Review

# Bench-to-bedside review: Rhabdomyolysis – an overview for clinicians

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See commentary, page 141 [http://ccforum.com/content/9/2/R141], and see related research by Naka *et al.* in this issue [http://ccforum.com/content/9/2/R90]

## Abstract

Rhabdomyolysis ranges from an asymptomatic illness with elevation in the creatine kinase level to a life-threatening condition associated with extreme elevations in creatine kinase, electrolyte imbalances, acute renal failure and disseminated intravascular coagulation. Muscular trauma is the most common cause of rhabdomyolysis. Less common causes include muscle enzyme deficiencies, electrolyte abnormalities, infectious causes, drugs, toxins and endocrinopathies. Weakness, myalgia and tea-colored urine are the main clinical manifestations. The most sensitive laboratory finding of muscle injury is an elevated plasma creatine kinase level. The management of patients with rhabdomyolysis includes early vigorous hydration.

## Introduction

Rhabdomyolysis means destruction or disintegration of striated muscle [1]. This syndrome is characterized by muscle breakdown and necrosis resulting in the leakage of the intracellular muscle constituents into the circulation and extracellular fluid [2]. Rhabdomyolysis ranges from an asymptomatic illness with elevation in the creatine kinase (CK) level to a life-threatening condition associated with extreme elevations in CK, electrolyte imbalances, acute renal failure (ARF) and disseminated intravascular coagulation.

The cause of rhabdomyolysis is usually easily identified; however, in some instances the etiology is elusive. Muscular trauma is the most common cause of rhabdomyolysis. Less common causes include muscle enzyme deficiencies, electrolyte abnormalities, infectious causes, drugs, toxins and endocrinopathies. Rhabdomyolysis is commonly associated with myoglobinuria, and if this is sufficiently severe it can result in ARF. Weakness, myalgia and tea-colored urine are the main clinical manifestations.

The most sensitive laboratory finding of muscle injury is an elevated CK level. In the absence of myocardial or brain infarction, CK >5000 U/l indicates serious muscle injury. The management of patients with rhabdomyolysis includes advanced life support (airway, breathing and circulation) followed by measures to preserve renal function – the latter includes vigorous hydration. The use of alkalizing agents and osmotic diuretics, while commonly used, remains of unproven benefit.

## Historical aspects

Rhabdomyolysis has been described for millennia. In the Bible a condition with characteristics similar to rhabdomyolysis is described when the Jews suffered a 'plague' during their exodus from Egypt, after abundant consumption of quail [3]. This biblical catastrophe is assumed to have been caused by intoxication with hemlock herbs that quails consume during spring migration [4].

Musculoskeletal trauma, in particular crush syndrome, accounts for a large proportion of the cases of rhabdomyolysis. The first cases of crush syndrome were reported in 1908 in the German military literature [5]. Crush victims who developed ARF were reported during the bombing of London during the Second World War. Pigmented casts were found in the renal tubules at autopsy; however, at that time the relationship between muscle injury and renal failure was unclear [5]. Additional cases were described during the Korean War [6]. The incidence of post-traumatic ARF decreased during the Vietnam War – this was ascribed to the faster evacuation techniques and improved fluid resuscitation of injured soldiers [7].



ARF = acute renal failure; CK = creatine kinase.

The role of myoglobin in the development of rhabdomyolysis was first described in experimental studies in the early 1940s. Bywaters and Stead injected rabbits with myoglobin and reported this 'toxin' to be responsible for the ARF following muscle injury [8]. In 1959, Korein and colleagues divided rhabdomyolysis into exertional and nonexertional groups [9]. In 1972, Rowland and Penn described a series of inherited enzyme deficiencies associated with myoglobinuria [10]. Increased recognition of nontraumatic, nonexertional causes of rhabdomyolysis soon occurred [11].

## Epidemiology

About 10–50% of patients with rhabdomyolysis develop ARF [12]. Indeed, it has been suggested by some authors that rhabdomyolysis from all causes leads to 5–25% of cases of ARF [11]. A recent clinical series of patients developing ARF reports mortality rates of 7–80% [13]. Rhabdomyolysis occurs in up to 85% of patients with traumatic injuries. Patients with severe injury who develop rhabdomyolysis-induced renal failure have a mortality of approximately 20% [14]. Mortality is higher in patients with multiorgan dysfunction syndrome [15].

Rhabdomyolysis and crush syndrome are common results of natural disasters such as earthquakes. The Marmara region of Turkey was devastated by one of the most catastrophic earthquakes recorded, registering a magnitude of 7.4 on the Richter Scale, on 17 August 1999 [16]. The Marmara region, a densely populated and highly industrialized area, is located in the northwestern part of Turkey with a population of 20 million. According to official reports, the disaster caused 17,480 deaths. Owing to the efforts of the Turkish Society of Nephrology and the International Society of Nephrology, detailed epidemiological data were collected [16]. Since almost all of the hospitals situated in the disaster area were partly or completely destroyed, victims were transferred by boat, helicopter or road to 35 reference hospitals located in adjacent cities. A total of 9843 patients were admitted to these reference hospitals, of whom 5392 were hospitalized and 425 died. Age was the only independent predictor of outcome. The average time under the rubble was 11.7 hours, which was not significantly different between survivors and nonsurvivors. Six hundred and thirty-nine patients developed renal failure (12% of all hospitalized patients), of whom 477 (74.6%) were treated by dialysis.

## Causes and pathophysiology

There are multiple causes of rhabdomyolysis, which can be classified as physical and nonphysical causes (see Table 1). The major causes of rhabdomyolysis in patients admitted to the emergency department of an urban population in the United States were reported to be cocaine, exercise and immobilization [17]. In the United States, rhabdomyolysis is commonly diagnosed in intoxicated patients subjected to prolonged muscle compression as they lay motionless, in elderly patients following a fall or stroke and in patients with

**Table 1**

**Causes of rhabdomyolysis**

| | |
|---|---|
| *Physical causes* | |
| Trauma and compression | Crush injuries |
| | Motor vehicle accidents |
| | Long-term confinement without changing position |
| | Physical torture and abuse |
| | Prolonged hours of surgery without changing position |
| Vessel occlusion | Embolism |
| | In *situ* thrombosis |
| | Vessel clamping during surgery |
| Shock states | |
| Strainful muscle exercise | Amphetamine overdose |
| Excessive muscle activity | Delirium tremens |
| | Epilepsy |
| | Overexertion (e.g. long distance running) |
| Tetanus | |
| Electrical current | Cardioversion |
| | High-voltage electrical injury |
| | Lightning |
| Hyperthermia | Exercise |
| | Malignant hyperthermia |
| | Neuroleptic malignant syndrome |
| Sepsis | |
| *Nonphysical causes* | |
| Metabolic syndromes | Carnitine deficiency |
| | Creatinine palmitoyl transferase deficiency |
| | McArdle disease (myophosphorylase deficiency) |
| | Mitochondrial respiratory chain enzyme deficiencies |
| | Phosphofruktokinase deficiency |
| Toxins | Heavy metals |
| | Insect venoms |
| | Snake venoms |
| Drugs | *See Table 2* |
| Infections | Coxsackievirus |
| | Falciparum malaria |
| | Herpes viruses |
| | HIV |
| | Legionella |
| | Salmonella |
| | Streptoccocus |
| | Tularemia |
| Electrolyte imbalances | Hyperosmotic conditions |
| | Hypernatremia |
| | Hypocalcemia |
| | Hyponatremia |
| | Hypokalemia |
| | Hypophosphatemia |
| Endocrine disorders | Hyperaldosteronism |
| | Hypothyroidism |
| | Ketoacidosis |
| | Hyperaldosteronism |
| Autoimmune diseases | Polymyositis |
| | Dermatomyositis |

seizure disorders [17]. Trauma and crush injuries following motor vehicle accidents and the collapse of buildings are other common causes of rhabdomyolysis [18,19]. During the collapse of the World Trade Center on 11 September 2001, nephrologists in New York City were prepared to dialyze large numbers of people with ARF in the days following the terrorist attack. Few patients were hospitalized with crush injuries, however, with only one reported case of rhabdomyolysis [20,21]. This case occurred in a 38-year-old policeman who was trapped under debris for 24 hours, who required hemodialysis for 1 month before fully recovering. Traumatic rhabdomyolysis may also occur in people who struggle against restraints and in children following abuse. Rhabdomyolysis has rarely been reported when a surgical procedure is performed in an improper position or following the prolonged use of a tourniquet [22–25].

Myoglobinemia and myoglobinuria and a mild elevation of creatine phosphokinase (CK) may occur after strenuous physical exertion [26]. When physical exertion is extreme, however, it can cause myolysis with severe rhabdomyolysis; this is especially likely to occur when strenuous exercise is performed under conditions of high temperature and humidity [27]. Hypokalemia increases the risk of rhabdomyolysis during strenuous exercise. This may be related to the fact that hypokalemia limits vasodilatation in the muscle microvasculature [28]. Athletes who abuse diuretics are therefore at a high risk of developing rhabdomyolysis during strenuous exercise. The pathogenesis of rhabdomyolysis following severe exertion appears to be due to a combination of mechanical and thermal muscle injury and ATP depletion. Excess muscle activity may also lead to rhabdomyolysis in conditions such as status epilepticus myoclonus and severe dystonia [29].

Rhabdomyolysis may complicate a high-voltage electrical injury and lightning strikes [30]. Rhabdomyolysis has been reported in 10% of subjects that survive an electrical shock. The degree of rhabdomyolysis is not related to the size of the wounds or to the site of entry [31]. The clinical course following an electrical burn is similar to that following a crush injury [32]. Myolysis following an electrical injury is attributed to the electrical disruption of sarcolemmal membranes, with loss of barrier function and massive calcium influx [33].

Hyperthemia may cause muscle damage. The syndromes of malignant hyperthermia and neuroleptic malignant syndrome are characterized by fever, generalized muscular contraction and rigidity, metabolic acidosis and rhabdomyolysis [34]. Malignant hyperthermia is an autosomal dominant genetic disorder in 50% of cases and an autosomal recessive genetic disorder in 20% of cases that affects males more frequently than females [35]. It occurs abruptly with the administration of anesthetic agents. The most common agents that cause malignant hyperthermia are succinylcholine and halothane [36]. The onset of malignant hyperthermia is usually within 1 hour of the administration of general anesthesia. Malignant hyperthermia results in excessive sweating, causing hypokalemia, which as previously stated potentiates the muscle injury [37].

Neuroleptic malignant syndrome is an idiosyncratic reaction to antipsychotic agents such as butyrophenones, phenothiazines and thioxanthenes, with haloperidol being the most common offending agent [38]. In this syndrome, there is a gradual development of hyperthermia, muscle rigidity, rhabdomyolysis, fluctuating consciousness and autonomic instability [39]. This clinical entity is believed to result from central nervous system dopamine receptor blockade, or from withdrawal of exogenous dopaminergic agonists [40]. Neuroleptic malignant syndrome can also develop in patients with Parkinson's disease following withdrawal of levodopa therapy [41–43].

Heat stroke is another cause of hyperthermia leading to rhabdomyolysis. By definition, patients with heat stroke have a core body temperature in excess of 40.5°C and their course is often complicated by acute respiratory distress syndrome, disseminated intravascular coagulation, renal or hepatic failure, rhabdomyolysis and seizures [44,45]. Heat stroke has a reported mortality approaching 21% [46]. Hypothermia can also cause rhabdomyolysis [47]. By reducing muscle perfusion, cold induces tissue ischemia and freezing causes cellular destruction [48,49].

Inherited disorders of carbohydrate metabolism can cause rhabdomyolysis [50]. McArdle's disease (myophosphorylase deficiency) is an autosomal recessive condition in which there is selective necrosis of type 2 muscle fibers [51]. These fibers are more dependent on glycolysis for generation of ATP and are therefore more sensitive to an enzyme defect that prevents the formation of glucose from glycogen. ATP depletion is responsible for rhabdomyolysis in this disease. Other diseases that affect the glycolytic/glycogenolytic pathways and cause rhabdomyolysis include Tarui's disease (congenital phosphofruktokinase deficiency) and phosphoglycerate mutase deficiency [52]. Other inherited metabolic disorders that are associated with rhabdomyolysis include carnitine palmitoyltransferase deficiency, an autosomal recessive disorder that has been considered the most common hereditary disease causing rhabdomyolysis [53]. In this deficiency disease, muscle pain and rhabdomyolysis develop after prolonged exercise with inadequate nutrient intake [54].

Medications and recreational drugs are important causes of rhabdomyolysis (see Table 2). Drug-induced rhabdomyolysis encompasses a large group of substances that can affect muscles by different mechanisms. Any drug that directly or indirectly impairs the production or use of ATP by skeletal muscle, or increases energy requirements that exceed the rate of ATP production, can cause rhabdomyolysis [55]. The potential mechanism of drug-induced sarcolemmal injury is presumably due to changes in the viscosity of sarcolemma

**Table 2**

**Drugs that may induce rhabdomyolysis**

| | |
|---|---|
| Antipsychotics and antidepressants | Drugs of addiction |
| Amitriptyline | Heroin |
| Amoxapine | Cocaine |
| Doxepine | Amphetamine |
| Fluoxetine | Methadone |
| Fluphenazine | D-lysergic acid diethylamide (LSD) |
| Haloperidol | Antihistamines |
| Lithium | Diphenhydramine |
| Protriptyline | Doxylamine |
| Phenelzine | Other drugs |
| Perphenazine | Alcohol |
| Promethazine | Amphotericin B |
| Chlorpromazine | Azathrioprine |
| Loxapine | Butyrophenones |
| Promazine | Emetics |
| Trifluoperazine | Epsilon-aminocaproic acid |
| Sedative hypnotics | Halothane |
| Benzodiazepines | Laxatives |
| Diazepam | Moxalactam |
| Nitrazepam | Narcotics |
| Flunitrazepam | Oxprenolol |
| Lorazepam | Paracetamol |
| Triazolam | Penicillamine |
| Barbiturates | Pentamidine |
| Gluthetimide | Phencyclidine |
| Antilipemic agents | Phenylpropanolamine |
| Lovastatin | Quinidine |
| Pravastatin | Salicylates |
| Simvastatin | Strychnine |
| Bezafibrate | Succinylcholine |
| Clozafibrate | Theophyline |
| Ciprofibate | Terbutaline |
| Clofibrate | Thiazides |
| | Vasopressin |

caused by activation of phospholipase A. These changes result in increased permeability of the sarcolemma, permitting leakage or intracellular contents, as well as an increase in the entry of sodium ions into the cell [56–58]. The increased intracellular sodium ion concentration activates $Na^+$,$K^+$-ATPase, a process that requires energy. This exhausts the supplies of ATP and impairs cellular transport proteins [59]. The increase in cellular sodium ion concentration leads to the accumulation of intracellular calcium, which activates neutral proteases causing further cellular injury [60].

The use of 3-hydroxy-3-methyl glutaryl coenzyme A reductase inhibitors, or statins, has been shown to reduce major cardiovascular events in both primary and secondary prevention. Statins have consequently become one of the most widely prescribed class of medications, with more than 76 million prescriptions filled in the United States in 2000 [61]. Statins are well tolerated by most patients. The most serious side effects of these drugs are myositis with rhabdomyolysis. This risk was emphasized by the withdrawal of cerivastatin in August 2001 after the drug was associated with approximately 100 rhabdomyolysis-related deaths [62]. Statins have been postulated to interfere with ATP production by reducing levels of coenzyme Q, a component of the electron transport chain [63]. Rhabdomyolysis may developed acutely soon after initiating therapy (2–3 weeks) or months or years later after a precipitating event such as an intercurrent illness or infection, strenuous exercise or a drug interaction. Clinically important rhabdomyolysis with statins is rare, with an overall reported incidence of fatal rhabdomyolysis of 0.15 deaths per one million prescriptions [64]. The FDA MedWatch Reporting system lists 3339 cases of statin-associated rhabdomyolysis reported between 1 January 1990 and 31 March 2002 [61].

Statins are also associated with a chronic myositis syndrome, characterized by muscle pain and weakness with or without evidence of clinically detectable rhabdomyolysis [65]. Few data are available on the frequency of the chronic myositis syndrome, which may affect between 0.1% and 1% of patients. Risk factors for the development of a statin-induced myopathy include high dosages, increasing age, female sex, renal and hepatic insufficiency, diabetes mellitus and concomitant therapy with drugs such as fibrates, cyclosporine, macrolide antibiotics, warfarin and digoxin [61]. Individual statins may differ in their risk of inducing rhabdomyolysis, with some patients developing this syndrome when switching from one statin to another. Other patients develop rhabdomyolysis when exposed to any statin. It is probable that genetic factors play a role in the pathogenesis of this syndrome.

Rhabdomyolysis has been reported in solid organ transplant recipients [66]. The use of immunosuppressive drugs, particularly cyclosporine, has been implicated in these cases. Alcohol directly injures the sarcolemma and increases sodium permeability [67]. Analysis of skeletal muscle from chronic alcoholics and experimental animals fed ethanol demonstrates a marked depletion of intracellular potassium, phosphorus and magnesium, and demonstrates elevated sodium, chloride, calcium and water content [68,69]. Acute alcohol-induced rhabdomyolysis can occur after binge drinking or a sustained period of alcohol abuse, and is associated with pain and swelling of muscles, particularly the quadriceps [70]. Drugs such as D-lysergic acid diethylamide (known as LSD), sympathomimetics and phencyclidine, which induce delirium or agitation, and those that cause prolonged involuntary muscle contraction, lead to increased ATP demand and to eventual exhaustion of its energy stores [71].

**Table 3**

**Mechanisms of cellular destruction in rhabdomyolysis**

| Method | Mechanism |
|---|---|
| Direct injury to cell membrane | Crushing, tearing, burning, pounding, poisoning, dissolving |
| Muscle cell hypoxia leading to depletion of ATP | Anerobic conditions: shock states, vascular occlusion, and tissue compression |
| Electrolyte disturbance disrupting the sodium–potassium pump | Hypokalemia: vomiting, diarrhea, extensive diuresis |
| | Hyponatremia: water intoxication |

Cocaine is a common cause of both traumatic and non-traumatic rhabdomyolysis. Twenty-four percent of emergency department patients presenting with cocaine-related disorders have acute rhabdomyolysis [72]. Cocaine produces rhabdomyolysis by several different mechanisms. Prolonged vasoconstriction of intramuscular arteries can produce muscle ischemia and acute rhabdomyolysis. In addition, large doses of cocaine can have a direct toxic effect and can produce acute skeletal myofibrillar degeneration. Cocaine may also produce traumatic rhabdomyolysis by causing generalized tonic–clonic seizures, or by coma and secondary physical compression of a major muscle group for prolonged periods of time [73].

A number of electrolyte abnormalities are associated with rhabdomyolysis [74]. Examples include chronic hypokalemia, hypophosphatemia and hyponatremia as well as rapid correction of hyponatremia [11,75,76]. Overuse of diuretic or cathartic drugs can lead to massive total body potassium depletion, causing rhabdomyolysis [77]. Potassium depletion-induced rhabdomyolysis can occur in the presence of normal or elevated serum potassium levels, which are maintained by the ongoing release of potassium from dying myocytes [28,31]. Any condition that produces major electrolyte losses, such as hyperemesis gravidarum, can be associated with rhabdomyolysis [78].

Polymyositis and dermatomyositis are chronic autoimmune conditions that in rare cases can progress to rhabdomyolysis [79,80]. An interesting and challenging cause of rhabdomyolysis is the ingestion of large quantities of licorice. It is well known that licorice contains a mineralocorticoid-type agent that causes renal potassium wasting [81]. Hyperosmolar states such as hyperglycemic hyperosmolar nonketotic coma have been reported to cause rhabdomyolysis [82,83]. On rare occasions rhabdomyolysis has been associated with thyroid storm and pheochromocytoma; both conditions increase sympathetic stimulation and metabolic demands, resulting in an extreme hypermetabolic state [84].

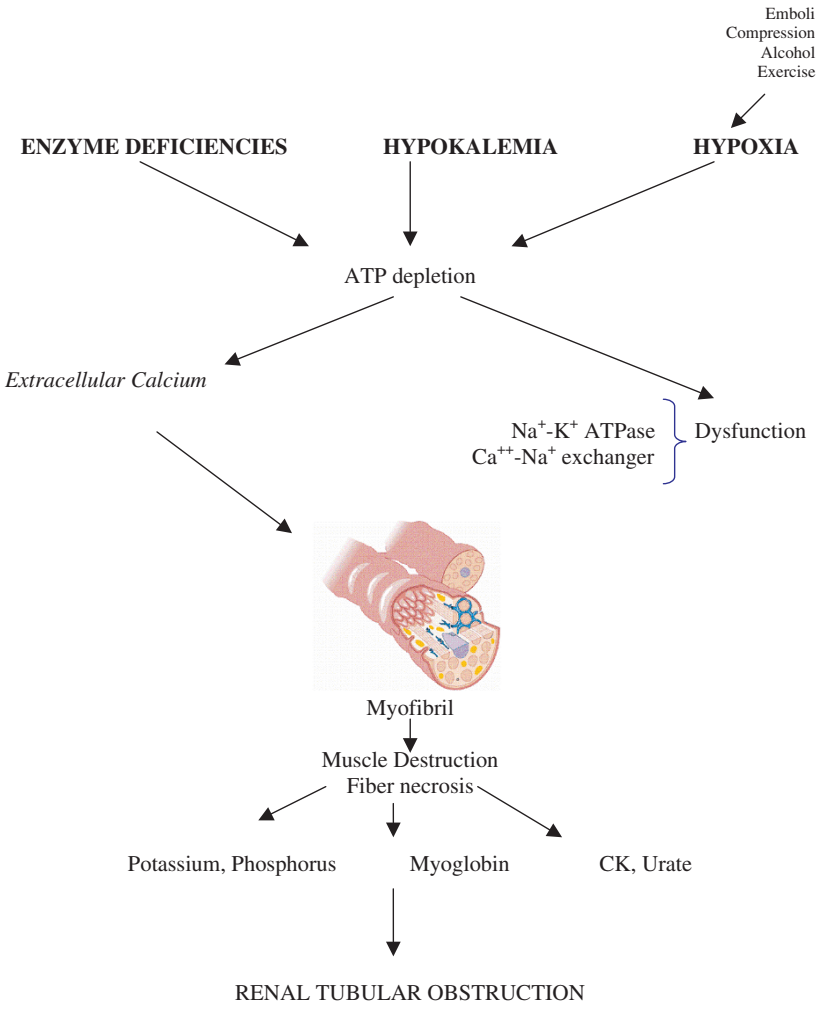
Infections have also been reported to cause rhabdomyolysis [85]. This includes bacterial pyomyositis, which presents with localized signs of muscle infection with erythema, edema and tenderness [86]. Legionella infection is classically associated with rhabdomyolysis [87]. Rhabdomyolysis can be seen in septic patients without direct muscle infection [88]. In these instances, muscle damage can be caused by a toxin, or from associated fever, rigors and dehydration [89]. Acute viral infections with influenza A and influenza B, Coxsackievirus, Epstein–Barr virus, herpes simplex virus, parainfluenza, adenovirus, echovirus, HIV and cytomegalovirus have been associated also with rhabdomyolysis [90,91].

Muscle injury, regardless of mechanism, results in a cascade of events that leads to leakage of extracellular calcium ions into the intracellular space [57] (Fig. 1 and Table 3). The excess of calcium causes a pathologic interaction of actin and myosin, and activates cellular proteases with muscle destruction and fiber necrosis [51]. The final common effector pathway is thought to be an increase in free cytosolic ionized calcium, which may start a cascade of effects leading to major cell permeability and capillary leak [22]. Mechanisms affecting membrane ion channels, activity of the membrane sodium–potassium pump and the production of ATP link the initial causes of rhabdomyolysis to the final effector pathway. Such mechanisms are initiated by direct damage to the membrane caused by toxins, severe exercise or compression, or failure to provide adequate ATP following ischemia or a defective oxidative metabolism. With muscle injury, large quantities of potassium, phosphate, myoglobin, CK and urate leak into the circulation. Myoglobin in the renal glomerular filtrate can precipitate and cause renal tubular obstruction, leading to renal damage [57].

## Mechanisms of ARF in rhabdomyolysis patients

It has been suggested that there are two crucial factors in the development of myoglobinuric ARF; these include hypovolemia/dehydration and aciduria. Three main mechanisms influence heme protein toxicity: renal vasoconstriction with diminished renal circulation, intraluminal cast formation and direct heme protein-induced cytotoxicity. In the absence of hypovolemia and aciduria, heme proteins have minimal nephrotoxic effects; when these conditions are present, however, heme proteins can induce renal dysfunction by a variety of mechanisms [67].

**Figure 1**




Overview of the pathophysiology of rhabdomyolysis. CK, creatine kinase.

Released heme proteins produce a synergistic effect on renal vasoconstriction initiated through hypovolemia and activation of the cytokine cascade [92] (Fig. 2). This effect possibly occurs through the scavenging of nitric oxide, which acts as a vasodilatory mediation, or through the activation of endothelin receptors consequent upon free-radical formation induced by heme protein. The enhanced renal vasoconstriction and resultant ischemia add, through depletion of tubular ATP, to the potential for damage to the renal tubular cells, already threatened by the heme-protein-induced free radicals [93].

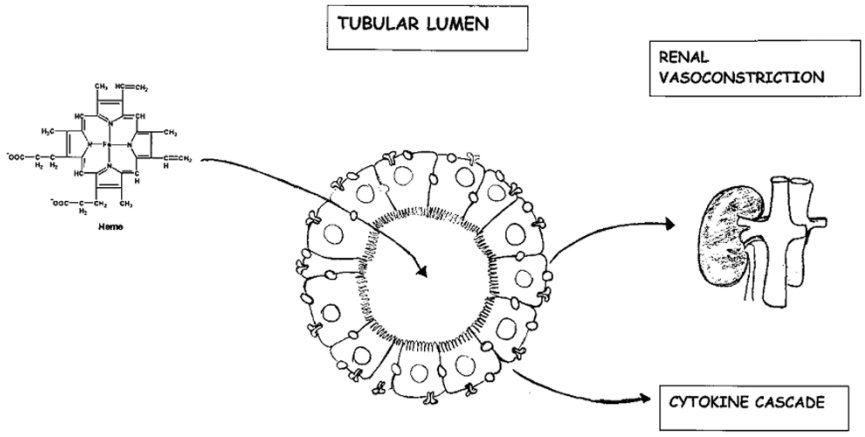
Pigmented casts are a characteristic of rhabdomyolysis-associated ARF (see Fig. 3). These are a result of the interaction of Tamm–Horsfall protein with myoglobin, which is enhanced at a low pH [53]. It has been suggested that ARF is caused by a tubular obstruction causing increased intraluminal pressure and thus opposing glomerular filtration [94].

Alternative mechanisms that have been suggested include the precipitation of heme protein providing a ready supply of material that can generate toxic free radicals [95]. The propensity for cast formation is determined by the pH, the filtered load of myoglobin and the flow through the renal tubule [96]. Heme-produced free radicals induce oxidative damage to the renal tubule [95]. Investigational work has suggested that myoglobin is central to the oxidative injury manifested as lipid peroxidation, and that this may be inhibited by an alkaline pH [97].

## Clinical manifestations

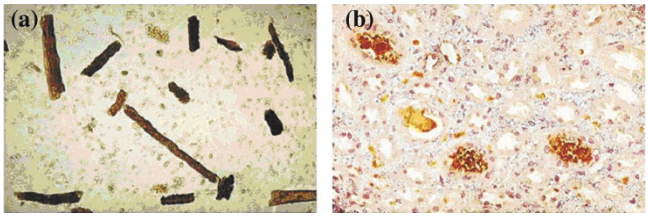
There is a wide variation in the clinical presentation of rhabdomyolysis. The 'classic' triad of symptoms includes muscle pain, weakness and dark urine [98]. The clinical manifestations can be classified as musculo-skeletal signs, general manifestations and complications. The muscle pain,

**Figure 2**




Mechanisms of heme-induced renal failure.

**Figure 3**




Pigmented casts. Analysis of urinary sediment (× 400) pigmented casts, leukocyturia, and hematuria without dysmorphic red cells. **(a)** Pigmented casts, leukocyturia, hematuria with dysmorphic cells; **(b)** with antibody against human myoglobin.

weakness, tenderness and contracture may involve specific groups of muscles or may be generalized [99]. The most frequently involved muscle groups are the calves and the lower back. The muscles can be tender and swollen, and there can be skin changes indicating pressure necrosis. However, these classic features are seen in less than 10% of the patients. Some patients experience severe excruciating pain. The calf pain may erroneously result in a work-up for deep venous thrombosis and the back pain can mimic renal colic. Similarly, involvement of the chest musculature can present with 'anginal' type chest pain. Over 50% of the patients may not complain of muscle pain or weakness [17]. The initial clinical sign of rhabdomyolysis may be the appearance of discolored urine. Urine can range from pink-tinged, to cola-colored, to dark black [100].

The general manifestations of rhabdomyolysis include malaise, fever, tachycardia, nausea and vomiting. The complications can be classified as early or late complications. The early complications include hyperkalemia, hypocalcemia, elevated liver enzymes, cardiac dysrrhythmias and cardiac arrest, while the late complications include ARF and disseminated intravascular coagulation.

Severe hyperkalemia occurs secondary to massive muscle breakdown, causing cardiac dysrrhythmias and possibly cardiac arrest. Hepatic dysfunction occurs in 25% of patients with rhabdomyolysis. Proteases released from injured muscle cause hepatic injury [101]. ARF and diffuse intravascular coagulation are late complications, developing 12–72 hours after the acute insult.

## Laboratory findings

Although the patient history and physical examination can provide clues, the diagnosis of rhabdomyolysis is confirmed by laboratory studies. CK levels are the most sensitive indicator of myocyte injury in rhabdomyolysis [47]. Normal CK enzyme levels are 45–260 U/l. CK rises in rhabdomyolysis within 12 hours of the onset of muscle injury, peaks in 1–3 days, and declines 3–5 days after the cessation of muscle injury. The peak CK level may be predictive of the development of renal failure [12]. Abnormal CK levels are commonly seen in injured intensive care unit patients, and a level of 5000 U/l or greater is related to renal failure [102]. The half-life of CK is 1.5 days and so it remains elevated longer than serum myoglobin levels [29]. Estimation of myoglobin in serum and urine is useful, particularly in the early phases of the disease [103]. Myoglobin is filtered by the kidney and appears in the urine when the plasma concentration exceeds 1.5 mg/dl [29,104]. It imparts a dark red–brown color to urine when the urine concentration exceeds 100 mg/dl. Myoglobin has a short half-life (2–3 hours) and is rapidly cleared by renal excretion and metabolism to bilirrubin [17]. Serum myoglobin levels may return to normal within 6–8 hours.

Other muscle markers can also be used. For example, carbonic anhydrase III is present in skeletal muscles but not in myocardium, and an increase in its levels is more specific for skeletal muscle injury than are CK levels [105]. Aldolase is another glycolytic pathway enzyme that is found in high concentration in skeletal muscle, the liver and the brain. While increased aldolase levels are not as specific or as sensitive for muscle disease as CK levels, increased aldolase together with an increased CK level is highly suggestive of muscle injury [106]. In addition to these enzymes, troponin I and troponin T can be helpful in diagnosing early rhabdomyolysis [107].

Both ARF and the increased release of creatinine from skeletal muscle increase the serum concentrations of urea nitrogen and creatinine. However the creatinine is elevated to a greater extent than the blood urea nitrogen, narrowing the normal 10:1 ratio of urea nitrogen to creatinine to a ratio of 6:1 or less [108]. A classic pattern of changes in serum electrolytes occurs in rhabdomyolysis. Serum levels of potassium and phosphate increase as these components are released from the cells; levels then decrease as they are excreted in urine [109]. Serum concentrations of calcium are initially decreased as calcium moves into the cells and then gradually increase. Electrolyte levels in each patient depend on the severity of the rhabdomyolysis, the stage of the illness and the therapeutic interventions that have been initiated [18]. The classic laboratory finding is an elevated serum CK of at least five times the normal value, where the creatinine kinase isoenzyme found predominately in striated muscle (CK-MM) predominates [109]. Myoglobin becomes detectable in urine and produces pigmenturia. Other findings include hyperkalemia, hypocalcemia, hyperphosphatemia and hyperuricemia along with elevated levels of other muscle enzymes like lactate dehydrogenase, aldolase, aminotransferases and carbonic anhydrase III [29].

Clotting studies are useful for detecting rhabdomyolysis – disseminated intravascular coagulation and toxicological screening should be performed if drugs are the suspected causal agent [110]. Urinalysis in patients with rhabdomyolysis will reveal the presence of protein, brown casts and uric acid crystals, and may reflect electrolyte wasting consistent with renal failure [111]. A urine dipstick is a quick way to screen for myoglobinuria, as the reagent on the dipstick that reacts with hemoglobin also reacts with myoglobin [18]. These reactants will detect hemoglobin at concentrations of 0.3 mg/l, and a similar concentration would be predicted for myoglobin [112]. Myoglobin imparts its characteristic red–brown color to urine at concentrations above 300 mg/l (see Table 3).

## Management

The treatment of rhabdomyolysis includes initial stabilization and resuscitation of the patient while concomitantly attempting to preserve renal function [113]. Retrospective analysis demonstrates that early aggressive fluid replacement with saline is beneficial in minimizing the occurrence of renal failure. The longer it takes for rehydration to be initiated, the more likely it is that renal failure will develop [22,24]. Forced diuresis, when started within 6 hours of admission, has been reported to minimize the risk of ARF [53,114].

Mannitol and bicarbonate are commonly employed following the initial resuscitation with saline [115–119]. Experimental studies suggested that mannitol may be protective due to the associated diuresis that minimizes intratubular heme pigment deposition [53,67,116]. It has also been suggested that mannitol acts as a free-radical scavenger, thereby minimizing cell injury [22]. Furthermore, mannitol reduces blood viscosity and is a renal vasodilator [120–125]. Furosemide and other loop diuretics have also been advocated for use in patients with myoglobinuric renal impairment in an attempt to initiate diuresis and convert anuric to oliguric renal failure [126–128].

Alkalinization of the urine has been suggested to minimize renal damage after rhabdomyolysis [129]. After resuscitation and restoration of normal renal perfusion, the kidneys clear a large acid load resulting in an acidic urine. It has been postulated that these patients may be unable to alkalinize their urine without the administration of bicarbonate, and this increases the risk of tubular cast development and renal injury [130–132]. Knochel and Moore, and Knottenbelt, however, have argued that large-volume infusion of crystalloid alone creates a solute diuresis sufficient to alkalinize the urine [133,134]. Furthermore, large doses of bicarbonate may worsen the degree of hypocalcemia, especially if hypovolemia is corrected [135].

While mannitol and bicarbonate are considered the standard of care in preventing ARF in patients with rhabdomyolysis [115–119], there is little clinical evidence to support the use of these agents. While randomized controlled trials are lacking, the available evidence suggests that mannitol and bicarbonate have no benefit over and above aggressive fluid resuscitation [120–123]. In a retrospective study of 24 patients Homsi and colleagues demonstrated that volume expansion with saline alone prevented progression to renal failure and that the addition of mannitol and bicarbonate had no additional benefit [119]. Using their Trauma Registry and intensive care unit database, Brown and colleagues reviewed the case records of 1771 trauma patients with increased CK levels [102]. Overall 217 patients (12%) developed renal failure, with 97 requiring dialysis. In this study, peak CK >5000 U/l was associated with an increased risk of developing renal failure. Of the 382 patients with CK >5000 U/l, 154 patients (40%) received mannitol and bicarbonate whereas 228 patients did not. There was no significant difference in the incidence of renal failure (22% versus 18%), of dialysis (7% versus 6%) or of mortality (15% versus 18%) between the two groups. Based on these data it would appear that mannitol and bicarbonate have little additional benefit over aggressive volume replacement with saline alone.

### The role of free-radical scavengers and antioxidants

The magnitude of muscle necrosis caused by ischemia-reperfusion injury has been reduced in experimental models by the administration of free-radical scavengers [136]. Many of these agents have been used in the early treatment of crush syndrome to minimize the amount of nephrotoxic material released from the muscle [137]. Pentoxyphylline is a xanthine derivative used to improve microvascular blood flow. In addition, pentoxyphylline acts to decrease neutrophil adhesion and cytokine release [138]. Vitamin E (alfa tocopherol), vitamin C (ascorbic acid), lazaroids (21-aminosteroids) and minerals such as zinc, manganese and selenium all have antioxidant activity and may have a role in the treatment of the patient with rhabdomyolysis [139,140].

### Dialysis

Despite optimal treatment, some patients will develop ARF, often with severe acidosis and hyperkalemia [141]. These patients will require renal replacement therapy to correct fluid, electrolyte and acid–base abnormalities. Daily hemodialysis or continuous hemofiltration may be required initially to remove urea and potassium that are released from damaged muscles [142]. This allows gradual removal of solutes and the slow correction of fluid overload. Normalization of potassium is the priority, because hyperkalemic cardiac arrest is a life-threatening early complication [143]. Peritoneal dialysis is inadequate to remove the large solute loads in patients with rhabdomyolysis-induced ARF, but it can offer temporary help [144]. The removal of myoglobin by plasma exchange has not demonstrated any benefit [145].

A unique management issue in rhabdomyolysis-induced ARF is the development of hypercalcemia during the recovery phase in 20–30% of patients [146,147]. To minimize this complication, the administration of calcium should be avoided during the renal failure phase, unless the patient has symptomatic hypocalcemia or severe hyperkalemia [148–150].

## Conclusions

Rhabdomyolysis is a potentially life-threatening condition that must be suspected in all patients with a history of any circumstance that can result in damage of skeletal muscle. Important clinical signs and symptoms (i.e. muscle pain, muscle tenderness and dark urine) and laboratory tests such as an elevated serum CK level and a urinalysis that reveal casts and is positive for hemoglobin, without red blood cells on microscope examination, are common. Aggressive hydration may prevent the complications of this illness. Mannitol and bicarbonate, although commonly recommended, are of unproven benefit.

## Competing interests

The author(s) declare that they have no competing interests.

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