDD and receiving meropenem (n = 10) Meropenem administered as 1 g dose, as an intravenous infusion over a period of 30 mins, 6 h before EDD was started Blood samples were drawn before administration of the drug, at 0.5, 1, 2, 4, and 6 h after its administration; before EDD; during EDD, at time points 2, 4, and 6 h; at the end of EDD; and at 0.5, 1, 3, and 8 h after EDD. Additional blood samples were drawn pre- and post-dialyzer in order to calculate the dialyzer clearance	The average (mean ± SD) dialysis time during the study was 480 ± 6 min, and mean blood and countercurrent dialysate flow was 160 ± 3 mL/min T1/2off was 8.7 h [4.7–30] T1/2on was 3.7 h [2.1–4.7] Vd was 0.72 L/kg [0.35–2.78] CLoff 5.01 L/h [2.44–11.15] CLdial was 2.3 L/h [0.7–3.7] (estimated from the drug amount recovered in the dialysate) and 5.1 L/h [4.3–5.7] (estimated from drug concentrations before and after application of the dialysis membrane)	Meropenem is significantly eliminated by EDD. Compared with PK results in the literature for intermittent dialysis and CRRT, dosing regimens cannot be used for critically ill septic patients with renal failure being treated with EDD EDD eliminates meropenem at least to an extent similar to CWH. Thus, physicians run the risk of underdosing. A dose of 0.5 to 1.0 g meropenem every 8 h is recommended. The exact dose should be tailored according to weight and severity of illness as well as to the current MIC against the incriminated bacteria. Whenever possible, therapeutic drug monitoring should be performed
Lorenzen et al. [83]	The aims of this study were to evaluate the PK of ampicillin/sulbactam in critically ill patients with AKI undergoing extended dialysis and to establish a dosing recommendation for this treatment method	Ampicillin-sulbactam	Prospective, open-label, observational study 12 adult ICU patients with anuric AKI PK after a single dose of ampicillin/sulbactam (2 g/1 g) over a period of 30 min was obtained in 12 patients. Multiple-dose PK after 4 days of twice-daily ampicillin/sulbactam (2 g/1 g) was obtained in three patients. The average dialysis time was 442 ± 77 min and mean blood and counter current dialysate flow was 162 ± 6 mL/min, resulting in a mean urea reduction ratio of 50.1% ± 2.7%. ED was started 3 h after the end of the ampicillin/sulbactam infusion	Cmax 280.9 ± 174.9 mg/L Tmax 0.5 h AUClast 847.5 ± 499.5 mg·h/L t1/2 2.8 ± 0.8 Vd (L) 13.1 ± 11.1 Clot 61.1 ± 55.2 mL/min CLdial 80.1 ± 7.7 mL/min	Ampicillin/sulbactam is eliminated by ED Current dosing recommendations from patients undergoing IHD (3 g every 24 h) would cause a significant underdosing of the drug in patients treated with ED Ampicillin/sulbactam concentrations exceeded MIC90 values of <i>Enterobacteriaceae</i> , such as <i>Escherichia coli</i> or <i>Klebsiella pneumoniae</i> (MIC90 < 2.0 mg/L) or <i>Enterococcus faecalis</i> (MIC90 = 2.0 mg/L), only for 8 h (approximately 30% of the dosing interval for patients on intermittent hemodialysis) after start of infusion. A dosage of 3 g every 12 h in patients undergoing ED does not lead to a significant accumulation of the drug
Burkhardt et al. [84]	To evaluate PK of ertapenem, with once-daily dosing, in critically ill patients with anuric acute renal failure undergoing EDD	Ertapenem	Prospective, open-label study 6 adult ICU patients undergoing EDD treated with 1 g ertapenem as a single intravenous dose Blood samples were collected before ertapenem infusion and 0.5, 1, 2, 4, 6, 8 h after the end of the infusion and also 2, 4, 6, and 8 h after the start of EDD. Additional blood samples were drawn pre- and post-dialysis in order to calculate the dialyzer clearance. To study post-EDD PK, samples were drawn 0.5, 1, 3, and 8 h after the end of EDD	Cmax 81.3 ± 12.1 mg/L AUC0-inf 687.4 ± 212.0 mg·h/L T1/2off 18.9 ± 5.4 h T1/2on 6.7 ± 0.4 h Vd (L) 15.9 ± 3.2 CLoff 19.3 ± 11.4 mL/min CLdial 49.5 ± 10.9 mL/min T > MIC was 100% (MIC90 ≤ 1 mg/L) and 85% (MIC90 ≤ 2 mg/L)	1 g ertapenem per day to critically ill patients with ARF in the ICU that undergo EDD is necessary to ensure optimal free concentrations of ertapenem. A reduction of the dose is not supported by our data. Further dosing recommendations for patients with renal failure in the ICU treated with such effective modes of renal replacement therapy should be developed to avoid excess mortality due to under-dosing of life-saving medication

Table 2 PK/PD studies of beta-lactams in patients with sustained low-efficiency dialysis or extended daily dialysis (Continued)

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Tamme et al. [85]	To describe the PK of piperacillin and tazobactam during extended high volume hemodiafiltration to define optimal dosing	Piperacillin-tazobactam	Prospective, observational study 10 adult ICU patients with sepsis and AKI requiring CRRT A single dose of 4000 mg of piperacillin and 500 mg tazobactam was administered as a 30-min intravenous infusion 1 h after the start of HVHDF Blood samples of 4 mL were collected before and immediately after the end of piperacillin/tazobactam infusion and 60, 90, 120, 150, 180, 240, 300, 360, 420, and 480 min after the start of drug administration	The plasma concentration-time profiles of piperacillin and especially tazobactam demonstrated high interindividual variability For piperacillin CL (range) 6.9 L/h (6.1–7.9) Vd central compartment (range) 9.0 L (7.4–11.0) Vd peripheral compartment (range) 11.2 L (8.9–14.2) For tazobactam CL (range) 5.1 L/h (4.1–6.3) Vd central compartment (range) 8.6 L (6.9–10.7) Vd peripheral compartment (range) 8.9 L (6.6–12.0) Using Monte Carlo simulation, the probability of 100% ft > MIC target attainment for piperacillin/tazobactam 4.5 g dosed every 6 and 8 h as 4-h infusion were 88.6% and 61.0%, respectively, for MIC 16 mg/l	Application of extended HVHDF for the treatment of AKI in septic shock patients results in considerable clearance of piperacillin and tazobactam Piperacillin/tazobactam doses of 4.5 g, administered every 8 h as 0.5-h infusion during HVHDF, ensured more than 80% probability of attaining the 50% ft > MIC target for intermediately susceptible bacteria (MIC 16 mg/l) While aiming for 100% ft > MIC of 16 mg/l, increasing doses to 4.5 g every 6 h and prolonging the infusion time to 4 h would be necessary

AKI acute kidney injury, ARF acute renal failure, *Cl_{dial}* dialysis clearance, *Cl_{off}* drug clearance without dialysis, *EDD* extended daily dialysis, *ICU* intensive care unit, *SD* standard deviation, *SLED* sustained low-efficiency dialysis, *T_{1/2off}* half-life before/after EDD, *T_{1/2on}* half-life during EDD, *V_d* volume of distribution

Table 3 PK/PD studies of beta-lactams in patients with extracorporeal membrane oxygenation

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Donadello et al. [100]	To investigate whether ECMO could alter the pharmacokinetics of meropenem and piperacillin/tazobactam in ICU patients	Meropenem Piperacillin/tazobactam	Retrospective, case-control study in 67 ICU patients Antibiotics daily dosing was done according to renal dosing Beta-lactam plasma concentrations were measured T2 and just before administration of the subsequent dose T0 TDM results in ECMO patients (W or VA) were matched (1:1) to TDM results of non-ECMO patients (total 41 TDM matches) according to the following criteria: drug regimen; renal function (same CrCl or, if on CRRT, same CRRT intensity with an acceptable difference of 10%); total body weight; SOFA score at the time of treatment initiation; age	For both antibiotics, there were numerical differences but with no statistical significance in V_d , $t_{1/2}$ and CL between ECMO patients and controls The proportions of insufficient (13/41 vs 12/41), adequate (15/41 vs 19/41), and excessive (13/41 vs 10/41) drug concentrations were similar in ECMO and control patients	PK parameters and TDM results were not significantly altered in ECMO patients compared with control ICU patients Almost 30% of the overall TDM results were associated with insufficient antibiotic concentrations to optimally treat <i>P. aeruginosa</i>
Shekar et al. [101]	To describe single-dose meropenem PK during ECMO using critically ill patients with sepsis and not receiving ECMO as controls	Meropenem	Open-label, descriptive, matched-cohort PK study Adult ICU patients on ECMO (no RRT $n = 6$; on RRT $n = 5$) and controls (no renal dysfunction $n = 5$; on RRT $n = 5$) Meropenem doses in ECMO: 1 g/8/8 h ($n = 8$); 1.5 g/8/8 h ($n = 2$) and 2 g bolus; and 1 g/8/8 h ($n = 1$) Controls: 1.5 g bolus and 1 g/8/8 h (no renal dysfunction); 1 g/8/8 h	Controls vs ECMO Cmax: 65.4 (58.7–74.4) vs 55.3 (37.8–60.4) mg/L Cmin: 4.2 (0.0–5.7) vs 7.2 (4.0–17.2) mg/L Vd: 0.45 ± 0.17 vs 0.41 ± 0.13 L/kg, $P = 0.21$ Clearance: 7.9 ± 5.9 vs 11.7 ± 6.5 L/h, $P = 0.18$ ECMO patients, trough concentrations > 2 mg/L were achieved in all patients. Through concentrations > 8 mg/L (targeting less susceptible microorganisms) were achieved in only 8 out of 11 patients, 5 of them being on RRT	Standard meropenem dosing (1 g IV 8-hourly) as an intermittent bolus infusion in ECMO patients is likely to result in drug concentrations sufficient to treat highly susceptible Gram-negative pathogens However, when treating less susceptible <i>P. aeruginosa</i> (MIC ₅₀ 8 mg/L) and <i>Acinetobacter</i> species (MIC ₅₀ 16 mg/L) higher meropenem doses may have to be considered
Welsch et al. [102]	To report the cases of two patients on W ECMO for refractory ARDS following lung transplantation and treated empirically with imipenem	Imipenem	Case report Imipenem 1 g every 6 h Serum and mini-BAL samples (native and transplant lung) collected at steady state after 2 days of therapy immediately before the fifth drug dose <i>Enterobacter cloacae</i> was isolated from the respiratory sample of patient 1 and <i>Klebsiella pneumoniae</i> was isolated from the respiratory sample of patient 2 MIC of the two isolated strains were 0.125 and 0.25 mg/L, respectively	BAL concentrations were undetectable (<0.5 mg/L) Serum T > MIC of both microorganisms was 100% Considering more resistant microorganisms, such as <i>P. aeruginosa</i> with MIC > 2 mg/L, the probability of achieving a fractional time above MIC > 50% or 100% was also high There was great variability in the residual serum concentration of imipenem between the two patients	An elevated dosing regimen (4 g/24 h) is more likely to optimize drug exposure, and therapeutic drug monitoring is recommended

ARDS acute respiratory distress syndrome, BAL bronchial-alveolar lavage, CL clearance, CrCl creatinine clearance, CRRT continuous renal replacement therapy, ECMO extracorporeal membrane oxygenation, ICU intensive care unit, MIC minimal inhibitory concentration, RRT renal replacement therapy, SOFA sequential organ failure assessment, T > MIC percentage of time above minimal inhibitory concentration, T0 0 h after the start of infusion, T2 2 h after the start of infusion, TDM therapeutic drug monitoring, V_d volume of distribution, venous-arterial, WV venous-venous

However results are conflicting concerning decreased mortality and bacteriological and clinical cure rates [144]. A sub-analysis from the DALI study compared intermittent bolus vs prolonged infusions of beta-lactams in patients with respiratory infection and concluded that patients receiving beta-lactams via prolonged infusion demonstrated significantly better 30-day survival [145].

Falagas et al. [114] conducted a meta-analysis of 14 studies comparing continuous and short-term infusion of carbapenems and piperacilin-tazobactam, involving 1229 patients. Mortality was lower among patients receiving extended or continuous infusion of carbapenems or piperacillin/tazobactam compared to those receiving short-term infusion (risk ratio (RR) 0.59, 95% confidence interval (CI) 0.41–0.83). Patients with pneumonia who received extended or continuous infusion had lower mortality than those receiving short-term infusion (RR 0.50, 95% CI 0.26–0.96) [114].

An interesting retrospective study by Huang et al. [120] reviewed 68 neurosurgical patients with post-operative intracranial infections treated with 4 g/day cefepime over 24 h as a continuous infusion (CI; $n = 34$) or 2 g every 12 h as intermittent infusion (II; $n = 34$). CI controlled the intracranial infection more rapidly and effectively than II (6.6 ± 1.9 days vs 7.8 ± 2.6 days; $P = 0.036$). PD targets were more achievable with CI: for plasma cefepime concentrations, the percentage $fT > MIC$ in the CI group was higher than in the II group (for MICs of 8 $\mu\text{g/mL}$, 100% vs 75%, respectively). For cerebral spinal fluid (CSF) cefepime concentrations, the percentage $fT > MIC$ in the CI group was higher than in the II group (for MICs of 4 $\mu\text{g/mL}$ and 8 $\mu\text{g/mL}$, 83.3% and 75% vs 25% and 0%, respectively) [120].

De Waele et al. [27] reviewed 343 patients from 68 ICUs across ten countries and concluded that use of intermittent infusion was the most significant factor associated with target non-attainment, for both 50% and 100% $fT > MIC$. Other risk factors for target non-attainment were CLCr, recent surgery, and timing from initial antibiotic therapy and sampling. However, the type of infusion was such a significant covariate in the model that it eliminated the effects of other variables [27].

Site of infection

Usually drug concentrations in blood are used to determine PD parameters, such as percentage of time drug levels exceed the MIC and peak drug AUC/MIC level, due to the relative accessibility of this body fluid. Because infection usually occurs at extravascular sites, the use of drug concentrations in blood is only satisfactory if blood levels are an adequate surrogate for levels at the site of infection [13]. In septic shock, blood misdistribution in the microcirculation might decrease antibiotic concentration at the infection site [1].

Boydjiev et al. [146] studied ertapenem penetration into muscle in mechanically ventilated patients and concluded that average muscle free-ertapenem concentrations were above the MIC values of targeted pathogens except in a few patients. Karjagin et al. [147] evaluated the PK/PD relations of meropenem in plasma and peritoneal fluid by microdialysis and showed that area under the concentration–time curve was lower in peritoneal fluid than in plasma, concluding that in patients with severe peritonitis associated with septic shock, a dosing regimen of 1 g infused over 20 min every 8 h is sufficient against susceptible bacteria, but not always against intermediately susceptible bacteria. Also, beta-lactam PK is variable between plasma and subcutaneous interstitial fluid in septic patients [148]. Thus, prediction of microbiological outcome based on concentrations in plasma results in overestimation of antimicrobial activity at the site of infection.

Special anatomic barriers (e.g., brain, eye, and prostate) can result in drug levels being much lower than free drug levels in plasma [13]. The combination of tight junctions and active transport systems that form the blood–brain barrier creates a substantial impediment to the penetration of most antibiotics into the CSF. However, the presence of inflammation within the meninges significantly alters the permeability of the blood–brain barrier, increasing CSF exposure for the majority of antibiotics [20]. For meningitis, CSF levels are appropriate for determination of PD parameters.

Very few studies have investigated PK/PD issues in the CSF (Table 4). Five case reports, one randomized clinical trial in a paediatric population, and three prospective observational studies found good probability of target attainment for susceptible strains but standard dosing may not be optimal for less susceptible strains. Prolonged and/or continuous infusion is of benefit in the attempt to achieve PD targets. No data regarding intermittent versus continuous CSF ventricular drainage were found and conceptually these two types of drainage may alter the beta-lactam PK profile.

There is very sparse data on possible surrogate central nervous system penetration factors for beta-lactams, so no conclusions can be made. We recommend to use higher than standard dosing, preferably with continuous or prolonged infusions, especially when treating less susceptible bacterial strains. Toxicity did not increase at increased doses. Finally, none of these studies addressed clinical outcome.

Though there are PK models of plasma concentrations of beta-lactams specifically for the critically ill population with pneumonia, it is suggested that epithelial lining fluid (ELF) concentrations are important determinants of efficacy of treatment of bacterial pneumonia. ELF-to-serum penetration ratios may vary widely among

Table 4 PK/PD studies of beta-lactams in cerebral spinal fluid

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Goldwater et al. [149]	To evaluate antibiotic CSF penetration and antimicrobial efficacy	Ceftriaxone Cefotaxime	Randomised, open, comparative trial 120 paediatric patients with meningitis, 33 with repeated lumbar puncture at different times CRO 100 mg/kg once daily; CTX 50 mg/kg every 6 h	All 33 repeated lumbar punctures were sterile The lowest CSF level recorded (0.45 µg/mL for CTX) was 45 times the MIC (0.01 µg/mL). The highest levels (24–35 µg/mL for CRO) were up to 8750 times the MIC of the patient's causative organism	Antibiotic levels achieved in CSF were therapeutic, being well above the MIC for all organisms encountered CSF cell count had no apparent influence on antibiotic levels
Lonsdale et al. [150]	To illustrate issues in the management of CSF antibiotic concentrations	Meropenem	Case report Neurosurgical patient with external ventricular drain-related ventriculitis	Adequate plasma through concentrations achievable after increasing dosing to 2 g, four times daily CSF concentrations of meropenem were similar to those seen in plasma There were variations in CSF drug penetration	Achieving CSF therapeutic antibacterial concentrations in neurosurgical critically ill patients is difficult Standard antibacterial prescription is potentially flawed in this setting, suggesting the need for therapeutic drug monitoring
Abdul-Aziz et al. [151]	To report the difficulty in achieving and maintaining target antibiotic exposure in critically ill patients with deep-seeded infections	Flucloxacillin	Case report Critical care patient with CNS infection	Trough plasma concentrations were below the MIC; CSF concentrations were undetectable (intermittent doses 2 g 6/6 h) With continuous infusion and increasing the dose to 20 g daily, the plasma and CSF levels became detectable, albeit lower than the predefined targets	Antibiotic pharmacokinetics may be significantly altered in critically ill patients Applying continuous infusion and monitoring plasma and CSF levels is of significance to optimize antibiotic delivery
Cies et al. [152]	To describe the pharmacokinetics of continuous-infusion of meropenem	Meropenem	Case report. Paediatric patient with ventriculitis	Serum levels were 12 µg/mL at 2 h and “undetectable” at 4 h, with CSF levels of 1 and 0.5 µg/mL at 2 and 4 h, respectively (MIC < 0.25) On continuous infusion, serum, and CSF levels were noted to be 13 and 0.5 µg/mL, respectively	The continuous-infusion dosing regimen allowed for 100% probability of target attainment in the serum and CSF and a successful clinical outcome
Dahyot-Fizler et al. [153]	To describe brain distribution of cefotaxime by microdialysis in patients with acute brain injury	Cefotaxime	Observational, prospective study 5 ICU patients with acute brain injury, treated for lung infection Cefotaxime 2 g 8/8 h	Mean AUC_{0-6h}/AUC_{plasma} ratio was $26.1 \pm 12.1\%$ Unbound cefotaxime brain concentrations were much lower than corresponding plasma concentrations Simulated brain concentration at two dosage regimens (used for treatment of meningitis) showed $T > MIC$ higher than 90% of the dosing interval for both dosing regimens (4 g every 6 h or 8 h) for susceptible strains and only for 4 g every 6 h for resistant ones	There is limited brain distribution of cefotaxime Higher cefotaxime dosage (4 g 6/6 h) is required to treat meningitis with resistant bacterial strains

Table 4 PK/PD studies of beta-lactams in cerebral spinal fluid (Continued)

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Morita et al. [154]	To assess the efficacy, safety, and concentration of meropenem in cerebrospinal fluid	Meropenem	Observational, prospective study 5 adult ICU patients with meningitis Meropenem 2 g/8h (duration of infusion was variable from 0.5 to 2 h) CSF and blood were obtained pre-treatment and on days 3, 7, 14, and 21	Concentrations in cerebrospinal fluid ranged from 0.27 to 6.40 µg/mL up to 8.47 h and were over 1 µg/mL 3 h after starting meropenem infusion The CSF/plasma concentration ratio ranged from 0.008 to 0.013, 0.011 to 0.953, and 0.633 to 1.821, respectively, within 2 h, 2–6 h, and after 6 h from the start of drug infusion The relationship of CSF concentration to CSF cell counts, CSF/plasma glucose ratio, and CSF protein concentration, respectively, was statistically correlated with CSF cell counts and CSF protein concentration, and inversely correlated with CSF/plasma glucose ratio	Concentration of meropenem in CSF exceeded the minimal inhibitory concentration for the pathogens involved (penicillin sensitive <i>S. pneumoniae</i> and <i>S. salivarius</i> ; methicillin-sensitive <i>Staph aureus</i>) No serious adverse event and no discontinuation of treatment occurred The meropenem concentration in the CSF can be expected to be high because of the presence of inflammation
Tsumura et al. [155]	To examine PK and PD of meropenem in cerebrospinal fluid	Meropenem	Observational, prospective study 6 neurosurgical patients Meropenem (0.5 g every 8 h) was administered during 0.5 h Lumbar CSF and venous blood samples were obtained at 0.5–16 h after the start of the first infusion	Penetration into the CSF with the AUC ratio was 0.10 ± 0.03 (mean ± SD) A dosage of 0.5 g q8h achieved a > 90% PTA (50% of the T > MIC), and 1 g q8h was needed for a > 90% PTA (100% of the T > MIC) for susceptible isolates For <i>P. aeruginosa</i> , 2 g q8h achieved a lower PTA	Less susceptible bacterial CNS infections may not be optimized with standard meropenem dosage
Nicasio et al. [156]	To describe the use and cerebral spinal fluid penetration of a prolonged infusion meropenem regimen in a patient with <i>Serratia marcescens</i> meningitis	Meropenem	Case report Adult patient with meningitis and epidural abscess Meropenem 2 g q8h, 3-h infusion	The prolonged (3 h) infusion regimen of 2 g 8 h resulted in concentrations in both serum and CSF above the MIC of 0.047 µg/mL, for 100% of the dosing interval CSF penetration was 6.4%	The use of a high-dose prolonged infusion of meropenem resulted in adequate exposure at the site of infection and a successful clinical response At follow-up, the patient had completed a 4-week course without relapse or adverse events
Frasca et al. [157]	To describe PK–PD profile of cefotaxime in the CSF	Cefotaxime	Case report Adult ICU patient with TBI Cefotaxime 4 g q8h, 30-min infusion Microdialysis was performed on day 4, after the 12th dose	Unbound plasma Cmax was 118.8 µg/mL CSF Cmax was 11.4 µg/mL T > MIC in the brain were, respectively, 78% (6.2 h) and 46% (3.7 h) for MIC values of 2 and 4 µg/mL	ECF brain concentrations indicate that an adequate exposure to cefotaxime is achieved in prevention and treatment of most CNS infections with the standard dosage regimen
Wang et al. [158]	To explore whether there is increased CSF penetration of cefoperazone/sulbactam when there is blood–brain barrier impairment following craniotomy; and whether extended infusion time affects drug concentrations	Cefoperazone/sulbactam	Observational, prospective study Dosing was 3.0 g in a 3-h infusion every 6 h, after craniotomy Venous blood and CSF were collected before the start of drug administration and at hour 1, 2, 3, 4, 6, 12, 15, 16, and 18 after administration 8 neurosurgical adult patients enrolled	CSF penetration: Peak concentrations (CSF/serum): 8.6% ± 7.2% for cefoperazone and 13.5% ± 11.9% for sulbactam Trough concentrations (CSF/serum): 13.4% ± 5.3% for cefoperazone and 106.5% ± 87.5% for sulbactam Ratio of the AUC of CSF and serum: 14.5% for cefoperazone and 22.6% for	If cefoperazone/sulbactam single infusion time is extended to 3 h, the serum drug concentration achieved the PK/PD standard of > 50%T > MIC (MIC ₉₀ 64 mg/L) It is very difficult to achieve this PK/PD standard in the CSF, and a higher dose might be needed to treat intracranial infections

Table 4 PK/PD studies of beta-lactams in cerebral spinal fluid (Continued)

Study	Endpoints	Antibiotic	Design	Results	Conclusions
				sulbactam Cefoperazone serum concentrations achieved > 50%T > MIC for <i>Pseudomonas</i> and <i>Acinetobacter</i> (MIC ₉₀ 64 mg/L) and 100%T > MIC for more susceptible bacteria CSF cefoperazone T > MIC% was almost 100% (<i>Escherichia coli</i> MIC ₅₀) and T > MIC% was more than 50% (<i>Acinetobacter baumannii</i> MIC ₅₀) Sulbactam serum concentrations achieved > 50%T > MIC for <i>Acinetobacter</i> (MIC ₉₀ 16 mg/L) CSF sulbactam concentrations did not reach the level of MIC ₅₀ of 8 mg/L for <i>Acinetobacter</i>	Destruction of the blood–brain barrier after craniotomy can increase the CSF concentration to a certain extent

AUC area under the curve, CNS central nervous system, CRO ceftriaxone, CSF cerebral spinal fluid, CTX cefotaxime, ICU intensive care unit, MIC minimal inhibitory concentration, MIC₅₀ minimal inhibitory concentration for 50% of isolates, PTA probability of target attainment, SD standard deviation, T > MIC percentage of time above minimal inhibitory concentration, TBI trauma brain injury

beta-lactams [13, 20, 159]. The impact of infection on their penetration into ELF in humans is unknown [159], though some reports state that ELF penetration increases in acute lung injury [160].

Only a few studies have investigated beta-lactam PK/PD issues in critically ill patients with pneumonia (Table 5) and in only seven of them were ELF drug concentrations measured. A standard dosage of beta-lactams derived from healthy patients' PK profiles may be insufficient for treatment of critically ill patients with pneumonia, especially when caused by multidrug-resistant pathogens. Continuous or prolonged infusions and higher than standard doses improve the PD profiles of these antibiotics. This is very important to achieve an adequate PD profile when treating less susceptible bacterial strains. Therapy drug monitoring would be extremely helpful in this setting.

New beta-lactam drugs and beta-lactamase combinations

Of great concern is the worldwide increase in the number of infections caused by Gram-negative multidrug-resistant bacteria. Treatment choices for these infections have been limited, especially for infections caused by bacteria that produce carbapenemases and/or extended-spectrum beta-lactamases.

Ceftolozane–tazobactam and ceftazidime–avibactam are 2 beta-lactams/beta-lactamase combinations with anti-Gram-negative bacteria activity that were recently approved for the treatment of complicated intra-abdominal infections, complicated urinary tract infections, and nosocomial pneumonia.

Ceftolozane is an oxymino-aminothiazolyl cephalosporin with a pyrazole substituent at the 3-position side chain instead of the lighter pyridium present in ceftazidime. This heavier side chain provides improved steric hindrance to prevent hydrolysis mediated through AmpC beta-lactamases.

Ceftolozane–tazobactam combines a novel cephalosporin with an established beta-lactam beta-lactamase inhibitor, whereas ceftazidime–avibactam couples a well-known cephalosporin with a novel non-beta-lactam beta-lactamase inhibitor.

Both tazobactam and avibactam target the active site of serine beta-lactamases. Tazobactam, a beta-lactam sulfone, binds irreversibly to the active site of beta-lactamases and avibactam is a diazabicyclooctane non-beta-lactam that binds covalently and reversibly to beta-lactamases. This reversibility is a unique feature that allows avibactam to undergo recyclization to inactivate another beta-lactamase. The crucial advantage of avibactam is its ability to inhibit extended spectrum beta lactamases, AmpC beta-lactamases (as expressed in *Pseudomonas aeruginosa* and *Enterobacteriaceae*), and class A carbapenemases of the *Klebsiella pneumoniae* carbapenemase (KPC and OXA-48) family.

The pharmacokinetic and safety profiles of this antibiotic have been established in healthy adults and subjects with various degrees of renal function [170, 171]. The currently approved dosages for adult patients with an estimated CrCl > 50 mL/minute are ceftolozane 1 g with tazobactam 500 mg every 8 h and ceftazidime 2 g with avibactam 500 mg every 8 h for complicated urinary tract infections and intra-abdominal infections [172] and ceftolozane 2 g with tazobactam 1 g every 8 h for nosocomial pneumonia [173].

However, data guiding its use in critically ill patients are currently sparse, being entirely derived from studies with very few patients and/or case reports.

Veillette et al. [174] presented PK data for ceftazidime–avibactam from two patients with bloodstream infections caused by carbapenemase (KPC)-producing *K. pneumoniae*; the patients had renal impairment and one of them was obese. In both patients half-lives were prolonged and Vd larger than predicted. They conclude that recommended doses and intervals may not be sufficient for obese patients with renal failure, especially for those infected with KPC-producing organisms [174].

Oliver et al. [175] evaluated the adequacy of extended-infusion ceftolozane–tazobactam to achieve target PK and PD goals in a critically ill patient with *Pseudomonas aeruginosa* pneumonia and septic shock on CVVH. A dosage of 1.5 g every 8 h (3-h infusion) was given. All estimated plasma-free drug concentrations achieved the PD goals and remained well above the isolated organism's MIC of 1.5 $\mu\text{g/mL}$ and above the susceptibility breakpoint of 4 $\mu\text{g/mL}$ throughout the dosing interval, although the authors could not comment on drug concentrations at the site of infection. The authors conclude that, given the lowest estimated free-drug concentration was fivefold greater than the susceptibility breakpoint, the estimated half-life of 28 h and the low extraction ratio observed, a lower total daily dose might be utilized and an extended infusion time may not be necessary for patients on CVVH [175].

Bremmer et al. [176] performed a PK analysis of intravenous ceftolozane–tazobactam 3 g every 8 h in a critically ill patient with *P. aeruginosa* pneumonia on CVVHDF. They concluded that, compared with a patient with normal renal function, this patient had decreased ceftolozane clearance. A ceftolozane–tazobactam dosage of 1.5 g every 8 h should adequately achieve a desired drug concentration above the minimum inhibitory concentration of 8 $\mu\text{g/mL}$ for the treatment of pneumonia [176].

Stokem et al. reported the successful treatment with ceftolozane–tazobactam 3 g every 12 h for a pulmonary exacerbation in a 35-year-old female post-lung transplant, with cystic fibrosis, malnutrition, chronic kidney disease, and multi-drug resistant *P. aeruginosa* infection. Optimal time above MIC (estimated 100% time above

Table 5 PK/PD studies of beta-lactams in bronchial-alveolar lavage

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Kikuchi et al. [52]	Compare the PK/PD parameters of biapenem in bronchial ELF given as 0.5-h and 3-h infusions	Biapenem	Prospective, non-blinded, crossover study 6 healthy adult volunteers	The percentage (mean ± SD) of T > MIC in bronchial ELF ranged from zero (MIC 4 µg/mL) to 34.6% ± 5.2% (MIC 0.25 µg/mL) after the 0.5-h infusion and from 5.1% ± 5.6% (MIC 4 µg/mL) to 52.2% ± 17.0% (MIC 0.25 µg/mL) after the 3-h infusion	A 3-h infusion of biapenem tended to produce a higher T > MIC in bronchial ELF, as well as in plasma, than a 0.5-h infusion. These results support the use of prolonged infusions for successful treatment of lower respiratory tract infections based on PK/PD parameters in bronchial ELF
Rodvold et al. [159]	To define the exposure targets in lung associated with good microbiological activity in a murine model To determine drug penetration into ELF in humans	Ceftibiprol	Prospective, observational study Pre-clinical, murine model Clinical study with 24 healthy volunteers 500 mg every 8 h, 2-h infusion regimen	Murine model: for cell kills of 1 and 2 log ₁₀ CFU/g, total drug must be present in ELF at a concentration in excess of the MIC of 12.9% and 24% of a 24-h interval, respectively. ELF penetration was 69% (median) Mean ELF penetration in human volunteers was 25.5% (median, 15.3%; interquartile range, 7.9% to 30.4%) Target attainment falls below 90% for a cell kill of 2 log ₁₀ CFU/g at a MIC of 1.0 mg/L and for a 1 log ₁₀ CFU/g cell kill at a MIC of 2.0 mg/L	Ceftibiprol penetrated into ELF very differently in humans compared to mice. For seriously ill patients, particularly in the ICU, higher doses or longer infusion times (to prolong the time > MIC), or both, will be required to comfortably ensure a 90% target attainment for seriously ill patients with MRSA pneumonia
Conte et al.	Determine the plasma and intrapulmonary ELF and AC pharmacokinetic parameters of intravenously administered meropenem	Meropenem	Prospective, observational Four doses (8/8 h) of 0.5 g, 1.0 g or 2.0 g were administered intravenously to 20, 20, and 8 healthy adult subjects, respectively	C _{max} , AUC, T _{1/2z} : 0.5 g group: serum 25.8 ± 5.8 µg/mL, 28.57 µg h/mL, 0.77 h; ELF 5.3 ± 2.5 µg/mL, 12.27 µg h/mL, 1.51 h; AC 1.0 ± 0.5 µg/mL, 4.30 µg h/mL, 2.61 h 1 g group: serum 53.5 ± 19.7 µg/mL, 55.49 µg h/mL, 1.31 h; ELF 7.7 ± 3.1 µg/mL, 15.34 µg h/mL, 0.95 h; AC 5.0 ± 3.4 µg/mL, 14.07 µg h/mL, 2.17 h 2 g group: serum 131.7 ± 18.2 µg/mL, 156.7 µg h/mL, 0.89 h T ≥ MIC (MIC90 values of 0.12–4 µg/mL): 0.5 g group: serum 28–78%, ELF 18–100%, AC 0–100% 1 g Group: serum 45–100%, ELF 25–88%, AC 24–100%	The prolonged T > MIC90 and high intrapulmonary drug concentrations following every 8 h administration of 0.5–2.0 g doses of meropenem are favorable for the treatment of common respiratory pathogens.
Boselli et al. [161]	Determine the steady-state serum and alveolar concentrations of piperacillin/tazobactam administered in continuous infusion (12/1.5 g/day or 16/2 g/day) at various degrees of renal failure (terminal renal failure excluded)	Piperacillin/tazobactam	Prospective, open-label, comparative, single center 40 ICU patients with ventilator-associated pneumonia Samples collected after 2 days of treatment	Median (interquartile) serum and alveolar piperacillin concentrations No/mild renal failure (creatinine clearance ≥ 50 mL/min) Serum 25.3 mg/L (23.1–32.6) and alveolar 12.7 mg/L (6.7–18.0) for 12/1.5 g/day Serum 38.9 mg/L (32.9–59.6) and alveolar 19.1 mg/L (14.0–21.5) for 16/2 g/day Moderate/advanced renal failure (creatinine clearance < 50 mL/min) Serum 102.4 mg/L (97.4–112.6) and alveolar 44.1 mg/L (33.4–48.3) for 12/2 g/day Serum 135.3 mg/L (119.5–146.2) and alveolar 54.9 mg/L (45.2–110.3) for 16/2 g/day Alveolar penetration was 40–50% for piperacillin and 65–85% for tazobactam There was a positive linear relationship between ELF and serum concentration in both patients receiving 12/1.5 g/day	The administration of daily continuous infusions of P/T 12/1.5 g or even 16/2 g might provide insufficient alveolar concentration to eradicate high-risk pathogens with high MICs such as multi-drug resistant <i>P. aeruginosa</i> . A linear relationship between alveolar and serum piperacillin concentration was observed in this study, with ELF piperacillin concentration being 40–50% of corresponding serum values

Table 5 PK/PD studies of beta-lactams in bronchial-alveolar lavage (Continued)

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Cousson et al. [162]	Compare continuous vs intermittent administration of drug Determine average concentration in ELF Determine the fraction of time, during the first 48 h of treatment, where serum concentrations remained above the 20 mg/L plasma threshold	Ceftazidime	Single-center, controlled, randomized trial in two parallel groups comparing two modes of administration: group A, loading dose 20 mg/kg + 60 mg/kg/day; group B, 20 mg/kg over 30 min every 8 h 34 adult ICU patients with ventilator-associated pneumonia due to Gram-negative bacilli. The mean MIC of ceftazidime for <i>P. aeruginosa</i> was estimated at 2 mg/L. The target threshold value for ceftazidime in the ELF was thus fixed at 8 mg/L, with a corresponding serum threshold of 20 mg/L	($r = 0.8437$, $P < 0.0001$) and 16/2 g/day ($r = 0.7935$, $P < 0.0001$) T > MIC was 100% (assuming MIC > 16 mg/L) in both serum and ELF in all patients with moderate/advanced renal failure; in patients with no/mild renal failure this threshold was not reached in 6 patients with 12/0.5 g dosing and 4 patients with 16/2 g dosing Plasma T > 20 mg was 100% in group A versus 46% in group B ($P < 0.003$) In the ELF, the median concentration was 12 mg/L in group A versus 6 mg/L in group B ($P < 0.08$) A threshold of 8 mg/L in the epithelial lining fluid was achieved twice as often in group A as in group B	Continuous infusion presents advantages in terms of PD and predictable efficacy in patients presenting ventilator-associated pneumonia
Burkhardt et al. [163]	Determine in vivo penetration into LT, ELF, and AC after 1 g of etrapenem (infusion period 30 min) for perioperative prophylaxis	Etrapenem	Single-center, prospective, observational of 15 patients undergoing thoracotomy BAL performed at 1, 3, and 5 h after etrapenem infusion Plasma concentration collected up to 24 h after infusion LT collected at time of lung extraction	Mean concentrations in plasma, ELF, and AC were: at 1.0 h, 63.1, 4.06, 0.004 mg/L; at 3.0 h, 39.7, 2.59, 0.003 mg/L; at 5.0 h, 27.2, 2.83, 0.007 mg/L Mean (range) concentration in LT was 7.60 (2.5–19.4) mg/kg tissue, 1.5 to 4.5 h after infusion and LT plasma concentration ratio was $23.6 \pm 12.3\%$ Mean level of ELF penetration was $7.48\% \pm 8.17\%$, and the highest degree of penetration was recorded at 5 h after infusion (mean \pm SD, $9.40\% \pm 10.7\%$) A 1-g dose, once daily, results in drug plasma concentrations higher than MICs of most community-acquired respiratory pathogens during the entire dosing interval Mean concentrations in ELF exceeded MIC90 values of highly or moderately penicillin-sensitive <i>Pneumococci</i> and of <i>H. influenzae</i> within 1 to 5 h after start of infusion and were partly below the MIC90 of penicillin-resistant <i>Pneumococci</i>	These results, combined with the reported MIC ₉₀ of most CAP bacteria, support the previously observed clinical efficacy of etrapenem in the treatment of community-acquired pneumonia
Boselli et al. [164]	Determine the steady-state serum and ELF concentrations of unbound etrapenem administered once daily to critically ill patients with early-onset ventilator-associated pneumonia	Etrapenem	Prospective, open-label study in an intensive care unit 15 patients with VAP received 1-h intravenous infusions of 1 g etrapenem once daily Samples obtained at steady-state, after 2 days of therapy ELF obtained by mini-BAL	Median (interquartile range) C _{max} , C ₁₂ , and C _{min} concentrations (mg/l) 1, 12, and 24 h after the end of infusion were: 30.3 (27.1–37.8), 4.8 (3.9–6.4), and 0.8(0.5–1.2) in serum and 9.4 (8.0–10.7), 2.0 (1.1–2.5), and 0.3 (0.2–0.4) in ELF, respectively Median free percentage penetration in ELF approximately 30–40%	Concentrations exceeding the MIC90 values of most of the causative pathogens (0.25–2 mg/l for <i>S. pneumoniae</i> , 0.06–0.125 mg/l for <i>Haemophilus influenzae</i> , 0.25–0.5 mg/l for oxacillin-susceptible <i>S. aureus</i> , and 0.03–0.125 mg/l for <i>Enterobacteriaceae</i> and anaerobes 0.5–1 mg/l was encountered in early-onset VAP during 50–100% time 1 g intravenous etrapenem once daily should be effective for early-onset VAP ICU patients with no known risk factors for multidrug-resistant pathogens

Table 5 PK/PD studies of beta-lactams in bronchial-alveolar lavage (Continued)

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Boselli et al. [165]	To determine the steady-state plasma and ELF concentrations of ceftazidime administered in continuous infusion to critically ill patients with severe nosocomial pneumonia	Ceftazidime	Prospective, open-label study. 15 adult patients with severe nosocomial bacterial pneumonia on mechanical ventilation. Administration of 30 min infusion of 2 g ceftazidime followed by continuous infusion 4 g over 24 h. Blood and mini-BAL samples collected by the third day of antibiotic therapy (8:00 am, 12:00 pm, and 6:00 pm)	The mean \pm SD steady-state plasma and ELF concentrations in continuous infusion were 39.6 ± 15.2 g/mL and 8.2 ± 4.8 g/mL, respectively, showing a mean \pm SD percentage penetration of ceftazidime into ELF of $20.6 \pm 8.9\%$	The administration of the applied dose in critically ill patients with severe nosocomial pneumonia provides concentrations in excess of the MIC of many susceptible organisms over the course of therapy in both serum and ELF. However, for some pathogens such as <i>P. aeruginosa</i> , higher doses of ceftazidime should be administered, or another agent should be used in combination
Felton et al. [166]	To assess plasma and intra-pulmonary PK of piperacillin/tazobactam in critically ill patients. Quantify pulmonary penetration. Identify factors that may influence pulmonary penetration	Piperacillin/tazobactam	Prospective, open label, single arm study. 18 ICU adult patients with pneumonia every 8 h daily. Samples collected at steady-state (mean of 88 doses (range 2–16)). Non-directed bronchial lavage was performed for ELF sampling	Median piperacillin and tazobactam pulmonary penetration ratio was 49.3% and 121.2% respectively. ELF protein/plasma protein ratio, as a surrogate measure of lung permeability, was found to have a statistically negative correlation between piperacillin pulmonary penetration ratio and pulmonary permeability. There was no statistically significant correlation for tazobactam. 14–18% of patients will have suboptimal drug exposure (ELF 50% \geq MIC when infected with a "susceptible" (MIC < 16 mg/L) organism. Piperacillin and tazobactam plasma concentrations do not precisely predict ELF concentrations of both drugs	Current piperacillin-tazobactam regimens is inadequate for effective treatment and suppression of emergence of antimicrobial resistance in an unacceptably high proportion of critically ill patients, especially those with pneumonia resulting from infection with a less susceptible organism
Boselli et al. [167]	To determine the steady-state plasma and ELF concentrations of cefepime administered in continuous infusion in critically ill patients with severe bacterial pneumonia	Cefepime	Prospective, open-label study. 20 ICU adult patients with severe nosocomial bacterial pneumonia. All subjects received a 30-min intravenous infusion of cefepime 2 g followed by a continuous infusion of 4 g over 24 h. Samples were collected after 48 h of therapy. Blood and mini BAL samples were collected at 8:00 am, 12:00 pm, and 6:00 pm	Mean \pm SD steady-state plasma and ELF concentrations were 13.5 ± 3.3 g/mL and 14.1 ± 2.8 g/mL, respectively, with a mean percentage penetration into epithelial lining fluid of about 100%	Administration of cefepime in continuous infusion in critically ill patients with severe nosocomial pneumonia appears to optimize the pharmacodynamic profile of this beta-lactam by constantly providing concentrations in excess of MIC of most of the susceptible organisms over the course of therapy in both serum and ELF. For some pathogens such as <i>Acinetobacter</i> spp. and <i>P. aeruginosa</i> higher doses should be administered
Boselli et al. [168]	To determine the steady-state plasma and epithelial lining fluid concentrations of piperacillin/tazobactam administered to critically ill patients with severe bacterial pneumonia	Piperacillin/tazobactam	Prospective, open label study. 10 ICU adult patients with severe nosocomial pneumonia. 30-min intravenous infusion 4/0.5 g every 8 h. Samples were obtained at steady-state, after 2 days of treatment. Blood samples were collected at three predetermined time points at 7:00 am (trough), 8:00 am (peak), and 12:00 pm (intermediate). Mini BAL was performed simultaneously to blood sampling at 12:00 pm	Mean \pm SD steady-state plasma trough, peak and intermediate concentrations were 8.5 ± 4.6 μ g/mL, 55.9 ± 21.6 μ g/mL, and 24.0 ± 13.8 μ g/mL for piperacillin, and 2.1 ± 1.0 μ g/mL, 4.8 ± 2.1 μ g/mL, and 2.4 ± 1.2 μ g/mL for tazobactam. Mean \pm SD steady-state intermediate ELF concentrations were 13.6 ± 9.4 μ g/mL for piperacillin and 2.1 ± 1.1 μ g/mL for tazobactam, respectively. Mean percentage penetration of piperacillin and tazobactam into ELF was 56.8% and 91.3%, respectively	Treatment of severe nosocomial pneumonia with a regimen of P/T 4/0.5 g every 8 h might provide insufficient concentrations into lung tissue to exceed the MIC of many causative pathogens

Table 5 PK/PD studies of beta-lactams in bronchial-alveolar lavage (Continued)

Study	Endpoints	Antibiotic	Design	Results	Conclusions
Lodise et al. [169]	To describe the PD profile of cefditoren in plasma and ELF	Cefditoren	Open, noncontrolled, dual-center, phase I study 24 adult patients with scheduled bronchoscopy Single oral dose 400 mg cefditoren Three sampling time windows (1–2 h, 2–3 h, or 3–4 h post-administration)	Plasma/ELF concentrations (mg/L)/penetration ratios 1–2 h: $1.78 \pm 1.27/0.39 \pm 0.21/38.1 \pm 50.1\%$ 2–3 h: $1.33 \pm 0.95/0.34 \pm 0.25/23.2 \pm 18.1\%$ 3–4 h: $1.03 \pm 0.51/0.30 \pm 0.18/31.8 \pm 19.2\%$ AUC _{ELF} /AUC _{plasma} penetration ratio (mean \pm SD) was 0.33 ± 0.48 PTA in plasma FT > MIC of 60 to 70% was < 90% for MICs of > 0.03 mg/L PTA in ELF T > MIC of 60% and 70% were 90.26% and 86.65%, respectively, at a MIC of 0.0125 mg/L and were significantly less for higher MICs	Cefditoren penetrates reasonably well into the ELF, as defined by the mean AUC _{ELF} /AUC _{plasma} penetration ratio The overall probability of target attainment (T > MIC > 50%) in plasma and ELF, however, was suboptimal (< 90%)

AC alveolar cells, AUC area under the curve, BAL bronchoalveolar lavage, CAP community-acquired pneumonia, C_{max} maximal concentration, C₁₂ concentration at 12 h, C_{min} trough concentration, CFU colony forming unit, ELF epithelial lining fluid, ICU intensive care unit, LT lung tissue, MIC minimal inhibitory concentration, MIC₉₀ minimal inhibitory concentration for 90% of isolates, MRSA methicillin-resistant *Staphylococcus aureus*, P/T piperacillin/tazobactam, PTA probability of target attainment, SD standard deviation, T_{1/2} half-life, T > MIC percentage of time above minimal inhibitory concentration, VAP ventilator-associated pneumonia

MIC of ceftolozane achieved against both isolates was 2 and 0.5 $\mu\text{g/mL}$) was likely attained at the dose and frequency provided in this case [177].

Toxicity

Beta-lactams are generally considered to have a high safety window with relatively few adverse effects, even when high doses are used [15]. Neurotoxicity is the most reported serious adverse effect of beta-lactams. Benzylpenicillin, cefepime, ceftazidime, and imipenem are considered to be the high-risk beta-lactams for neurotoxicity. Renal impairment, excess doses and/or concentrations, age, and a prior history of neurological disorders are known to be predisposing factors [2, 178–184].

Other adverse effects are found in a few case reports: acute renal failure [185] and electrolyte disorders [186]; severe intravascular haemolysis [187, 188]; extreme thrombocytosis [189]; severe thrombocytopenia [190–193]; leukopenia [194]; delayed-type hypersensitivity [195]; anaphylactic shock [196]; and severe cutaneous reactions [197].

Therapeutic dose monitoring

Several studies reported high PK variability of beta-lactams in sepsis/septic shock, both in different patients and in the same patient over time. In critically ill patients, hydrophilic and moderately lipophilic antimicrobials, being at higher risk of daily PK variations, should be more closely monitored and their dosages should be streamlined according to the underlying diseases in order to prevent under- or overexposure [2, 11].

Therapeutic drug monitoring (TDM) has been instituted for aminoglycosides and glycopeptides to reduce the rate of toxicity. However, because of the safety profile of beta-lactams, TDM was thought unnecessary for these drugs. In line with PK changes in critically ill patients, insufficient PD target attainment with beta-lactams has been reported in these patients, especially those with hypoalbuminemia, altered renal function, and low susceptibility bacterial strain infections [2, 35, 42, 198]. The challenges in achieving 'optimal' drug concentrations in the critically ill suggest beta-lactam TDM as a useful strategy to optimize drug exposure [199].

The TDM approach could be particularly useful in a certain group of critically ill patients in whom achieving target concentrations is more difficult, such as those with highly resistant bacterial strains, obese patients, immunocompromised patients, those undergoing renal-replacement therapies, and patients with augmented renal clearance [2, 198, 200, 201].

Though there are PK models to estimate antibiotic concentrations over a range of creatinine clearance (CrCl) and on renal replacement therapy [67, 202–205], the use of CrCl as a tool to optimize beta-lactam dosing

may not be reliable; although CrCl was significantly correlated with concentrations and clearance of broad-spectrum beta-lactams, changes in CrCl and RRT parameters do not reliably predict variations in drug PK/PD. In this setting, routine TDM should be considered to adapt beta-lactam doses [206].

Daily TDM of beta-lactams with dose adaptation in critically ill patients improves PD target attainment [207, 208]. Case reports have shown that TDM improved clinical outcome [209], but the clinical efficacy of using drug levels to achieve adequate concentrations had never been properly evaluated [1, 35, 210, 211] and there are reports concerning cost-effectiveness [111].

Facing poor implementation in beta-lactam TDM, Delattre et al. [212] proposed a predictive PK performance between an aminoglycoside and a beta-lactam. Due to physicochemical and PK similarities between aminoglycosides and beta-lactams, optimization of the beta-lactam dosage could be reached without any beta-lactam measurements, using TDM-related data of an aminoglycoside. The study aimed to characterize the PK of four beta-lactams (piperacillin, ceftazidime, cefepime, and meropenem) at the first dose in 88 critically ill septic patients co-medicated with amikacin, and to confirm the predictive performance of amikacin data on these PK, on a larger patient cohort, using a nonlinear mixed-effects modeling approach. There was a significant relationship between the exposure to amikacin and to beta-lactams. The population model presented was able to guide dosage adjustments for piperacillin, ceftazidime, cefepime, and meropenem during the early phase of severe sepsis in critically ill patients, using renal biomarkers or TDM-related aminoglycoside data [212].

Conclusions

The duration of infusion of beta-lactams has been shown to influence their $fT > \text{MIC}$ and an improved PD profile of beta-lactams may be obtained by longer exposure with more frequent dosing, extended infusions, or continuous infusions. This is particularly relevant in the critically ill patient, as V_d and ClCr are often increased, namely in the early phase of systemic hyperinflammatory states, promoting the risk of antibiotic underdosing.

The use of extracorporeal support techniques, either for renal replacement or ECMO, may further contribute to this problem and consequently concentrations below those expected are often found for beta-lactams. Given the heterogeneity of extracorporeal support therapy modes, it is difficult to suggest a specific dosage, but we recommend not to reduce dosage since no drug accumulation was found in the available literature and to use continuous or prolonged infusions to achieve the adequate PD profiles necessary to successfully treat infections caused by less susceptible strains.

More studies are needed to define optimal dosing of new beta-lactams and new beta-lactam/beta-lactamase combinations, which are increasingly important to effectively treat multidrug-resistant bacterial strains, namely in patients on extracorporeal support therapy and with difficult-to-treat sites of infection.

Although, it is not currently a clinical routine in most hospitals and its clinical efficacy has not yet been properly evaluated, a beta-lactam TDM approach with daily dose adaptation, allowing personalized antibiotic dosing, should be particularly useful in critically ill patients in whom achieving target concentrations is more difficult, such as obese patients, the immunocompromised, patients with augmented renal clearance, those undergoing extracorporeal support therapy, or those infected with highly resistant bacterial strains. Studies comparing TDM- versus non-TDM-based beta-lactam regimens should be promoted.

However, infection usually occurs at extravascular sites and prediction of outcome based on antibiotic plasma concentrations may result in overestimation of antimicrobial activity at the site of infection. Very few studies have investigated PK/PD issues concerning special anatomic barriers like the brain and lung, but most suggest that standard ICU dosing for beta-lactams may be insufficient for low susceptibility/high MIC pathogens in these sites. Therefore, although no studies have assessed clinical outcome, we recommend using higher than standard dosing, preferably with continuous or prolonged infusions, when treating severe infections caused by less susceptible bacterial strains at these sites, as PD profiles may improve and toxicity does not seem to increase.

Abbreviations

AKI: Acute kidney injury; AUC: Area under the curve; C: Concentration; CFU: Colony-forming units; CI: Continuous infusion; Cl: Clearance; CICr: Creatinine clearance; CNS: Central nervous system; CRRT: Continuous renal replacement therapy; CSF: Cerebral spinal fluid; CVHD: Continuous venous-venous hemodialysis; CVHDF: Continuous venous-venous hemodiafiltration; CVHF: Continuous venous-venous hemofiltration; ECMO: Extracorporeal membrane oxygenation; EDD: Extended daily dialysis; ELF: Epithelial lining fluid; ICU: Intensive care unit; Ii: Intermittent infusion; KPC: Carbapenemase producing *Klebsiella*; MBC: Minimal bactericidal concentration; MIC: Minimum inhibitory concentration; PAE: Post-antibiotic effect; PD: Pharmacodynamics; PK: Pharmacokinetics; PTA: Probability of target attainment; RRT: Renal replacement therapy; SLED: Sustained low-efficiency dialysis; TDM: Therapeutic drug monitoring; VAP: Ventilator associated pneumonia; Vd: Volume of distribution

Availability of data and materials

All data generated or analyzed during this study are included in this published article (see bibliography).

Authors' contributions

RV and JAP made substantial contributions to conception and design, acquisition of data, and analysis and interpretation of data; were involved in drafting the manuscript or revising it critically for important intellectual content; gave final approval of the version to be published; agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Gonçalves-Pereira J, Póvoa P. Antibiotics in critically ill patients: a systematic review of the pharmacokinetics of β -lactams. *Crit Care*. 2011;15(5):R206.
- Sinnollareddy MJ, Roberts MS, Lipman J, Roberts JA. Beta-lactam pharmacokinetics and pharmacodynamics in critically ill patients and strategies for dose optimization: A structured review. *Clin Exp Pharmacol Physiol*. 2012;39:489–96.
- Puskarič MA, Trzeciak S, Shapiro NI, Arnold RC, Horton JM, Studnek JR, Kline JA, Jones AE, On behalf of the Emergency Medicine Shock Research Network. Association between timing of antibiotic administration and mortality from septic shock in patients treated with a quantitative resuscitation protocol. *Crit Care Med*. 2011;39:2066–71.
- Kumar A, Roberts D, Wood KE, Light B, Parrillo JE, Sharma S, Suppes R, Feinstein D, Zanotti S, Taiberg L, Gurka D, Kumar A, Cheang M. Duration of hypotension before initiation of effective antimicrobial therapy is the critical determinant of survival in human septic shock. *Crit Care Med*. 2006;34:1589–96.
- Roberts JA, Paul SK, Akova M, Bassetti M, De Waele JJ, Dimopoulos G, Kaukonen KM, Koulenti D, Martin C, Montravers P, Rello J, Rhodes A, Starr T, Wallis SC, Lipman J, for the DALI Study. DALI: defining antibiotic levels in intensive care unit patients: are current β -lactam antibiotic doses sufficient for critically ill patients? *Clin Infect Dis*. 2014; <https://doi.org/10.1093/cid/ciu027>.
- Zilberberg MD, Shorr AF, Micek ST, Vazquez-Guillamet C, Kollef MH. Multi-drug resistance, inappropriate initial antibiotic therapy and mortality in Gram-negative severe sepsis and septic shock: a retrospective cohort study. *Crit Care*. 2014;18:596.
- Bloos F, Ruddel H, Thomas-Ruddel D, Schwarzkopf D, Pausch C, Harbarth S, Schreiber T, Grundling M, Marshall J, Simon P, Levy MM, Weiss M, Weyland A, Gerlach H, Schurholz T, Engel C, Matthaus-Kramer C, Scheer C, Bach F, Riessen R, Poidinger B, Dey K, Weiler N, Meier-Helmann A, Haberle HH, Wobker G, Kaisers UX, Reinhart K. Effect of a multifaceted educational intervention for anti-infectious measures on sepsis mortality: a cluster randomized trial. *Intensive Care Med*. 2017;43:1602–12.
- Ryoo SM, Kim WY, Sohn CH, Seo DW, Koh JW, Oh BJ, Lim KS. Prognostic value of timing of antibiotic administration in patients with septic shock treated with early quantitative resuscitation in emergency departments. *Am J Med Sci*. 2015;349:328–33.
- Vilella AL, Seifert CF. Timing and appropriateness of initial antibiotic therapy in newly presenting septic patients. *Am J Emerg Med*. 2014;32:7–13.
- Rhodes A, Evans LE, Alhazzani W, Levy MM, Antonelli M, Ferrer R, Kumar A, Sevransky JE, Sprung CL, Nunnally ME, Rochwerf B, Rubenfeld GD, Angus DC, Annane D, Beale RJ, Bellinghan GJ, Bernard GR, Chiche JD, Coopersmith C, De Backer DP, French CJ, Fujishima S, Gerlach H, Hidalgo JL, Hollenberg SM, Jones AE, Karnad DR, Kleinpell RM, Koh Y, Lisboa TC, Machado FR, Marini JJ, Marshall JC, Mazuski JE, McIntyre LA, McLean AS, Mehta S, Moreno RP, Myburgh J, Navalesi P, Nishida O, Osborn TM, Perner A, Plunkett CM, Ranieri M, Schorr CA, Seckel MA, Seymour CW, Shieh L, Shukri KA, Simpson SQ, Singer M, Thompson BT, Townsend SR, Van der Poll T, Vincent JL, Wiersinga WJ, Zimmerman JL, Dellinger RP. Surviving Sepsis Campaign:

- International guidelines for management of sepsis and septic shock: 2016. *Intensive Care Med.* 2017;43:304–77.
11. Bloos F, Rüdde H, Thomas-Rüddel D, Schwarzkopf D, Pausch C, Harbarth S, Schreiber T, Gründling M, Marshall J, Simon P, Levy MM, Weiss M, Weyland A, Gerlach H, Schürholz T, Engel C, Matth C, Scheer C, Bach F, Riessen R, Poidinger B, Dey K, Weiler N, Meier-Hellmann A, Häberle HH, Wöbker G, Kaisers UX, Reinhart K for the MEDUSA study group. Effect of a multifaceted educational intervention for anti-infectious measures on sepsis mortality: a cluster randomized trial. *Intensive Care Med.* 2017;43:1602–12.
 12. Pea F, Viale P, Furlanum T. Antimicrobial therapy in critically ill patients. A review of pathophysiological conditions responsible for altered disposition and pharmacokinetic variability. *Clin Pharmacokinet.* 2005;44:1009–34.
 13. Levison ME. Pharmacodynamics of antimicrobial drugs. *Infect Dis Clin N Am.* 2004;18:451–65.
 14. Roberts JA, Lipman J. Pharmacokinetic issues for antibiotics in the critically ill patient. *Crit Care Med.* 2009;37:840–51.
 15. McKinnon PS, Paladino JA, Schentag JJ. Evaluation of area under the inhibitory curve (AUC) and time above the minimum inhibitory concentration (T>MIC) as predictors of outcome for cefepime and ceftazidime in serious bacterial infections. *Int J Antimicrob Agents.* 2008;31:345–51.
 16. Roos JF, Lipman J, Kirkpatrick CMJ. Population pharmacokinetics and pharmacodynamics of ceftipime in critically ill patients against Gram-negative bacteria. *Intensive Care Med.* 2007;33:781–8.
 17. Sime BK, Roberts MS, Warner MS, Hahn U, Robertson TA, Yeend S, Phay A, Lehman S, Lipman J, Peake SL, Roberts JA. Altered pharmacokinetics of piperacillin in febrile neutropenic patients with haematological malignancy. *Antimicrob Agents Chemother.* 2014;53:533–7.
 18. Taccone FS, Cotton F, Vincent JL, Jacobs F. Optimal meropenem concentrations to treat multidrug-resistant *Pseudomonas aeruginosa* septic shock. *Antimicrob Agents Chemother.* 2012;56:2129–31.
 19. Ashley WS, Allen N, Rafferty KD, Fish DN, Toschlog E, Newell M, Waibel B. Pharmacokinetic analysis of piperacillin administered with tazobactam in critically ill, Morbidly Obese Surgical Patients. *Pharmacotherapy.* 2014;34:28–35.
 20. Onufrak NJ, Forrest A, Gonzalez D. Pharmacokinetic and pharmacodynamic principles of anti-infective dosing. *Clin Ther.* 2016;8:1930–47.
 21. Syamhanin A, Li JX, Wallis SC, Rudd M, Jarrett P, Paterson DL, Lipman J, Udy AA, Roberts JA. Pharmacokinetics of meropenem and piperacillin in critically ill patients with indwelling surgical drains. *Int J Antimicrob Agents.* 2013;42:90–3.
 22. Brink AJ, Richards JA, Schillack V, Kiem S, Schentag J. Pharmacokinetics of once-daily dosing of ertapenem in critically ill patients with severe sepsis. *Int J Antimicrob Agents.* 2009;33:432–6.
 23. Roberts JA, Udy AA, Jarret P, Wallis SC, Hope WW, Sharma R, Kirkpatrick CMJ, Kruger PS, Roberts MS, Lipman J. Plasma and target-site subcutaneous tissue population pharmacokinetics and dosing simulations of cefazolin in post-trauma critically ill patients. *J Antimicrob Chemother.* 2015;70:1495–502.
 24. Jeon S, Han S, Lee J, Hong T, Paek J, Woo H, Yima DS. Population pharmacokinetic analysis of piperacillin in burn patients. *Antimicrob Agents Chemother.* 2014;58:3744–51.
 25. Carlier M, Noe M, Roberts JA, Stove V, Verstraete AG, Lipman J, De Waele JJ. Population pharmacokinetics and dosing simulations of cefuroxime in critically ill patients: non-standard dosing approaches are required to achieve therapeutic exposures. *J Antimicrob Chemother.* 2014;69:2797–803.
 26. Couffignal C, Pajot O, Laouenan C, Burdet C, Foucrier A, Wolff M, Armand-Lefevre L, Mentré F, Massias L. Population pharmacokinetics of imipenem in critically ill patients with suspected ventilator-associated pneumonia and evaluation of dosage regimens. *Br J Clin Pharmacol.* 2014;78:1022–34.
 27. De Waele JJ, Lipman J, Akova M, Bassetti M, Dimopoulos G, Kaukonen M, Koukenti D, Martin C, Montravers P, Rello J, Rhodes A, Udy AA, Starr T, Wallis SC, Roberts JA. Risk factors for target non-attainment during empirical treatment with β -lactam antibiotics in critically ill patients. *Intensive Care Med.* 2014;40:1340–51.
 28. Meyer B, Traunmueller F, Bojic A, Locker G, Schmid R, Winkler S, Thalhammer F. Single-dose pharmacokinetics of cefodizime in critically ill elderly patients. *Int J Antimicrob Agents.* 2006;27:335–8.
 29. Blot SI, Pea F, Lipman J. The effect of pathophysiology on pharmacokinetics in the critically ill patient - Concepts appraised by the example of antimicrobial agents. *Adv Drug Deliv Rev.* 2014;77:3–11.
 30. Joynt JM, Lipman J, Gomersall CD, Young RJ, Wong ELY, Gin T. The pharmacokinetics of once-daily dosing of ceftriaxone in critically ill patients. *J Antimicrob Chemother.* 2001;47:421–9.
 31. Dalley AJ, Lipman J, Deans R, Vankatesh B, Rudd M, Roberts MS, Cross SE. Tissue accumulation of cephalothin in burns: a comparative study by microdialysis of subcutaneous interstitial fluid cephalothin concentrations in burn patients and healthy volunteers. *Antimicrob Agents Chemother.* 2009;53:210–5.
 32. Dalley AJ, Deans R, Lipman J, Venkatesh B, Rudd M, Roberts MS, Cross SE. Unbound cephalothin pharmacokinetics in adult burn patients are related to the elapsed time after injury. *Antimicrob Agents Chemother.* 2009;53:5303–5.
 33. Gonçalves-Pereira J, Silva NE, Mateus A, Pinho C, Póvoa P. Assessment of pharmacokinetic changes of meropenem during therapy in septic critically ill patients. *BMC Pharmacol Toxicol.* 2014;15:21.
 34. Conil JM, Georges B, Ravat F, Ruiz S, Seguin T, Metsu D, Fourcade O, Saivin S. Ceftazidime dosage recommendations in burn patients: from a population pharmacokinetic approach to clinical practice via Monte Carlo simulations. *Clin Ther.* 2013;35:1603–12.
 35. Isla A, Rodríguez-Gascón A, Trocóniz IF, Bueno L, Solinís MA, Maynar J, Sánchez-Izquierdo JA, Pedraz JL. Population pharmacokinetics of meropenem in critically ill patients undergoing continuous renal replacement therapy. *Clin Pharmacokinet.* 2008;47:173–80.
 36. Udy AA, Roberts JA, Lipman J. Clinical implications of antibiotic pharmacokinetic principles in the critically ill. *Intensive Care Med.* 2013;39:2070–82.
 37. Carlier M, Carrette S, Stove V, Verstraete AG. Does consistent piperacillin dosing result in consistent therapeutic concentrations in critically ill patients? A longitudinal study over an entire antibiotic course. *Int J Antimicrob Agents.* 2014;43:470–3.
 38. Burkhardt O. Ertapenem in critically ill patients with early-onset ventilator-associated pneumonia: pharmacokinetics with special consideration of free-drug concentration. *J Antimicrob Chemother.* 2007;59:277–84.
 39. Uldemolins M, Kumar V, Majcher-Peszynska J, Drewelow B, Derendorf H, Welte T. Flucloxacillin dosing in critically ill patients with hypoalbuminemia: special emphasis on unbound pharmacokinetics. *J Antimicrob Chemother.* 2010;65:1771–8.
 40. Taccone FS, Laterre PF, Dugernier T, Spapen H, Delattre I, Wittebole X, De Backer D, Layeux B, Wallemacq P, Vincent JL, Jacobs F. Insufficient β -lactam concentrations in the early phase of severe sepsis and septic shock. *Crit Care.* 2010;14:R126.
 41. Hanes SD, Wood C, Herring V, Croce MA, Fabian TC, Pritchard E, Boucher BA. Intermittent and continuous ceftazidime infusion for critically ill trauma patients. *Am J Surg.* 2000;179:436–40.
 42. Garot D, Respaud R, Lanotte P, Simon N, Mercier E, Ehrmann S, Perrotin D, Dequin PF, Le Guellec C. Population pharmacokinetics of ceftriaxone in critically ill septic patients: a reappraisal. *British J Clin Pharm.* 2011;72:758–67.
 43. Conil JM, Georges B, Mimoz O, Dieye E, Ruiz S, Cougot P, Samii K, Houin G, Saivin S. Influence of renal function on trough serum concentrations of piperacillin in intensive care unit patients. *Intensive Care Med.* 2006;32:2063–6.
 44. Kees MG, Minichmayr I, Moritz S, Beck S, Wicha SG, Kees F, Kloft C, Steinke T. Population pharmacokinetics of meropenem during continuous infusion in surgical ICU patients. *J Clin Pharmacol.* 2016;56:307–15.
 45. Ramon-Lopez A, Allen JM, Thompson AH, Dheansa BS, James SE, Hanlan GW, Davies JG. Dosing regimen of meropenem for adults with severe burns: a population pharmacokinetic study with Monte Carlo simulations. *J Antimicrob Chemother.* 2015;70:882–90.
 46. Blot S, Lipman J, Roberts DM, Roberts JA. The influence of acute kidney injury on antimicrobial dosing in critically ill patients: are dose reductions always necessary? *Diagn Microbiol Infect Dis.* 2014;79:77–84.
 47. Roberts JA, Pea F, Lipman J. *Clin Pharmacokinet.* 2013;52:1–8.
 48. Udy AA. Are standard doses of piperacillin sufficient for critically ill patients with augmented creatinine clearance? *Crit Care.* 2015;19:28.
 49. Udy AA, Lipman J, Jarret P, Klein K, Wallis SC, Patel K, Kirkpatrick CMJ, Kruger PS, Paterson DL, Roberts MS, Roberts JA. Augmented renal clearance in the intensive care unit: an illustrative case series. *Int J Antimicrob Agents.* 2010;35:606–8.
 50. Huttner A, Von Dach E, Renzoni A, Huttner B, Affaticati M, Pagani L, Daali Y, Pugin J, Karmime A, Fathi M, Lew D, Harbarth S. Augmented renal clearance, low β -lactam concentrations and clinical outcomes in the critically ill: An observational prospective cohort study. *Int J Antimicrob Agents.* 2015;45:385–39.
 51. Carlier M, Carrette S, Roberts JA, Stove V, Verstraete A, Hoste E, Depuydt P, Decruyenaere J, Lipman J, Wallis SC, De Waele JJ. Meropenem and piperacillin/tazobactam prescribing in critically ill patients: does augmented

- renal clearance affect pharmacokinetic/pharmacodynamic target attainment when extended infusions are used? *Crit Care*. 2013;17:R84.
52. Kikuchi E, Kikuchi J, Nasuhara Y, Oizumi S, Ishizaka A, Nishimura M. Comparison of the pharmacodynamics of biapenem in bronchial epithelial lining fluid in healthy volunteers given half-hour and three-hour intravenous infusions. *Antimicrob Agents Chemother*. 2009;53:2799–803.
 53. Mimos O, Soreda S, Padoin C, Tod M, Petitjean O, Benhamou D. Ceftriaxone pharmacokinetics during iatrogenic hydroxyethyl starch-induced hypoalbuminemia. *Anesthesiology*. 2000;93:735–43.
 54. Udy AA, Varghese JM, Altukroni M, Briscoe S, McWhinney BC, Ungerer JP, Lipman J, Roberts JA. Subtherapeutic initial b-lactam concentrations in select critically ill patients. Association between augmented renal clearance and low trough drug concentrations. *Chest*. 2012;142:30–9.
 55. Wong G, Briscoe S, Adnan S, McWhinney B, Ungerer J, Lipman J, Roberts JA. Protein binding of beta-lactam antibiotics in critically ill patients: can we successfully predict unbound concentrations? *Antimicrob Agents Chemother*. 2013;57:6165–70.
 56. Schleibinger M, Steinbach C, Topper C, Kratzer A, Liebchen U, Kees F, Salzberger B, Kees MG. Protein binding characteristics and pharmacokinetics of ceftriaxone in intensive care unit patients. *Br J Clin Pharmacol*. 2015;80:525–33.
 57. Matuszkiewicz-Rowińska J. Dosing of antibiotics in critically ill patients: are we left to wander in the dark? *Polskie Archiwum Medycyny Wewnętrznej*. 2012;122:630–40.
 58. Fissell WH. Antimicrobial dosing in acute renal replacement. *Adv Chronic Kidn Dis*. 2013; <https://doi.org/10.1053/jackd.2012.10.004>.
 59. Eyler RF, Mueller BA. Antimicrobial dosing in critically ill patients with acute kidney injury. *Nat Rev Nephrol*. 2011;7:226–35.
 60. Lewis SJ, Mueller BA. Antibiotic dosing in critically ill patients receiving CRRT: underdosing is overprevalent. *Semin Dial*. 2014;27:441–5.
 61. Harris LE, Reaves AB, Kraussa AG, Grinerb J, Hudson JQ. Evaluation of antibiotic prescribing patterns in patients receiving sustained low-efficiency dialysis: opportunities for pharmacists. *Int J Pharm Pract*. 2013;21:55–61.
 62. Morabito S, Guzzo I, Vitaliano E, Muzi L, Solazzo A, Pistolesi V, Pierucci A. Farmacocinetica degli Antibiotici nelle Terapie Sostitutive Renali Continue (CRRT). *Gionale Italiano di Nefrologia*. 2012;29:425–44.
 63. Morabito S, Guzzo I, Vitaliano E, Muzi L, Solazzo A, Pistolesi V, Pierucci A. Principi di farmacocinetica e aggiustamento posologico dei farmaci nelle terapie sostitutive renali continue (CRRT). *Giornale Italiano di Nefrologia*. 2006;36:5127–38.
 64. Choi G, Gomersall CD, Tian Q, Joynt GM, Freebairn R, Lipman J. Principles of antibacterial dosing in continuous renal replacement therapy. *Crit Care Med*. 2009;37:2268–82.
 65. Hites M, Dell'Anna AM, Scolletta S, Taccone FS. The challenges of multiple organ dysfunction syndrome and extra-corporeal circuits for drug delivery in critically ill patients. *Adv Drug Deliv Rev*. 2014;77:12–21.
 66. Jamal J-A, Udy AA, Lipman J, Roberts JA. The impact of variation in renal replacement therapy settings on piperacillin, meropenem, and vancomycin drug clearance in the critically ill: an analysis of published literature and dosing regimens. *Crit Care Med*. 2014;42:1640–50.
 67. Fish DN, Teitelbaum I, Abraham E. Pharmacokinetics and pharmacodynamics of imipenem during continuous renal replacement therapy in critically ill patients. *Antimicrob Agents Chemother*. 2005;49:2421–8.
 68. Malone RS, Fish DN, Abraham E, Teitelbaum I. Pharmacokinetics of cefepime during continuous renal replacement therapy in critically ill patients. *Antimicrob Agents Chemother*. 2001;45:3148–55.
 69. Mueller SC, Majcher-Peszynska J, Hickstein H, Francke A, Pertschy A, Schulz A, Mundkowski R, Drewelow B. Pharmacokinetics of piperacillin-tazobactam in anuric intensive care patients during continuous venovenous hemodialysis. *Antimicrob Agents Chemother*. 2002;46:1557–60.
 70. Roberts DM, Roberts JA, Roberts MS, Liu X, Nair P, Cole L, Lipman J, Bellomo R, on behalf of the RENAL Replacement Therapy Study Investigators. Variability of antibiotic concentrations in critically ill patients receiving continuous renal replacement therapy: A multicentre pharmacokinetic study. *Crit Care Med*. 2012;40:1523–8.
 71. Banyai M, Thalhammer F, El Menyawi I, Heinz G, Traunmüller F, Siostrzonek P. Pharmacokinetics of ceftiprome during continuous venovenous hemofiltration: Rationale for an 8-hour dosing interval. *Clin Pharmacol Therapeut*. 2000;67:368–72.
 72. Eyler RF, Vilay AM, Nader AM, Heung M, Pleva M, Sowinski KM, DePestel DD, Sörgel F, Kinzig M, Mueller BA. Pharmacokinetics of ertapenem in critically ill patients receiving continuous venovenous hemodialysis or hemodiafiltration. *Antimicrob Agents Chemother*. 2014;58:1320–6.
 73. Vossen MG, Wenisch JM, Maier-Salamon A, Fritsch A, Saria K, Zuba C, Jilch S, Lemmerer R, Unger M, Jaehde U, Jäger W, Thalhammer F. Doripenem treatment during continuous renal replacement therapy. *Antimicrob Agents Chemother*. 2016;60:1687–94.
 74. Carlier M, Taccone FS, Beumier M, Seyler L, Cottone F, Jacobs F, Roberts JA. Population pharmacokinetics and dosing simulations of cefepime in septic shock patients receiving continuous renal replacement therapy. *Int J Antimicrob Agents*. 2015;46:413–9.
 75. Seyler L, Cotton F, Taccone FS, De Backer D, Macours P, Vincent J-L, Jacobs F. Recommended b-lactam regimens are inadequate in septic patients treated with continuous renal replacement therapy. *Crit Care*. 2011;15:R137.
 76. Roberts DM, Liu X, Roberts JA, Nair P, Cole L, Roberts MS, Lipman J, Bellomo R, on behalf of the RENAL Replacement Therapy Study Investigators. A multicenter study on the effect of continuous hemodiafiltration intensity on antibiotic pharmacokinetics. *Crit Care*. 2015;19:84.
 77. Ohchi T, Hidaka S, Goto K, Shitomi R, Nishida T, Abe T, Yamamoto S, Yasuda N, Hagiwara S, Noguchi T. Effect of hemopurification rate on doripenem pharmacokinetics in critically ill patients receiving high-flow continuous hemodiafiltration. *Yakugaku Zasshi*. 2011;131:1395–9.
 78. Arzuaga A, Maynar J, Gascón AR, Isla A, Corral E, Fonseca F, Sánchez-Izquierdo JA, Rello J, Canut A, Pedraz JL. Influence of renal function on the pharmacokinetics of piperacillin/tazobactam in intensive care unit patients during continuous venovenous hemofiltration. *J Clin Pharmacol*. 2005;45:168–76.
 79. Isla A, Maynar J, Sánchez-Izquierdo JA, Gascón AR, Arzuaga A, Corral E, Pedraz JL. Meropenem and continuous renal replacement therapy: in vitro permeability of 2 continuous renal replacement therapy membranes and influence of patient renal function on the pharmacokinetics in critically ill patients. *J Clin Pharmacol*. 2005;45:1294–304.
 80. Ulldemolins A, Soy D, Llaurodo-Serra M, Vaquer S, Castro P, Rodríguez AH, Pontes CJ, Calvo G, Torres A, Martín-Loeches I. Meropenem population pharmacokinetics in critically ill patients with septic shock and continuous renal replacement therapy: influence of residual diuresis on dose requirements. *Antimicrob Agents Chemother*. 2015;59:5520–8.
 81. Bouman CSC, van Kan HJM, Koopmans RP, Korevaar JC, Schultz MJ, Vroom MB. Discrepancies between observed and predicted continuous venovenous hemofiltration removal of antimicrobial agents in critically ill patients and the effects on dosing. *Intensive Care Med*. 2006;32:2013–9.
 82. Kielstein JT, Czock D, Schöpke T, Hafer C, Bode-Böger SM, Kuse E, Keller F, Fliser D. Pharmacokinetics and total elimination of meropenem and vancomycin in intensive care unit patients undergoing extended daily dialysis. *Crit Care Med*. 2006;34:51–6.
 83. Lorenzen JM, Broll M, Kaever V, Burhenne H, Hafer C, Clajus C, Knitsch W, Burkhardt O, Kielstein JT. Pharmacokinetics of ampicillin/sulbactam in critically ill patients with acute kidney injury undergoing extended dialysis. *Clin J Am Soc Nephrol*. 2012;7:385–90.
 84. Burkhardt O, Hafer C, Langhoff A, Kaever V, Kumar V, Welte T, Haller H, Fliser D, Kielstein JT. Pharmacokinetics of ertapenem in critically ill patients with acute renal failure undergoing extended daily dialysis. *Nephrol Dial Transplant*. 2009;24:267–71.
 85. Tamme K, Oselin K, Kipper K, Tasa T, Metsvaht T, Karjagin J, Herodes K, Kern H, Starkopf J. Pharmacokinetics and pharmacodynamics of piperacillin/tazobactam during high volume haemodiafiltration in patients with septic shock. *Acta Anaesthesiol Scand*. 2016;60:230–40.
 86. Bourquin V, Ponte B, Saudan P, Martin P-Y. Adaptation posologique des médicaments couramment utilisés en réanimation lors d'épuration extrarénale continue. *Néphrol Thérapeut*. 2009;5:533–41.
 87. Trotman RL, Williamson JC, Shoemaker DM, Salzer WL. Antibiotic dosing in critically ill adult patients receiving continuous renal replacement therapy. *Clin Infect Dis*. 2005;41:1159–66.
 88. Brett H, Matzke GR, Dager WE. Antimicrobial dosing concepts and recommendations for critically ill adult patients receiving continuous renal replacement therapy or intermittent hemodialysis. *Pharmacotherapy*. 2009;29:562–77.
 89. De Waele JJ, Carlier M. Beta-lactam antibiotic dosing during continuous renal replacement therapy: how can we optimize therapy? *Crit Care*. 2014;18:158.

90. Kielstein JT, Burkhardt O. Dosing of antibiotics in critically ill patients undergoing renal replacement therapy. *Curr Pharm Biotechnol*. 2011;12:2015–9.
91. Churchwell MD, Mueller BA. Drug dosing during continuous renal replacement therapy. *Semin Dial*. 2009;22:185–8.
92. Vossen MF, Thalhammer F. Effects of renal replacement therapy on antimicrobial therapy. *Curr Clin Pharmacol*. 2013;8:39–45.
93. Goldstein SL, Nolin TD. Lack of drug dosing guidelines for critically ill patients receiving continuous renal replacement therapy. *Clin Pharmacol Therapeut*. 2014;96:159–61.
94. Scoville BA, Mueller BA. Medication dosing in critically ill patients with acute kidney injury treated with renal replacement therapy. *Am J Kidney Dis*. 2013;61:490–500.
95. Mueller BA, Pasko DA, Sowinski KM. Higher renal replacement therapy dose delivery influences on drug therapy. *Artif Organs*. 2003;27:808–14.
96. Ulldemolins M, Vaquer S, Llauredó-Serra M, Pontes C, Calvo G, Soy D, Martín-Loeches I. Beta-lactam dosing in critically ill patients with septic shock and continuous renal replacement therapy. *Crit Care*. 2014;18:227.
97. Shekar, Fraser JF, Smith MT, Roberts JA. Pharmacokinetic changes in patients receiving extracorporeal membrane oxygenation. doi: <https://doi.org/10.1016/j.jcrrc.2012.02.013>.
98. Shekar K, Roberts JA, McDonald CI, Fisque S, Barnett AG, Mullany DV, Ghassabian S, Wallis SC, Fung YL, Smith MT, Fraser JF. Sequestration of drugs in the circuit may lead to therapeutic failure during extracorporeal membrane oxygenation. *Crit Care*. 2012;16:R194.
99. Dzierba AL, Abrams D, Brodie D. Medicating patients during extracorporeal membrane oxygenation: the evidence is building. *Crit Care*. 2017;21:66.
100. Donadello K, Antonucci E, Cristallini S, Roberts JA, Beumier M, Scolletta S, Jacobs F, Rondelet B, de Backer D, Vincent J-L, Taccone FS. β -Lactam pharmacokinetics during extracorporeal membrane oxygenation therapy: A case-control study. *Int J Antimicrob Agents*. 2015; 45: 278–282.
101. Shekar K, Fraser JF, Taccone FS, Welch S, Wallis SC, Mullany DV, Lipman J, Roberts JA, and on behalf of the ASAP ECMO Study Investigators. The combined effects of extracorporeal membrane oxygenation and renal replacement therapy on meropenem pharmacokinetics: a matched cohort study. *Crit Care*. 2014;18:565.
102. Welsh C, Augustin P, Allyn J, Massias L, Montravers P, Allou N. Alveolar and serum concentrations of imipenem in two lung transplant recipients supported with extracorporeal membrane oxygenation. *Transpl Infect Dis*. 2015;17:103–5.
103. Perrott J, Mabasa VH, Ensom MHH. Comparing outcomes of meropenem administration strategies based on pharmacokinetic and pharmacodynamic principles: a qualitative systematic review. *Ann Pharmacother*. 2010;44:557–64.
104. Mah GT, Mabasa VH, Chow I, Ensom MHH. Evaluating outcomes associated with alternative dosing strategies for piperacillin/tazobactam: a qualitative systematic review. *Ann Pharmacother*. 2012;46:265–75.
105. Burgess SV, Mabasa VH, Chow I, Ensom MHH. Evaluating outcomes of alternative dosing strategies for cefepime: a qualitative systematic review. *Ann Pharmacother*. 2015; <https://doi.org/10.1177/1060028014564179>.
106. Bauer KA, West JE, O'Brien JM, Goff DA. Extended-infusion cefepime reduces mortality in patients with *Pseudomonas aeruginosa* Infections. *Antimicrob Agents Chemother*. 2013;57:2907–12.
107. Roberts JA, Boots R, Rickard CM, Thomas P, Quinn J, Roberts DM, Richards B, Lipman J. Is continuous infusion ceftriaxone better than once-a-day dosing in intensive care? A randomized controlled pilot study. *J Antimicrob Chemother*. 2007;59:285–91.
108. Lorente L, Lorenzo L, Martín MM, Jiménez A, Mora ML. Meropenem by continuous versus intermittent infusion in ventilator-associated pneumonia due to Gram-negative bacilli. *Ann Pharmacother*. 2006;40:219–23.
109. Roberts JA, Kirkpatrick CMJ, Roberts MS, Robertson TA, Dalley AJ, Lipman J. Meropenem dosing in critically ill patients with sepsis and without renal dysfunction: intermittent bolus versus continuous administration? Monte Carlo dosing simulations and subcutaneous tissue distribution. *J Antimicrob Chemother*. 2009;64:142–50.
110. Rafati MR, Rouini MR, Mojtahedzadeh M, Najafi A, Tavakoli H, Gholami K, Fazeli MR. Clinical efficacy of continuous infusion of piperacillin compared with intermittent dosing in septic critically ill patients. *Int J Antimicrob Agents*. 2006;28:122–7.
111. Duszynska W, Taccone FS, Switala M, Hurkacz M, Kowalska-Krochmal B, Kübler A. Continuous infusion of piperacillin/tazobactam in ventilator-associated pneumonia: a pilot study on efficacy and costs. *Int J Antimicrob Agents*. 2012;39:153–8.
112. Roberts JA, Kirkpatrick CMJ, Roberts MS, Dalley AJ, Lipman J. First-dose and steady-state population pharmacokinetics and pharmacodynamics of piperacillin by continuous or intermittent dosing in critically ill patients with sepsis. *J Antimicrob Agents*. 2010;35:156–63.
113. Chytra I, Stepan M, Benes J, Pelnar P, Zidkova A, Bergerova T, Pradl R, Kasal E. Clinical and microbiological efficacy of continuous versus intermittent application of meropenem in critically ill patients: a randomized open-label controlled trial. *Crit Care*. 2012;16:R113.
114. Falagas ME, Tansarli GS, Ikawa K, Vardakas KZ. Clinical outcomes with extended or continuous versus short-term intravenous infusion of carbapenems and piperacillin/tazobactam: a systematic review and meta-analysis. *Clin Infect Dis*. 2013;56:272–82.
115. Lorente L, Jimenez A, Palmero S, Jimenez JJ, Iribarren JL, Santana M, Martín MM, Mora ML. Comparison of clinical cure rates in adults with ventilator-associated pneumonia treated with intravenous ceftazidime administered by continuous or intermittent infusion: a retrospective, nonrandomized, open-label, historical chart review. *Clin Ther*. 2007;29:2433–9.
116. Dulhunty JM, Roberts JA, Davis JS, Webb SAR, Bellomo R, Gomersall C, Shirwadkar C, Eastwood GM, Myburgh J, Paterson DL, Lipman J. Continuous infusion of beta-lactam antibiotics in severe sepsis: a multicenter double-blind, Randomized Controlled Trial. *Clin Infect Dis*. 2013;56:236–44.
117. Gonçalves-Pereira J, Oliveira BS, Janeiro S, Estilita J, Monteiro C, Salgueiro A, Vieira A, Gouveia J, Paulino C, Bento L, Póvoa P. Continuous infusion of piperacillin/tazobactam in septic critically ill patients—a multicenter propensity matched analysis. *PLoS One*. 2012;7:e4984. <https://doi.org/10.1371/journal.pone.0049845>.
118. Abdul-Aziz MH, Staatz CE, Kirkpatrick CMJ, Lipman J, Roberts JA. Continuous infusion vs. bolus dosing: implications for beta-lactam antibiotics. *Minerva Anesthesiol*. 2012;78:94–10.
119. Nicolau DP, McNabb J, Lacy MK, Quintiliani R, Nightingale CH. Continuous versus intermittent administration of ceftazidime in intensive care unit patients with nosocomial pneumonia. *Int J Antimicrob Agents*. 2001;17: 497–504.
120. Huang H, Hang S, Zhu P, Xi X. Continuous versus intermittent infusion of cefepime in neurosurgical patients with post-operative intracranial infections. *Int J Antimicrob Agents*. 2014;43:68–72.
121. van Zanten ARH, Oudijk M, Nohlmans-Paulssen MKE, van der Meer YG, Girbes ARJ, Polderman KH. Continuous vs. intermittent cefotaxime administration in patients with chronic obstructive pulmonary disease and respiratory tract infections: pharmacokinetics/pharmacodynamics, bacterial susceptibility and clinical efficacy. *Br J Clin Pharmacol*. 2006;63:100–9.
122. Lee GC, Liou H, Yee R, Quan CF, Neldner K. Outcomes of extended-infusion piperacillin-tazobactam: a retrospective analysis of critically ill patients. *Clin Ther*. 2012;34:2297–300.
123. Jaruratanasirikul S, Limapichat T, Jullangkoon M, Ainlang N, Ingviya N, Wongpoowarak W. Pharmacodynamics of meropenem in critically ill patients with febrile neutropenia and bacteraemia. *Int J Antimicrob Agents*. 2011;38:231–6.
124. Angus BJ, Smith MD, Suputtamongkol Y, Mattie H, Walsh AL, Wuthiekanun V, Chaowagul W, White NJ. Pharmacokinetic-pharmacodynamic evaluation of ceftazidime continuous infusion vs intermittent bolus injection in septicemic melioidosis. *Br J Clin Pharmacol*. 2000;49:184–91.
125. Roos JF, Bulitta J, Lipman J, Kirkpatrick CMJ. Pharmacokinetic-pharmacodynamic rationale for cefepime dosing regimens in intensive care units. *J Antimicrob Chemother*. 2006;58:987–93.
126. Breilh D, Fleureau C, Gordien J-B, Joanes-Boyau O, Texier-Maugein J, Rapaport S, Boselli E, Janvier G, Saux M-C. Pharmacokinetics of free eropenem in critically ill septic patients: intermittent versus continuous infusion. *Minerva Anesthesiol*. 2011;77:1058–62.
127. Roberts JA, Roberts MS, Robertson TA, Daley AJ, Lipman J. Piperacillin penetration into tissue of critically ill patients with sepsis—Bolus versus continuous administration? *Crit Care Med*. 2009;37:926–33.
128. Lodise TP, Lomaestro B, Drusano GL. Piperacillin-tazobactam for *Pseudomonas aeruginosa* infection: clinical implications of an extended-infusion dosing strategy. *Clin Infect Dis*. 2007;44:357–63.
129. Sakka SG, Glauner AK, Bulitta JB, Kinzig-Schippers M, Pfister W, Drusano GL, Sorgel F. Population pharmacokinetics and pharmacodynamics of continuous versus short-term infusion of imipenem-cilastatin in critically ill patients in a randomized, controlled trial. *Antimicrob Agents Chemother*. 2007;51:3304–10.
130. Nicasio AM, Ariano RE, Zelenitski SA, Kim A, Crandon JL, Kuti JL, Nicolau DP. Population pharmacokinetics of high-dose, prolonged-infusion cefepime in

- adult critically ill patients with ventilator-associated pneumonia. *Antimicrob Agents Chemother.* 2009;53:1476–81.
131. Arnold HM, Hollands JM, Skrupky LP, Smith JR, Juang PH, Hampton NB, McCormick S, Reichley RM, Hoban A, Hoffman J, Micek ST, Kollef MH. Prolonged infusion antibiotics for suspected Gram-negative infections in the ICU: a before-after study. *Ann Pharmacother.* 2013;47:170–80.
 132. Yusuf E, Spapen H, Piérard D. Prolonged vs. intermittent infusion of piperacillin/tazobactam in critically ill patients: A narrative and systematic review. *J Crit Care.* 2014;29:1089–95.
 133. Cheatham SC, Fleming MR, Healy DP, Kyoung E, Shea KM, Humphrey ML, Kays MB. Steady-state pharmacokinetics and pharmacodynamics of meropenem in morbidly obese patients hospitalized in an intensive care unit. *J Clin Pharmacol.* 2013;54:324–30.
 134. Laterre P-F, Wittebole X, Van de Velde S, Muller AE, Mouton JW, Carryn S, Tulkens PM, Dugernir T. Temocillin (6 g daily) in critically ill patients: continuous infusion versus three times daily administration. *J Antimicrob Chemother.* 2015;70:891–8.
 135. Langgartner J, Lehn N, Gluck T, Herzog H, Kees F. Comparison of the pharmacokinetics of piperacillin and sulbactam during intermittent and continuous intravenous infusion. *Chemotherapy.* 2007;53:370–7.
 136. Langgartner J, Vasold A, Glück T, Reng M, Kees F. Pharmacokinetics of meropenem during intermittent and continuous intravenous application in patients treated by continuous renal replacement therapy. *Int Care Med.* 2008; 34:1091–96.
 137. Langgartner J, Vasold A, Glück T, Reng M, Kees F. Pharmacokinetics of meropenem during intermittent and continuous intravenous application in patients treated by continuous renal replacement therapy. *Intens Care Med.* 2008;34:1091–6.
 138. Jamal J-A, Mat-Nor MB, Mohamad-Nor F-S, Udy AA, Wallis SC, Lipman J, Roberts JA. Pharmacokinetics of meropenem in critically ill patients receiving continuous venovenous haemofiltration: A randomised controlled trial of continuous infusion versus intermittent bolus administration. *Int J Antimicrob Agents.* 2015;45:41–5.
 139. Asín-Prieto E, Rodríguez-Gasco A, Tracóniz I, Soraluca A, Maynar J, Sánchez-Izquierdo JA, Isla A. Population pharmacokinetics of piperacillin and tazobactam in critically ill patients undergoing continuous renal replacement therapy: application to pharmacokinetic/pharmacodynamic analysis. *J Antimicrob Chemother.* 2014;69:180–9.
 140. Obrink-Hansen K, Juul RV, Storgaard M, Thomsen MK, Hardlei TF, Brock B, Kreilgaard M, Gjedsted J. Population pharmacokinetics of piperacillin in the early phase of septic shock: does standard dosing result in therapeutic plasma concentrations? *Antimicrob Agents Chemother.* 2015;59:7018–26.
 141. D'Agostino C, Rhodes NJ, Skoglund E, Roberts JA. Microbiologic clearance following transition from standard infusion piperacillin-tazobactam to extended-infusion for persistent Gram-negative bacteremia and possible endocarditis: A case report and review of the literature. *J Infect Chemother.* 2015;21:742–6.
 142. Lips M, Siller M, Strojil J, Urbánek K, Balik M, Suchánková H. Pharmacokinetics of imipenem in critically ill patients during empirical treatment of nosocomial pneumonia: A comparison of 0.5-h and 3-h infusions. *Int J Antimicrob Agents.* 2014;44:358–62.
 143. De Waele J, Carlier M, Hoste E, Depuydt P, Decruyenaere J, Wallis SC, Lipman J, Roberts JA. Extended versus bolus infusion of meropenem and piperacillin: a pharmacokinetic analysis. *Minerva Anesthesiol.* 2014;80:1302–9.
 144. Dulhunty JM, Roberts JA, Davis JS, SAR W, Bellomo R, Gomersall C, Shirwadkar C, Eastwood GM, Myburgh J, Paterson DL, Starr T, Paul SK, Lipman J, for the BLING II Investigators for the ANZICS Clinical Trials Group. A multicenter randomized trial of continuous versus intermittent b-lactam infusion in severe sepsis. *Am J Respir Crit Med.* 2015;192:1298–305.
 145. Abdul-Aziz MH, Lipman J, Akova M, Bassetti M, De Waele JJ, Dimopoulos G, Dulhunty J, Kaukonen K-M, Kouletis D, Martin C, Montravers P, Rello J, Rhodes A, Starr T, Wallis SC, Roberts JA, on behalf of the DALI Study Group. Is prolonged infusion of piperacillin/tazobactam and meropenem in critically ill patients associated with improved pharmacokinetic/pharmacodynamic and patient outcomes? An observation from the Defining Antibiotic Levels in Intensive Care unit patients (DALI) cohort. *J Antimicrob Chemother.* 2016;71:196–207.
 146. Boyadjiev I, Boulamery A, Simon N, Martin C, Bruguerolle B, Leone M. Penetration of ertapenem into muscle measured by in vivo microdialysis in mechanically ventilated patients. *Antimicrob Agents Chemother.* 2011;55:3573–5.
 147. Karjagin J, Lefeuvre S, Oselins K, Kipper K, Marchand S, Tikkerberi A, Starkopf J, Coue W, Sawchuk RJ. Pharmacokinetics of meropenem determined by microdialysis in the peritoneal fluid of patients with severe peritonitis associated with septic shock. *Clin Pharmacol Ther.* 2008;83:452–9.
 148. Zeitlinger MA, Erovic BM, Sauermaier R, Georgopoulos A, Muller M, Joukhadar C. Plasma concentrations might lead to overestimation of target site activity of piperacillin in patients with sepsis. *J Antimicrob Chemother.* 2005;56:703–8.
 149. Goldwater PN. Cefotaxime and ceftriaxone cerebrospinal fluid levels during treatment of bacterial meningitis in children. *Int J Antimicrob Agents.* 2005; 26:408–11.
 150. Do L, Udy AA, Roberts JA, Lipman J. Antibacterial therapeutic drug monitoring in cerebrospinal fluid: difficulty in achieving adequate drug concentrations. *J Neurosurg.* 2013;118:297–301.
 151. Abdul-Aziz MH, McDonald C, McWhinney B, Ungerer JPJ, Lipman J, Roberts JA. Low fluclxacillin concentrations in a patient with central nervous system infection: the need for plasma and cerebrospinal fluid drug monitoring in the ICU. *Ann Pharmacother.* 2014;48:1380–4.
 152. Cies JJ, Moore WS, Calaman S, Brown M, Narayan P, Parker J, Chopra A. Pharmacokinetics of continuous-infusion meropenem for the treatment of *Serratia marcescens* ventriculitis in a pediatric patient. *Pharmacotherapy.* 2015;35:e32–6.
 153. Dahyot-Fizelier C, Frasca D, Grégoire N, Adier C, Mimoz O, Debaene B, Couet W, Marchand S. Microdialysis study of cefotaxime cerebral distribution in patients with acute brain injury. *Antimicrob Agents Chemother.* 2013;6:2738–42.
 154. Morita A, Kamei S, Minami M, Yoshida K, Kawabata S, Kuroda H, Suzuki Y, Araki N, Iwasaki Y, Kobayashi R, Hayashi N, Hirayama T, Ochiai J, Ueda M, Yamagishi Y, Niwa J, Shindo K, Fukushima Y, Takita T, Sato T, Sato S, Mikamo H, lwata S. Open-label study to evaluate the pharmacodynamics, clinical efficacy, and safety of meropenem for adult bacterial meningitis in Japan. *J Infect Chemother.* 2014;20:535–40.
 155. Tsumura R, Ikawa K, Morikawa N, Ikeda K, Shibukawa M, Iida K, Kurisu K. The pharmacokinetics and pharmacodynamics of meropenem in the cerebrospinal fluid of neurosurgical patients. *J Chemother.* 2008;20:615–21.
 156. Nicasio AM, Quintiliani R Jr, DeRyke CA, Kutl JL, Nicolau DP. Treatment of *Serratia marcescens* meningitis with prolonged infusion of meropenem. *Ann Pharmacother.* 2007;41:1077–81.
 157. Frasca D, Dahyot-Fizelier C, Couet W, Debaene B, Mimoz O, Marchand S. Brain microdialysis distribution study of cefotaxime in a patient with traumatic brain injury. *Br J Anesthesiol.* 2012;109:830–1.
 158. Wang Q, Wu Y, Chen B, Zhou J. Drug concentrations in the serum and cerebrospinal fluid of patients treated with cefoperazone/sulbactam after craniotomy. *BMC Anesthesiol.* 2015;15:33.
 159. Rodvold KA, Nicolau DP, Lodise TP, Khashab M, Noel GJ, Kahn JB, Gotfried M, Murray SA, Nicholson S, Laohavaleeson S, Tessier PR, Drusano GL. Identifying exposure targets for treatment of staphylococcal pneumonia with ceftobiprole. *Antimicrob Agents Chemother.* 2009;53:3294–301.
 160. Bayat S, Louchahi K, Verdière B, Anglade D, Rahoui A, Sorin P-M, Tod M, Petitjean O, Fraisse F, Grimbert FA. Comparison of 99mTc-DTPA and urea for measuring cefepime concentrations in epithelial lining fluid. *Eur Respir J.* 2004;24:150–6.
 161. Boselli E, Breilh B, Rimmelé T, Guillaume C, Xuereb F, Saux M-C, Bouvet L, Chassard D, Allaouchiche B. Alveolar concentrations of piperacillin/tazobactam administered in continuous infusion to patients with ventilator-associated pneumonia. *Crit Care Med.* 2008;36:1500–6.
 162. Cousson J, Floch T, Guillard T, Vernet V, Raclot P, Wolak-Thierry A, Jolly D. Lung concentrations of ceftazidime administered by continuous versus intermittent infusion in patients with ventilator-associated pneumonia. *Antimicrob Agents Chemother.* 2015;59:1905–9.
 163. Burkhardt O, Majcher-Peszynska J, Borner K, Mundkowski R, Drewelow B, Derendorf H, Welte T. Penetration of ertapenem into different pulmonary compartments of patients undergoing lung surgery. *J Clin Pharmacol.* 2005; 45:659–65.
 164. Boselli E, Breilh D, Saux M-C, Gordien J-B, Allaouchiche B. Pharmacokinetics and lung concentrations of ertapenem in patients with ventilator-associated pneumonia. *Intensive Care Med.* 2006;32:2059–62.
 165. Boselli E, Breilh D, Rimmel T, Poupelin J-C, Saux M-C, Chassard D, Allaouchiche A. Plasma and lung concentrations of ceftazidime administered in continuous infusion to critically ill patients with severe nosocomial pneumonia. *Intensive Care Med.* 2004;30:989–91.

166. Felton TW, McCalman K, Malagon I, Ialska B, Whalley S, Goodwin J, Bentley AM, Hope WW. Pulmonary penetration of piperacillin and tazobactam in critically ill patients. *Clin Pharmacol Ther.* 2014;96:438–48.
167. Boselli, et al. Steady-state plasma and intrapulmonary concentrations of cefepime administered in continuous infusion in critically ill patients with severe nosocomial pneumonia. *Crit Care Med.* 2003;31(8):2102–6.
168. Boselli E, Breilh D, Cannesson M, Xuereb F, Rimmel T, Chassard D, Saux M-C, Allaouchiche B. Steady-state plasma and intrapulmonary concentrations of piperacillin/tazobactam 4 g/0.5 g administered to critically ill patients with severe nosocomial pneumonia. *Intens Care Med.* 2004;30:976–9.
169. Lodise TP, Kinzig-Schippers M, Drusano GL, Loos U, Vogel F, Bulitta J, Hinder M, Sorgel F. Use of population pharmacokinetic modeling and Monte Carlo simulation to describe the pharmacodynamic profile of cefditoren in plasma and epithelial lining fluid. *Antimicrob Agents Chemother.* 2008;52:1945–51.
170. Xiao AJ, Caro L, Popejoy MW, Huntigton JA, Kullar R. PK/PD target attainment with ceftolozane/tazobactam using Monte Carlo simulation in patients with various degrees of renal function, including augmented renal clearance and end-stage renal disease. *Infect Dis Ther.* 2017;6:137–48.
171. Chandorkar G, Xiao AJ, Mouksassi M-S, Hershberger E, Krishna G. Population pharmacokinetics of ceftolozane/tazobactam in healthy volunteers, subjects with varying degrees of renal function and patients with bacterial infections. *J Clin Pharmacol.* 2015;55:230–9.
172. Van Duin B, Bonomo RA. Ceftazidime/avibactam and ceftolozane/tazobactam: second-generation β -lactam/ β -lactamase inhibitor combinations. *Clin Infect Dis.* 2016;63:234–41.
173. Xiao AJ, Miller BW, Huntington JA, Nicolau DP. Ceftolozane/tazobactam pharmacokinetic/pharmacodynamic-derived dose justification for phase 3 studies in patients with nosocomial pneumonia. *J Clin Pharmacol.* 2016;56:56–66.
174. Veillette JJ, Truong J, Forland SC. Pharmacokinetics of ceftazidime-Avibactam in two patients with KPC-Producing *Klebsiella pneumoniae* Bacteremia and renal Impairment. *Pharmacotherapy.* doi.org/https://doi.org/10.1002/phar.1840.
175. Oliver WD, Heil EL, Gonzales JP, Mehrotra S, Robinett K, Saleeb P, Nicolau DP. Ceftolozane-tazobactam pharmacokinetics in a critically ill patient on continuous venovenous hemofiltration. *Antimicrob Agents Chemother.* 2016;60:1899–901.
176. Bremmer DN, Nicolau DP, Burcham P, Chunduri A, Shidham G, Bauer KA. Ceftolozane/tazobactam pharmacokinetics in a critically ill adult receiving continuous renal replacement therapy. *Pharmacotherapy.* Doi.org/https://doi.org/10.1002/phar.1744.
177. Stokem K, Zuckerman JB, Nicolau DP, Wungwattana M, Sears EH. Use of ceftolozane-tazobactam in a cystic fibrosis patient with multidrug-resistant pseudomonas infection and renal insufficiency. *Respir Med Case Rep.* 2018; 23:8–9.
178. Fugate JE, Kalimullah EA, Hocker SE, Clark SL, Wijidicks EFM, Rabinstein AA. Cefepime neurotoxicity in the intensive care unit: a cause of severe, underappreciated encephalopathy. *Crit Care.* 2013;17:R264.
179. Shaheen T, Volles D, Calland F, Sifri CD, Mytinger J, Hagspiel K, Sawyer R, Bonati H. Cefepime-associated status epilepticus in an ICU patient with renal failure. *J Chemother.* 2009;21:65–7.
180. Alvarez SD, Aragon MCFG, Moreno A, Alonso FP, Cook HJ, Cárdenas G, Soto-Hernández JL. Clinical and electroencephalographic assessment of cefepime during treatment of nosocomial infections in neurological patients. *Cent Nerv Syst Agents Med Chem.* 2011;11:1–5.
181. Finkelsztein A, Cabral L, Bragatti JÁ, da Silva AV, Schuh AFS. Imipenem-associated encephalopathy. *Arquivo Neuropsiquiatria.* 2010;68:137–9.
182. Smith NL, Freebairn RC, Park MAJ, Wallis SC, Roberts JA, Lipman J. Therapeutic drug monitoring when using cefepime in continuous renal replacement therapy: seizures associated with cefepime. *Crit Care Resusc.* 2012;14:312–5.
183. Collins RD, Tverdek FP, Bruno JJ, Coyle EA. Probable nonconvulsive status epilepticus with the use of high-dose continuous infusion ceftazidime. *J Pharm Pract.* 2016;29:564–8.
184. Beumier M, Casu GS, Hites M, Wolff F, Cotton F, Vincent J-L, Jacobs F, Taccone FS. Elevated β -lactam concentrations associated with neurological deterioration in ICU septic patients. *Minerva Anesthesiol.* 2015;81:497–506.
185. Fritz G, Barner C, Schindler R, Boemke W, Falke K. Amoxicillin-induced acute renal failure. *Nephrol Dial Transp.* 2003;18:1660–2.
186. Polderman KH, Girbes ARJ. Piperacillin-induced magnesium and potassium loss in intensive care unit patients. *Intensive Care Med.* 2002;28:520–2.
187. Lambden SP, Akeru J, Barrett NA. Acute intravascular hemolysis associated with intravenous administration of meropenem in a sixty-year-old man. *Can J Clin Pharmacol.* 2010;17:e64–6.
188. Marik PE, Parekh P. Life-threatening piperacillin-induced immune haemolysis in a patient with cystic fibrosis. *BMJ Case Rep.* 2013; <https://doi.org/10.1136/bcr-2012-007801>.
189. Yang C-J, Hwang J-J, Hung J-Y, Chong I-W, Huang M-S. Extreme thrombocytosis under the treatment by amoxicillin/clavulanate. *Pharm World Sci.* 2006;28:326–8.
190. Lim PP, Chong CP, Abdul AN. Cefepime-associated thrombocytopenia in a critically ill patient. *Int J Clin Pharmacol.* 2011;33:902–4.
191. Macwilliam JL, Mistry R, Floyd MS Jr, Baird AD. Piperacillin/tazobactam induced thrombocytopenia delayed response. *BMJ Case Rep.* 2012; <https://doi.org/10.1136/bcr.2012.5981>.
192. Rousan TA, Aldoss IT, Cowley BD Jr, Curtis BR, Bougie DW, Aster RH, George JN. Recurrent acute thrombocytopenia in the hospitalized patient: Sepsis, DIC, HIT, or antibiotic-induced thrombocytopenia. *Am J Hematol.* 2010;85:71–4.
193. Nguyen VD, Tourigny JF, Roy R, Brouillette D. Rapid-onset thrombocytopenia following piperacillin-tazobactam reexposure. *Pharmacotherapy.* 2015;35:e326–30.
194. Whitman CB, Joseph JM, Sjöholm LO. Cephalosporin-induced leukopenia following rechallenge with cefoxitin. *Ann Pharmacother.* 2008;42:1327–32.
195. Lambourne J, Kitchen J, Hughes C, Merry C. Piperacillin/tazobactam-induced paresthesiae. *Ann Pharmacother.* 2006;40:977–9.
196. Nag DS, Samaddar DP, Kant S, Mahanty PR. Perianesthetic refractory anaphylactic shock with cefuroxime in a patient with history of penicillin allergy on multiple antihypertensive medications. *Rev Brasil Anestesiol.* 2017; 67:217–20.
197. Lin Y-F, Yang C-H, Sindy H, Lin J-Y, Hui C-YR, Tsai Y-C, Wu TS, Huang CT, Kao K-C, Hu H-C, Chiu C-H, Hung S-I, Chung W-H. Severe cutaneous adverse reactions related to systemic antibiotics. *Clin Infect Dis.* 2014;58:1377–85.
198. Hayashi, et al. Beta-Lactam therapeutic drug monitoring in the critically ill: optimising drug exposure in patients with fluctuating renal function and hypoalbuminaemia. *Int J Antimicrob Agents.* 2013;41:162–6.
199. Hites M, Taccone FS, Wolff F, Cotton F, Beumier M, De Backer D, Roisin S, Lorent S, Surin R, Seyler L, Vincent J-L, Jacobs F. Case-control study of drug monitoring of beta-lactams in obese critically ill patients. *Antimicrob Agents Chemother.* 2013;57:708–15.
200. Blondiaux N, Wallet F, Favory R, Onimus T, Nseir S, Courcol RJ, Durocher A, Roussel-Delvallez M. Daily serum piperacillin monitoring is advisable in critically ill patients. 2010; <https://doi.org/10.1016/j.jantimicag.2010.01.018>.
201. Pea F, Cojutti P, Sbrojavacca R, Cadeo B, Cristini F, Bulfoni A, Furlanut M. TDM-guided therapy with daptomycin and meropenem in a morbidly obese, Critically Ill Patient. *Ann Pharmacother.* 2011;45:e37.
202. Crandon JL, Ariano RE, Zelenitsky SA, Nicasio AM, Kuti JL, Nicolau DP. Optimization of meropenem dosage in the critically ill population based on renal function. *Intensive Care Med.* 2011;37:632–8.
203. Robatel C, Decosterd LA, Biollaz J, Eckert P, Schaller MD, Buclin T. Pharmacokinetics and dosage adaptation of meropenem during continuous venovenous hemodiafiltration in critically ill patients. *J Clin Pharmacol.* 2003; 43:1329–40.
204. Kawano S, Matsumoto K, Hara R, Kuroda Y, Ikawa K, Morikawa N, Horino T, Hori S, Kizu J. Pharmacokinetics and dosing estimation of meropenem in Japanese patients receiving continuous veno-venous hemodialysis. *J Infect Chemother.* 2015;21:476–8.
205. Bugge JF. Pharmacokinetics and drug dosing adjustments during continuous venovenous hemofiltration or hemodiafiltration in critically ill patients. *Acta Anaesthesiol Scand.* 2001;45:929–34.
206. Casu GS, Hites M, Jacobs F, Cotton F, Wolff F, Beumier M, De Backer D, Vincent J-L, Taccone FS. Can changes in renal function predict variations in beta-lactam concentrations in septic patients? *Int J Antimicrob Agents.* 2013;42:422–8.
207. De Waele JJ, Carrette S, Carlier M, Stove V, Boelens J, Claeys G, Leroux-Roels I, Hoste E, Depuydt P, Decruyenaere J, Verstraete AG. Therapeutic drug monitoring-based dose optimisation of piperacillin and meropenem: a randomised controlled trial. *Intensive Care Med.* 2014;40:380–7.
208. Sime FB, Roberts MS, Tiong IS, Gardner JH, Lehman S, Peake SL, Hahn U, Warner MS, Roberts JA. Can therapeutic drug monitoring optimize exposure to piperacillin in febrile neutropenic patients with haematological malignancies? A randomized controlled trial. *J Antimicrob Chemother.* 2015; 70:2369–75.

209. Neuner EA, Ahrens CL, Groszek JJ, Isada C, Vogelbaum MA, Fissell WH, Bhimraj A. Use of therapeutic drug monitoring to treat *Elizabethkingia* meningoseptica meningitis and bacteraemia in an adult. *J Antimicrob Chemother.* 2012; <https://doi.org/10.1093/jac/dks053>.
210. Roberts JA, Ullidemolins M, Roberts MS, McWhinney B, Ungerer J, Paterson DL, Lipman J. Therapeutic drug monitoring of Beta-lactams in critically ill patients: proof of concept. *Int J Antimicrob Agents.* <https://doi.org/10.1016/j.ijantimicag.2010.06.008>.
211. Roberts JA, Norris R, Paterson DL, Martin JH. Therapeutic drug monitoring of antimicrobials. *Br J Clin Pharmacol.* 2011;73:27–36.
212. Delattre IK, Musuamba FT, Jacqmin P, Taccone FS, Laterre P-F, Verbeeck RK, Jacobs F, Wallemacq P. Population pharmacokinetics of four β -lactams in critically ill septic patients co-medicated with amikacin. *Clin Biochem.* 2012; 45:780–6.

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