

Primary research

## Lack of agreement between tonometric and gastric juice partial carbon dioxide tension

Arnaldo Dubin, Julio Badie, Sofía Fernandez, Elisa Estenssoro, Héctor Canales, Guillermo Bordoli and Fernando Pálizas  
Clínica Bazterrica, Capital Federal, Argentina

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### Statement of findings

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Our goal was to compare measurement of tonometered saline and gastric juice partial carbon dioxide tension ( $PCO_2$ ). In this prospective observational study, 112 pairs of measurements were simultaneously obtained under various hemodynamic conditions, in 15 critical care patients. Linear regression analysis showed a significant correlation between the two methods of measuring  $PCO_2$  ( $r^2 = 0.43$ ;  $P < 0.0001$ ). However, gastric juice  $PCO_2$  was systematically higher (mean difference 51 mmHg). The 95% limits of agreement were 315 mmHg and the dispersion increased as the values of  $PCO_2$  increased. Tonometric and gastric juice  $PCO_2$  cannot be used interchangeably. Gastric juice  $PCO_2$  measurement should be interpreted with caution.

**Keywords:** gastric tonometry, intramucosal partial carbon dioxide tension, intramucosal pH

### Synopsis

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**Introduction:** In recent years there has been growing interest in tonometric estimation of gastric intramucosal pH (pHi). More recently, attention has focused on the gradient between intraluminal and arterial  $PCO_2$ . pHi appears to be a useful diagnostic and prognostic tool in critically ill patients, and may also be used as a therapeutic guide. However, intraluminal  $PCO_2$  is the parameter measured to calculate pHi, and it is assumed as equivalent to the  $PCO_2$  of the upper layers of the gastric mucosa.

Direct measurement of  $PCO_2$  in gastric juice might offer advantages over tonometry. Tonometer costs could be saved, and equilibration time would no longer be necessary. Additionally, preanalytic factors that account for poor reproducibility, such as inadequate volume of saline in the tonometer, errors in the dwell time of the sample or in the technique used to aspirate saline, mixing of the sample with tonometer dead space and delay in analysis, could be prevented. Nevertheless, to our knowledge few experimental or

clinical studies have examined  $PCO_2$  in gastric juice. Moreover, no comparison with simultaneous tonometric samples has been performed. Our goal was to compare simultaneous measurement of  $PCO_2$  in gastric juice and in saline samples from a tonometer. Data from the present study show that gastric juice  $PCO_2$  is systematically higher. Furthermore, differences widen at high  $PCO_2$  values, and data dispersion becomes even more striking. Therefore, tonometric  $PCO_2$  and gastric juice  $PCO_2$  are not interchangeable.

**Patients and methods:** The present study was approved by the local ethics committee, and informed consent was obtained from the next of kin of each patient.

We studied 15 consecutive mechanically ventilated patients from a medical/surgical intensive care unit, in whom tonometric monitoring was indicated by attending physicians. All patients were receiving 50 mg intravenous ranitidine every 8 h. Gastric tonometers were filled with saline, which was extracted after 90 min of equilibration time. At the same time, gastric juice was

Table 1

Clinical characteristics and first value of arterial, tonometer and gastric juice Pco<sub>2</sub>

Patient	Sex	Age (years)	Diagnosis	Inotropes (µg/kg per min)	Outcome	Pco <sub>2</sub> (mmHg)		
						Arterial	Tonometric	Gastric juice
1	Female	53	Stroke, ARDS	Dopamine 32	Survival	30	48	165
2	Female	73	Intestinal obstruction, septic shock	Dopamine 40	Death	26	44	92
3	Male	37	Multiple trauma		Survival	21	28	41
4	Male	56	Multiple trauma		Survival	39	42	49
5	Female	64	Acute pancreatitis, shock, ARDS	Dopamine 18	Death	30	34	80
6	Male	17	Multiple trauma		Survival	43	60	60
7	Female	18	Fat liver of pregnancy		Survival	30	40	44
8	Male	73	Necrotizing cellulitis, septic shock, ARDS	Epinephrine 1.2	Death	28	33	31
9	Male	64	Multiple trauma, pneumonia, ARDS		Death	36	41	57
10	Male	65	Lung cancer postoperatively, ARDS		Death	35	51	242
11	Male	65	Lung cancer postoperatively, ARDS	Dopamine 20	Death	36	30	125
12	Female	22	Neutropenia, septic shock, ARDS	Epinephrine 0.8	Death	50	69	81
13	Male	83	Perioperative shock	Dopamine 25	Survival	23	28	34
14	Male	52	Ventilator-associated pneumonia		Survival	43	43	126
15	Male	56	Colangitis, septic shock	Dopamine 36	Survival	38	44	92

ARDS, acute respiratory distress syndrome.

anaerobically extracted from the aspiration port of the tonometer. The initial 20 ml was discarded. Pco<sub>2</sub> in both samples was measured using a blood gas analyzer (AVL 945; AVL List GMBH, Gratz, Austria). These measurements were taken at various time points in each patient, and under various haemodynamic and oxygen transport conditions. All measurements were performed with the patient fasted. Correlation between the two measurements was examined using the Bland–Altman technique.

We also performed an *in vitro* study to quantify the precision and bias for the AVL 945. For this purpose, a stable Pco<sub>2</sub> in saline solution was achieved by bubbling 5% carbon dioxide calibration gas.

**Results:** We performed 112 pairs of measurements in 15 patients. Table 1 shows clinical data and the first values of arterial, tonometered and gastric juice Pco<sub>2</sub> for each patient. Regression analysis demonstrated a significant correlation between both methods of measuring Pco<sub>2</sub> ( $r^2 = 0.43$ ; gastric juice Pco<sub>2</sub> =  $-28.79 + [2.55 \times \text{tonometric Pco}_2]$ ;  $P < 0.0001$ ; Fig. 1). However, the bias calculated as the mean difference of gastric juice and tonometric Pco<sub>2</sub> was 51 mmHg. The 95% limits of agreement were 315 mmHg (Fig. 2). For mean Pco<sub>2</sub> values lesser than 100 mmHg, the bias and the 95% limits of agreement were 19 and 102 mmHg, respectively. As mean Pco<sub>2</sub> increased, the scattering of differences widened ( $r^2 = 0.71$ ;  $P < 0.0001$ ).

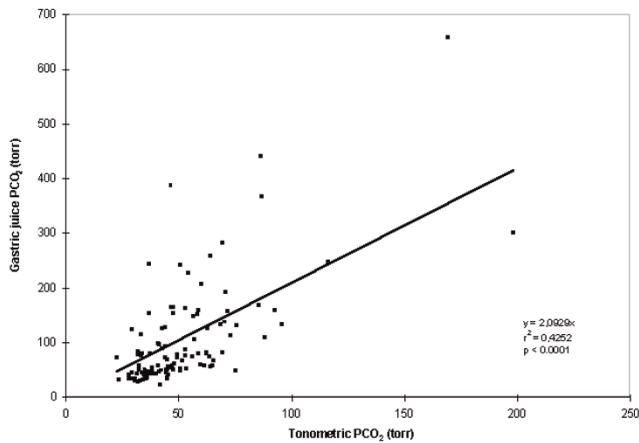
In an effort to prevent the bias related to multiple measurements per patient, we performed Bland–Altman analysis with the first measurement of each patient. After this the results remained similar (bias 55 mmHg, 95% limits of agreement 216 mmHg).

The AVL 945 blood gas analyzer showed a negative bias of 0.97 mmHg and a precision of 2.13 mmHg. This bias was considered negligible, so no further correction was made to saline tonometric values.

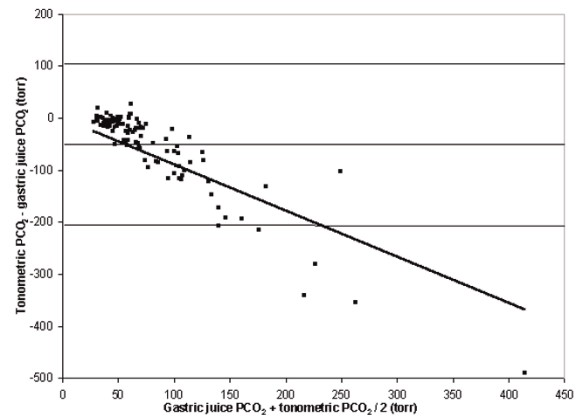
**Discussion:** The results of the present study show that tonometric Pco<sub>2</sub> and gastric juice Pco<sub>2</sub> are not interchangeable. Gastric juice Pco<sub>2</sub> is systematically higher. At high Pco<sub>2</sub> values the differences widen, and data dispersion becomes even more marked.

There is no clear cause for these observations. A possible explanation might be that tonometric Pco<sub>2</sub> is generated over a time interval, whereas gastric juice Pco<sub>2</sub> might reflect rapid changes in mucosal metabolism. Different equilibrium time could also account for data dispersion, but not for the positive bias for gastric juice. Rapid changes should occur in both directions.

Another potential confounding factor is the ability of blood gas analyzers to measure Pco<sub>2</sub> in gastric juice. Measurement of Pco<sub>2</sub> in 0.9% saline is an important source of error in the estimation of pH<sub>i</sub>. Variation in Pco<sub>2</sub> values may occur with different Pco<sub>2</sub> equilibration solutions. For example, bias is –66.5% when the Nova Stat Profile 7 blood gas analyzer (Nova

**Figure 1**

Correlation between gastric juice and tonometric  $\text{PCO}_2$ . We performed 112 pairs of measurements of gastric juice and tonometric  $\text{PCO}_2$  in 15 critical care patients under different haemodynamic and oxygen transport conditions. The linear regression coefficient is significant. However, the slope value indicates systematic overestimation of gastric juice  $\text{PCO}_2$  in relation to saline  $\text{PCO}_2$ .

**Figure 2**

Bland-Altman analysis of the differences between gastric juice and tonometric  $\text{PCO}_2$ . The bias calculated as the mean difference of gastric juice and tonometric  $\text{PCO}_2$  was 51 mmHg. The 95% limits of agreement were 315 mmHg. The bias and the scattering of differences widened as  $\text{PCO}_2$  increased.

Biomedical, Waltham, MA, USA) measures concentration of 1.95% of  $\text{CO}_2$  equilibrated in normal saline. However, bias changes to +45.4% when 1.95%  $\text{CO}_2$  is equilibrated in human albumin solution 4.5%.

It would not be surprising if gastric juice components such as proteins, mucopolisaccharides and others interfere with  $\text{CO}_2$  solubility and its subsequent measurement by blood gas analyzers. In this way, intersubject and intrasubject variation in gastric juice composition could also account for data dispersion. Fiddian-Green *et al* [1] measured  $\text{PCO}_2$  in gastric contents of anaesthetized dogs. They isolated the stomach from the oesophagus and the duodenum with ligatures, and washed it through a catheter with saline. Then, they instilled 250 ml 0.9% saline and took samples to measure  $\text{PCO}_2$  and to

estimate  $\text{pHi}$ . Simultaneously, mucosa  $\text{pHi}$  was recorded with a microglass probe. They found a statistically significant correlation between both methods. However, data dispersion in the graph was considerable.

We were able to exclude analyzer underestimation of  $\text{PCO}_2$  in saline as the cause for the present results. *In vitro* performance of the AVL 945 in blood was good. It showed a negative bias less than 1 mmHg and a precision of about 2 mmHg.

We cannot infer from the present data the technique that should be the gold standard for measuring  $\text{PCO}_2$  in gastric mucosa. However, the studies that have established the normal values for  $\text{pHi}$ , prognostic changes and its uses as a therapeutic index have been performed with tonometry. Hence, more data are needed for the routine measurement of  $\text{PCO}_2$  in gastric juice.

## Full article

### Introduction

In recent years there has been growing interest in tonometric estimation of gastric  $\text{pHi}$ . More recently attention has focused on the gradient between intraluminal and arterial  $\text{PCO}_2$ .  $\text{pHi}$  appears to be a useful diagnostic and prognostic tool in critically ill patients, and may also be used as a therapeutic guide [2,3]. However, intraluminal  $\text{PCO}_2$  is the parameter measured to calculate  $\text{pHi}$ , and it is assumed as equivalent to the  $\text{PCO}_2$  of the upper layers of the gastric mucosa [4].

Direct measurement of  $\text{PCO}_2$  in gastric juice might offer additional advantages over tonometry. First, tonometer costs could be saved. Second, equilibration time would no longer be necessary. Finally, preanalytic factors that account for poor tonometric reproducibility, such as inadequate volume of saline in the tonometer, errors in timing the dwell time of the sample or in the technique used to aspirate the tonometered saline sample, mixing the sample with air from the tonometer dead space, and delays in specimen analysis, might be prevented [5].

Nevertheless, to our knowledge, very few studies, either experimental or clinical, have examined  $\text{PCO}_2$  in gastric juice [1,6,7]. Moreover, no comparison with tonometric samples obtained at the same time has been done. Our goal was to compare  $\text{PCO}_2$  obtained simultaneously in gastric juice and in saline samples from tonometers. The results of the present study show that gastric juice  $\text{PCO}_2$  is systematically higher, and that for high  $\text{PCO}_2$  values this difference widens and data dispersion is even more marked. Therefore, tonometric  $\text{PCO}_2$  and gastric juice  $\text{PCO}_2$  are not interchangeable.

### Patients and methods

The present study was approved by the local ethics committee and informed consent was obtained from the next of kin of each patient.

We consecutively studied 15 mechanically ventilated patients from a medical/surgical intensive care unit, in whom tonometric monitoring was indicated by attending physicians. All patients were receiving 50 mg intravenous ranitidine every 8 h. Gastric tonometers were filled with saline and, after 90 min for equilibration, saline samples were collected, as has previously been described [2]. At the same time, gastric juice was anaerobically extracted from the aspiration port of the tonometer. Initial 20 ml were discarded. A blood gas analyzer (AVL 945) was used to measure  $\text{PCO}_2$  in both samples. These measurements were taken at various time points in each patient, and under various haemodynamic and oxygen transport conditions. All measurements were performed with the patient fasted. Correlation between both of measurements was examined using the Bland–Altman technique [8].

We also performed an *in vitro* study to quantify the precision and bias of the AVL 945. For this purpose, a stable  $\text{PCO}_2$  in saline solution was achieved by bubbling 5% carbon dioxide calibration gas. Bias (mean difference from expected  $\text{PCO}_2$ ) and precision (standard deviation of the bias) of  $\text{PCO}_2$  measured in saline was determined by comparison of measured  $\text{PCO}_2$  with expected  $\text{PCO}_2$ . The latter was calculated from the carbon dioxide content of the calibration gas and from barometric pressure, according to gas laws. Measurements were repeated six times.

### Results

We performed 112 pairs of measurements in 15 patients. Table 1 shows clinical data and the first values of arterial, tonometer and gastric juice  $\text{PCO}_2$  taken in each patient. Regression analysis demonstrated a significant correlation between both methods of measuring  $\text{PCO}_2$  ( $r^2 = 0.43$ ; gastric juice  $\text{PCO}_2 = -28.79 + [2.55 \times \text{tonometric } \text{PCO}_2]$ ;  $P < 0.0001$ ; Fig. 1). However, bias, calculated as the mean difference between gastric juice and tonometric  $\text{PCO}_2$ , was 51 mmHg. The 95% limits of agreement were 315 mmHg (Fig. 2). When mean  $\text{PCO}_2$  values were less than 100 mmHg,

bias and 95% limits of agreement were 19 and 102 mmHg, respectively. The scattering of differences widened as  $\text{PCO}_2$  increased ( $r^2 = 0.71$ ;  $P < 0.0001$ ).

The two types of measurements are clearly correlated. However, they cannot be considered as interchangeable, because gastric juice  $\text{PCO}_2$  was always higher and the 95% limits of agreement were clinically significant.

In an effort to prevent bias related to multiple measurements per patient we used another approach, taking into account the initial measurement of each patient. Despite this, results continued to be similar (bias 55 mmHg, 95% limits of agreement 216 mmHg).

The AVL 945 blood gas analyzer showed a negative bias of 0.97 mmHg and a precision of 2.13 mmHg. This was considered negligible, so no further correction was done to tonometric values.

### Discussion

The present data show that tonometric  $\text{PCO}_2$  and gastric juice  $\text{PCO}_2$  should not be considered interchangeable, because gastric juice  $\text{PCO}_2$  was systematically higher. With high  $\text{PCO}_2$  values, this difference widened, and data dispersion became even more marked.

There is no clear cause for these observations. It can be argued that tonometric  $\text{PCO}_2$  is a value that is measured over a predetermined period, whereas gastric juice  $\text{PCO}_2$  may generate data on minute-to-minute changes in mucosal metabolism. This different equilibrium time could account for data dispersion. However, the positive bias for gastric juice is harder to interpret, because such rapid changes should appear in both directions.

Another potential confounding factor is the ability of blood gas analyzers to measure  $\text{PCO}_2$  in gastric juice. Measurement of  $\text{PCO}_2$  in saline is an important source of error in the estimation of  $\text{pHi}$ . The error lies both in the kind of blood gas analyzer used and in the  $\text{PCO}_2$  value itself [9,10]. Different solutions might modify  $\text{PCO}_2$  measurement. For example, bias is  $-66.5\%$  when the Nova Stat Profile 7 blood gas analyzer measures concentration of 1.95% of  $\text{CO}_2$  equilibrated in normal saline. However, bias changes to  $+45.4\%$  when 1.95%  $\text{CO}_2$  is equilibrated in human albumin solution 4.5% [9].

It would not be surprising if different gastric juice buffers, such as proteins, mucopolisaccharides and others, interfere with  $\text{CO}_2$  solubility and its subsequent measurement. In this way, intersubject and intrasubject variation of gastric juice composition could also account for data dispersion. Therefore, analytic issues related to the various constituents of gastric juice could have added to the observed differences.

Fiddian-Green *et al* [1] measured  $\text{PCO}_2$  in gastric content of anaesthetised dogs. They isolated the stomach from the oesophagus and the duodenum with ligatures, and washed it through a catheter with saline. Then they instilled 250 ml saline and intermittently took samples to measure  $\text{PCO}_2$  and estimate  $\text{pHi}$ , which was compared with simultaneous direct mucosa  $\text{pHi}$  measurement performed using a microglass probe. They found a statistically significant correlation between both methods, but there was considerable data dispersion in the graph. However, in that study, as well as in others in which  $\text{PCO}_2$  was measured without a tonometer [6,7], some kind of saline lavage was used. On the other hand, we directly measured  $\text{PCO}_2$  in gastric juice, which could produce marked differences in the results.

Differences between tonometric  $\text{PCO}_2$  and gastric juice  $\text{PCO}_2$  could be due to blood gas analyzers that are calibrated for blood gas analysis, and could systematically underestimate  $\text{PCO}_2$  in saline [9,10]. Nevertheless, this explanation is not supported by the present *in vitro* results. The performance of the AVL 945 was fairly good. It showed a minor negative bias of less than 1 mmHg and a precision of approximately 2 mmHg. Thus, it appears to be a suitable device for saline tonometry.

We ascribe the high  $\text{PCO}_2$  values obtained in the present study to shock and subsequent gut hypoperfusion, as can be deduced from the clinical characteristics of our patients. Also, in the presence of hydrochloric acid, duodenal or gastric bicarbonate buffering could produce important amounts of carbon dioxide [11]. Although we cannot exclude spurious generation of carbon dioxide, measurements were performed in the fasted state, the position of the tonometer was checked by X-ray, and ranitidine was administered in each patient. If  $\text{PCO}_2$  in gastric juice and in the tonometer were interchangeable, their values would be similar, regardless of  $\text{CO}_2$  source or absolute value. However, gastric juice  $\text{PCO}_2$  and tonometric  $\text{PCO}_2$  were quite different. We believe that this is the novel point in this study.

Gastric juice  $\text{PCO}_2$  has been used in some clinical studies [6,7,12]. Mohsenifar *et al* [12] advocated its advantages over tonometry. However, the present results suggest that the two techniques are not interchangeable. From the present data we cannot infer which should be the gold standard technique for measurement of gastric mucosa  $\text{PCO}_2$ . Nevertheless, knowledge regarding  $\text{pHi}$  has evolved from gastric tonometry; studies regarding normal values, prognostic changes and its uses as a therapeutic guide were performed using tonometry. Hence, until more and clearer data are reported,  $\text{PCO}_2$  measurement in gastric juice should be considered with caution.

## References

1. Fiddian-Green RG, Pittenger G, Whitehouse WM: **Back-diffusion of  $\text{PCO}_2$  and its influence on the intramural pH in gastric mucosa.** *J Surg Res* 1982, **33**:39–48.
2. Doglio GR, Pusajo JF, Egurrola MA, *et al*: **Gastric mucosal pH as a prognostic index of mortality in critically ill patients.** *Crit Care Med* 1991, **19**:1037–1040.
3. Gutierrez G, Pálizas F, Doglio G, *et al*: **Gastric intramucosal pH as a therapeutic index of tissue oxygenation in critically ill patients.** *Lancet* 1992, **339**:195–199.
4. Antonsson JB, Boyle CC, Kruit KL, *et al*: **Validation of tonometric measurement of gut intramural pH during endotoxemia and mesenteric occlusion in pigs.** *Am J Physiol* 1990, **259**:G519–G523.
5. Oud L, Kruse JA: **Poor in vivo reproducibility of gastric intramucosal pH determined by saline-filled balloon tonometry.** *J Crit Care* 1996, **11**:144–150.
6. Gys T, Hubens A, Neels H, *et al*: **Prognostic value of gastric intramucosal pH in surgical intensive care patients.** *Crit Care Med* 1988, **16**:1222–1224.
7. Mohsenifar Z, Hay A, Hay J, *et al*: **Gastric intramucosal pH as a predictor of success or failure in weaning patients from mechanical ventilation.** *Ann Intern Med* 1993, **119**:794–798.
8. Bland JM, Altman DG: **Statistical methods for assessing agreement between two methods of clinical measurement.** *Lancet* 1986, **i**:307–310.
9. Riddington D, Venkatesh KB, Clutton-Brock T, *et al*: **Measuring carbon dioxide tension in saline and alternative solutions: quantification of bias and precision in two blood gas analyzers.** *Crit Care Med* 1994, **22**:96–100.
10. Takala J, Parviainen I, Silohao M, *et al*: **Saline  $\text{PCO}_2$  is an important source of error in the assessment of gastric intramucosal pH.** *Crit Care Med* 1994, **22**:1877–1879.
11. Stevens MH, Thirlbly RC, Feldman M: **Mechanism for high  $\text{PCO}_2$  in gastric juice: roles of bicarbonate secretion and  $\text{CO}_2$  diffusion.** *Am J Physiol* 1987, **253**:G527–G530.
12. Mohsenifar Z, Collier J, Koerner SK: **Gastric intramural pH in mechanically ventilated patients.** *Thorax* 1996, **51**:606–610.

**Authors' affiliation:** Servicio de Terapia Intensiva, Clínica Bazterrica, Capital Federal, Argentina

**Correspondence:** Arnaldo Dubin MD, Servicio de Terapia Intensiva, Clínica Bazterrica, Juncal 3002, (1425) Capital Federal, Argentina.  
E-mail: adee@infovia.com.ar