LETTER

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Soluble programmed cell death protein-1 and programmed cell death ligand-1 in sepsis

Debasree Banerjee^{1,2*}, Sean Monaghan^{2,3}, Runping Zhao^{2,3}, Thomas Walsh², Amy Palmisciano², Gary S. Phillips⁴, Steven Opal^{1,2} and Mitchell M. Levy^{1,2}

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Immunotherapy targeting the programmed cell death protein-1 (PD-1)–programmed cell death ligand-1 (PDL-1) axis in sepsis is poised for clinical trials, although optimal inclusion criteria and predictors of response are not well characterized.

We evaluated the kinetics of soluble (s)PD-1 and sPD-L1 in 30 septic intensive care unit (ICU) patients and 30 nonseptic ICU patients (Table 1). sPD-1 and sPD-L1 were significantly elevated among the septic cohort compared with the nonseptic ICU patients at enrollment (17.7 pg/ml vs. 4.5 pg/ml, p =0.002; and 29.9 pg/ml vs. 11.3 pg/ml, p = 0.02; respectively) and were associated with sepsis (Fig. 1). Higher sPD-L1 on day 3 was associated with mortality among septic patients (16.7 pg/ml vs. 3.0 pg/ml, p = 0.054) and also in the total ICU cohort (14.9 pg/ ml vs. 2.7 pg/ml, p = 0.026). Soluble PD-L1 regressed on interleukin (IL)-6 and interferon (IFN)y levels were significantly associated in the total ICU cohort and septic patients, possibly pointing to upstream triggers for post-transcriptional modifications. Tumor necrosis factor (TNF) α regressed on sPD-1 and sPD-L1 was significant in all populations including septic survivors, revealing possible downstream effects of sPD-1 and sPD-L1. Initial sPD-1 levels correlated with a drop in lymphocyte count to $< 1 \times$ $10^{9}/L$ (area under the receiver operating characteristic (ROC) curve 0.72, p = 0.006) and to $< 0.6 \times 10^9/L$ (area under the ROC curve 0.68, p = 0.02). sPD-L1

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also correlated with lymphocyte count drop to $< 1 \times 10^9$ /L during the hospital stay. The correlation between the two immune checkpoint molecules, sPD-1 and sPD-L1, was also significant on enrollment, and at days 1 and 3 (p < 0.001, p < 0.001, p = 0.004, respectively; Fig. 2).

sPD-1 and sPD-L1 are easily sampled, making them advantageous biomarkers in sepsis. A recent study demonstrated elevated sPD-1 among patients with infected pancreatitis [1]. sPD-1 may serve as an indicator of severity of sepsis among emergency room patients [2]. Lange et al. [3] reported that sPD-1 levels did not differ significantly between septic and nonseptic critically ill patients and had no association with outcome among septic patients. Our results may stem from sampling a different population to Lange and colleagues. Our controls, while critically ill, had lower severity of illness and mortality. Additionally, we excluded patients with immunocompromise, malignancy, and organ transplantation due to possible iatrogenic skewing of sPD-1 and sPD-L1. Approximately half of the control group in Lange et al. developed infections; thus, observations comparing sepsis versus nonseptic groups were limited to initial measurement only.

sPD-1 and sPD-L1 are point-of-care tests that might eventually guide personalized medicine in sepsis. These soluble immune checkpoints can risk-stratify patients for immunotherapy in sepsis and may potentially serve as targets themselves.

^{*} Correspondence: banerjed19@gmail.com

¹Departments of Medicine, Warren Alpert Medical School of Brown

University, Providence, RI, USA

²Lifespan Hospital System, Providence, RI, USA

1.0

0.8

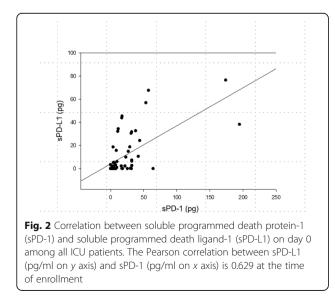
0.6

0.4

0.2

Sensitivity

PD-L1, A = 0.77 0.0 0.0 0.2 0.4 0.6 0.8 1.0 1 - Specificity Fig. 1 The area under the ROC curve for the discrimination of sepsis by soluble programmed death protein-1 (sPD-1) and soluble programmed death ligand-1 (sPD-L1). These are the day 0 area under the ROC curves for Sequential Organ Function Assessment (SOFA) score, sPD-1 (pg/ml), and sPD-L1 (pg/ml) for the discrimination of patients who have sepsis. Soluble PD-L1 outperforms sPD-1 for discrimination of sepsis



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Age (years), median (IQR)	63.3 (49.3–74.1)	58.6 (52.8–64.6)
Male, n (%)	11 (36)	19 (63)
White, <i>n</i> (%)	23 (76)	25 (83)
Past medical history		
COPD/asthma/fibrosis, n (%)	11 (36)	2 (7)
CAD/MI, CHF, AF, n (%)	16 (53)	7 (23)
Diabetes mellitus, n (%)	5 (17)	10 (33)
Malignancy, n (%)	8 (27)	6 (20)
CKD/ESRD, n (%)	5 (17)	1 (3)
Cirrhosis, n (%)	0 (0)	4 (13)
Connective tissue disease, n (%)	3 (10)	0
Arthritis, n (%)	3 (10)	3 (10)
Clinical assessment		
WBC (×10 ⁻⁹ /L), median (IQR)	12.8 (6.9–19.1)	9.1 (7.6–10.9)
SOFA score, median (IQR)	7 (5–9)	2 (1-4)
Shock, <i>n</i> (%)	24 (80)	2 (6)
Site of infection in septic patients (n)		
Pneumonia	13	
Genitourinary tract infection	11	
Abdominal infection	2	
Meningitis	1	
Multiple sites of infection	1	
Bacteremia	9	
Unknown	1	
Organism of infection if known in septic patients (<i>n</i>)		
Escherichia coli not extended-spectrum β -lactamase producer	4	

Table 1 Patient characteristics

Variables

SOFA Score, A = 0.87

PD-1, A = 0.74

E: Escherichia coli extended-spectrum β-2 lactamase producer Enterobacter 1 Enterococcus faecalis 1 Acinetobacter baumannii 1 Pseudomonas aeruginosa 1 Methicillin-resistant Staphylococcus aureus 1 Methicillin-sensitive Staphylococcus aureus 1 Candida albicans 1 Haemophilus influenzae 1 Bacillus species not anthracis 2

AF atrial fibrillation, COPD chronic obstructive pulmonary disease,

Klebsiella pneumoniae

CAD coronary artery disease, CHF congestive heart Failure, CKD chronic kidney disease, ESRD end-stage renal disease, IQR interquartile range, MI myocardial infarction, SOFA sequential organ function assessment, WBC white blood cell

1

Control subjects

(n = 30)

Septic patients

(n = 30)

Abbreviations

ICU: Intensive care unit; IFN: Interferon; IL: Interleukin; PD-1: Programmed cell death protein-1; PD-L1: Programmed cell death ligand-1; ROC: Receiver operator characteristic; TNF: Tumor necrosis factor

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MML and SO constructed the biorepository for this study. DB contributed to the IRB for this study and handling of specimens. TW enrolled patients and collected biosamples and AP supervised biobank maintenance. RZ performed the multiplex analysis. GSP performed the statistical analysis for this project, GSP and SM created the figures presented. DB drafted and edited the manuscript, while SM, SO, GSP, and MML edited and finalized the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

This study was approved by the Rhode Island Hospital IRB 4159-14. All participants consented prior to enrollment.

Competing interests

The authors declare that they have no competing interests.

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Author details

¹Departments of Medicine, Warren Alpert Medical School of Brown University, Providence, RI, USA. ²Lifespan Hospital System, Providence, RI, USA. ³Department of Surgery, Warren Alpert Medical School of Brown University, Providence, RI, USA. ⁴Department of Biomedical Informatics, Center for Biostatistics, Ohio State University, Columbus, OH, USA.

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